

# 4D Analysis of Lipid Nanoparticles (LNP) using Elute-timsTOF Pro 2 with VIP-HESI source

Beixi Wang,<sup>1</sup> Viola Jeck,<sup>2</sup> Matthew Lewis,<sup>2</sup> Xuejun Peng,<sup>1</sup> Surendar Tadi,<sup>3</sup> Erica Forsberg<sup>1</sup>

<sup>1</sup>Bruker Scientific LLC, 101 Daggett Drive, San Jose, CA 95134, USA

<sup>2</sup>Bruker Daltonik GmbH, Fahrenheitstraße 4, 28359 Bremen, Germany

<sup>3</sup>Bruker Scientific LLC, 40 Manning Road, Billerica, MA 01821, USA

## Introduction

Lipid nanoparticles (LNPs) have emerged from the recent COVID-19 pandemic as effective delivery vehicles for mRNA vaccines,<sup>1,2</sup> and pharmaceutical formulations in nucleic acid-based drugs. LNPs are typically composed of four critical lipid species components:

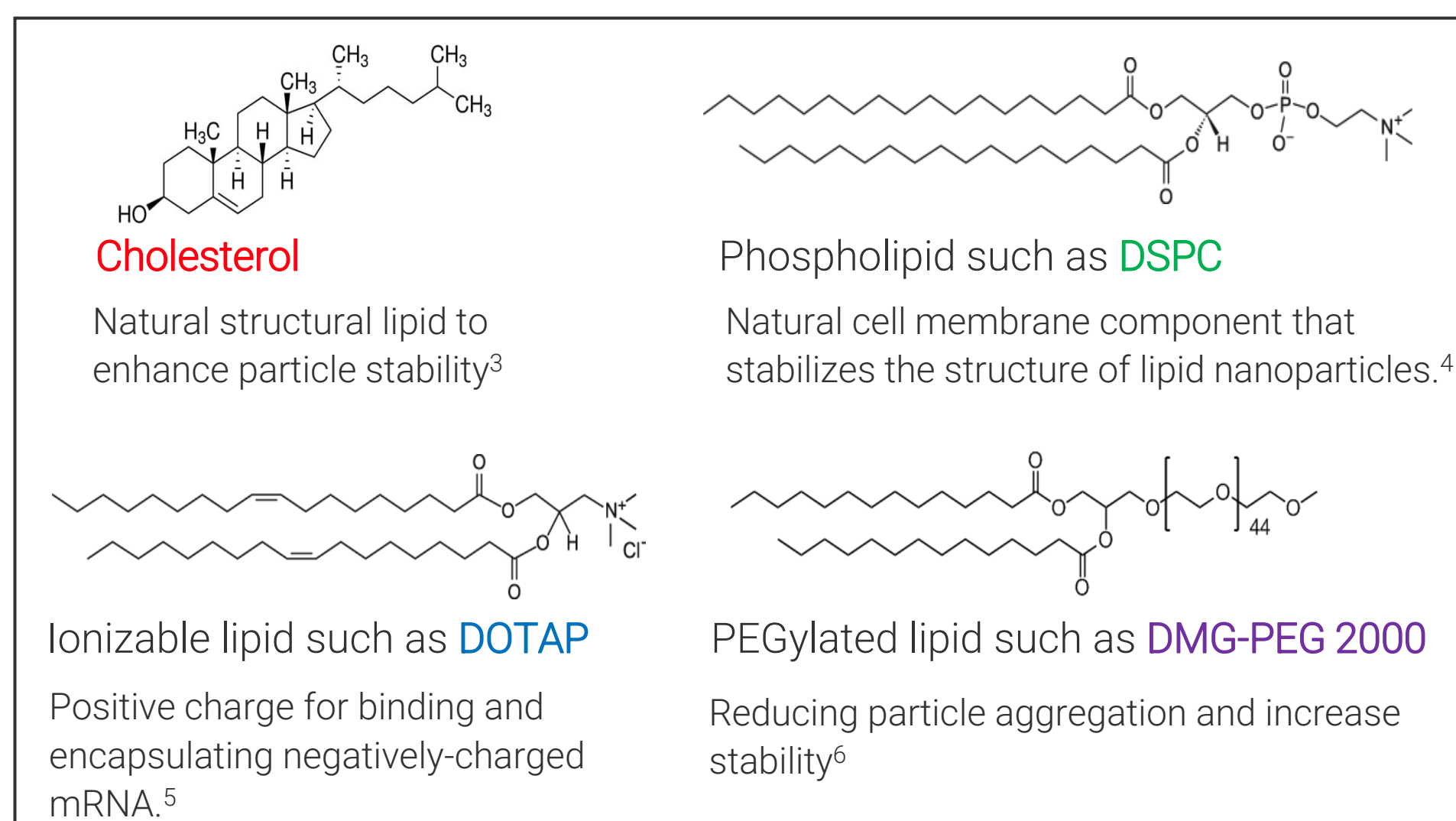


Figure 1. Structure of four components of lipid nanoparticles (LNP).

As LNPs become more commonly considered for use by drug and vaccine manufacturers, the research interest in their composition, stability, and biological interactions is driving a need for bioanalytical solutions capable of in-depth sample characterization. Bruker's 4D-Lipidomics™ solution leverages powerful multidimensional separation by combining ultra-high performance liquid chromatography (UHPLC) with trapped ion mobility spectrometry (TIMS) and high resolution mass spectrometry (HRMS) in a single hyphenated system capable of separating and describing sample components by their retention time, collision cross section (CCS), and mass-to-charge ( $m/z$ ) measurements, respectively. Here, four components of LNPs are analyzed using the Elute-timsTOF Pro 2 system with CCS-capability via a VIP-HESI source. Analysis of LNP lipid species in a complex biological matrix is also investigated.

## Methods

Working solution chemical standards of DSPC, DOTAP, DMG-PEG-2000 (Avanti Polar Lipids) and cholesterol (Sigma-Aldrich) with concentrations ranging from 1 ppb – 100 ppm of the resulting LNP component mixture were analyzed using Elute-timsTOF Pro 2 with both ESI and VIP-HESI sources.

Lipids were extracted from NIST SRM 1950 plasma (Sigma-Aldrich) by MTBE. One aliquot was analyzed as is and the other aliquot was spiked with LNP component mixture (5 ppm cholesterol, 1 ppm DMG-PEG 2000, and 50 ppb DOTAP/DSPC).

LC System	Elute	MS System	timsTOF Pro2
Column	YMC-Triart C18, 100x2.1 mm, 1.9 $\mu$ m, P/N TA12SP9-10Q1PT	Method	default 4D-Lipidomics_pos
Mobile Phase A	600/390/10 MeCN/Water/1M ammonium formate in 0.1% formic acid	Parameter	Value
Mobile Phase B	900/90/10 IPA/MeCN/1M ammonium formate in 0.1% formic acid	End Plate Offset	500 V
Gradient	Time	%A	%B
	0.0	50	50
	0.5	47	53
	4.0	45	55
Flow Rate	7.0	35	65
	7.5	20	80
	10.0	1	99
	13.0	1	99
	13.1	50	50
Column Temp	55 °C	ESI Source Parameter	Capillary Voltage
Sample Temp	4 °C	End Plate Offset	500 V
Injection Volume	2 $\mu$ L	Parameter	Value
		End Plate Offset	500 V
		Capillary Voltage	4000 V
		Nebulizer	2.2 Bar
		Dry Gas	10.0 L/min
		Dry Temp	220 °C

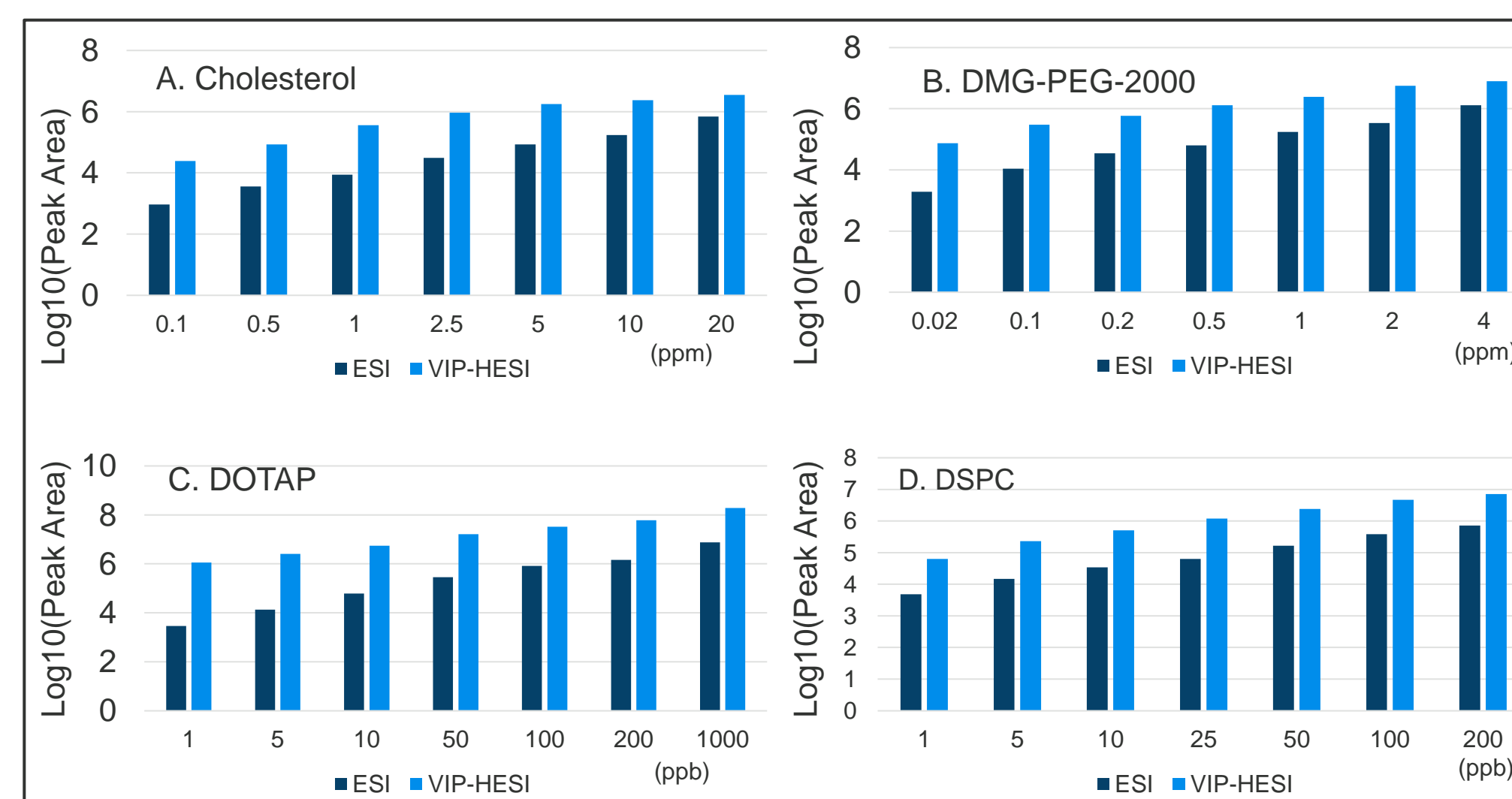


Figure 2. Log10 of peak area of representative EIC peaks from the four lipid species comparing standard ESI source vs. VIP-HESI source for cholesterol at  $m/z$  369.3516, DMG-PEG-2000 at  $m/z$  854.2496, DOTAP at  $m/z$  662.6087, and DSPC at  $m/z$  790.6320.

## Results

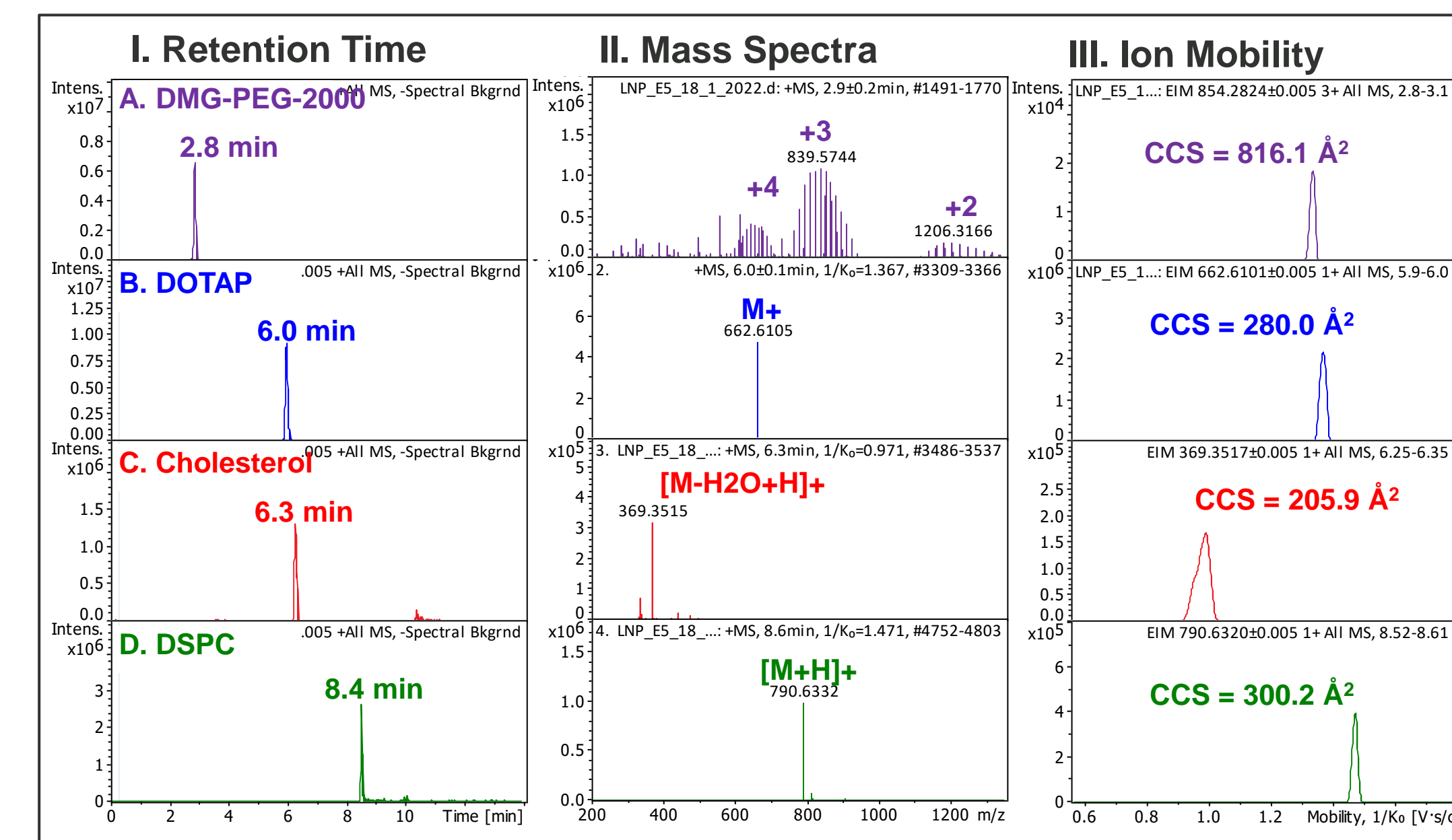


Figure 3. Results of (I) extracted ion chromatogram, (II) extracted mass spectra, and (III) extracted ion mobilograms for the analysis of a mixture of four LNP components: (A) DMG-PEG-2000 ( $m/z$  854.2496 for +3 of 44 repeating PEG unit), (B) DOTAP for the cation ( $m/z$  662.6087), (C) Cholesterol with water loss ( $m/z$  369.3516), and (D) DSPC ( $m/z$  790.6320)

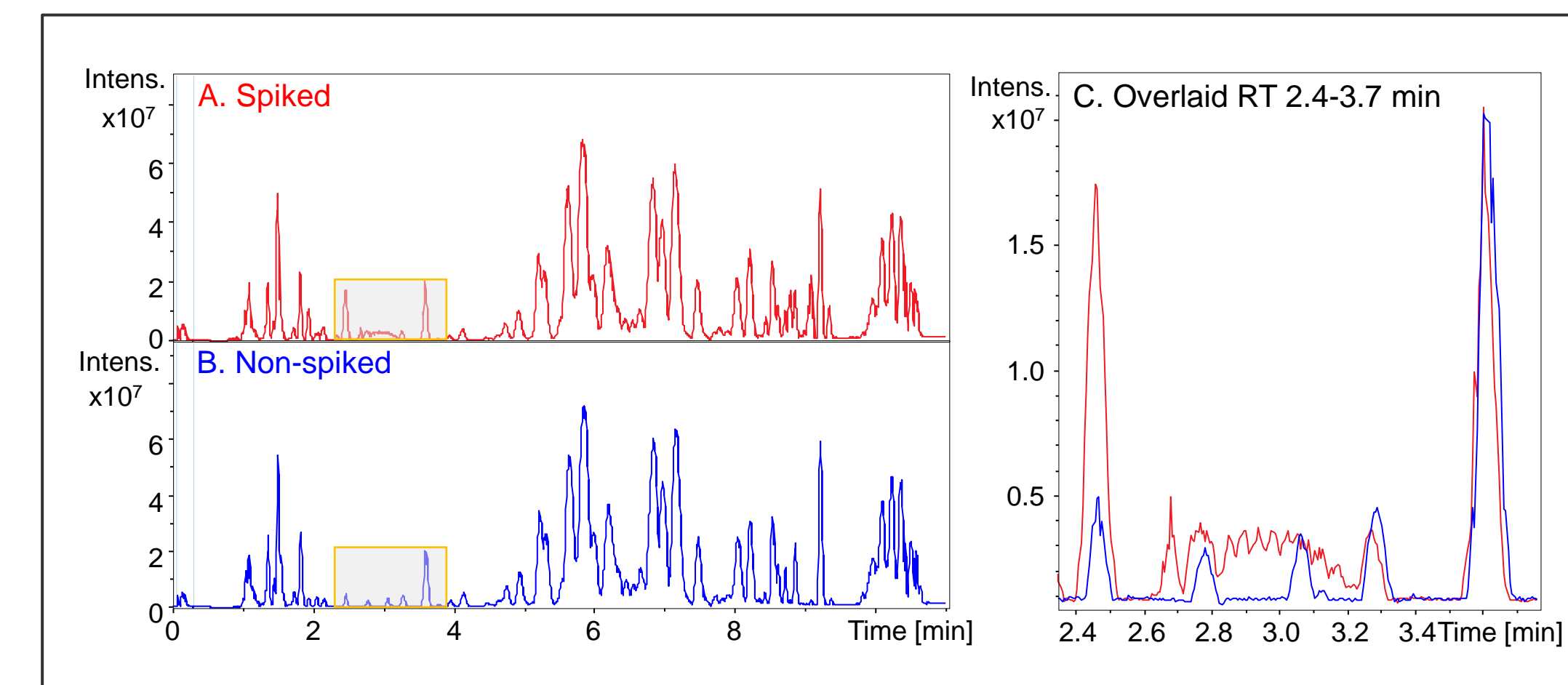


Figure 4. Base peak chromatogram of SRM 1950 lipid extract (A) with and (B) without LNP component mixture spike, (C) Overlaid chromatogram of RT 2.4 – 3.7 min.

## References

- [1] Anderson EJ, et al. (2020). Safety and immunogenicity of SARS-CoV-2 mRNA-1273 vaccine in older adults. *N. Engl. J. Med.* 383:2427–2438.
- [2] Polack FP, et al. (2020). Safety and efficacy of the BNT162b2 mRNA Covid-19 vaccine. *N. Engl. J. Med.* 383:2603–2615.
- [3] Hajj KA, Whitehead KA (2017). Tools for translation: non-viral materials for therapeutic mRNA delivery. *Nat. Rev. Mater.* 2:17056.
- [4] Koltover I, Salditt T, Rädler JO, Safinya CR (1998). An inverted hexagonal phase of cationic liposome-DNA complexes related to DNA release and delivery. *Science* 281:78–81.
- [5] Felgner J, Martin M, Tsai Y, Felgner PL (1993). Cationic lipid-mediated transfection in mammalian cells: "Lipofection". *J. Tissue Cult. Methods* 15:63–68.
- [6] Meng C, Chen Z, Li G, Welte T, Shen H (2021). Nanoplatforams for mRNA therapeutics. *Adv. Ther.* 4:2000099.

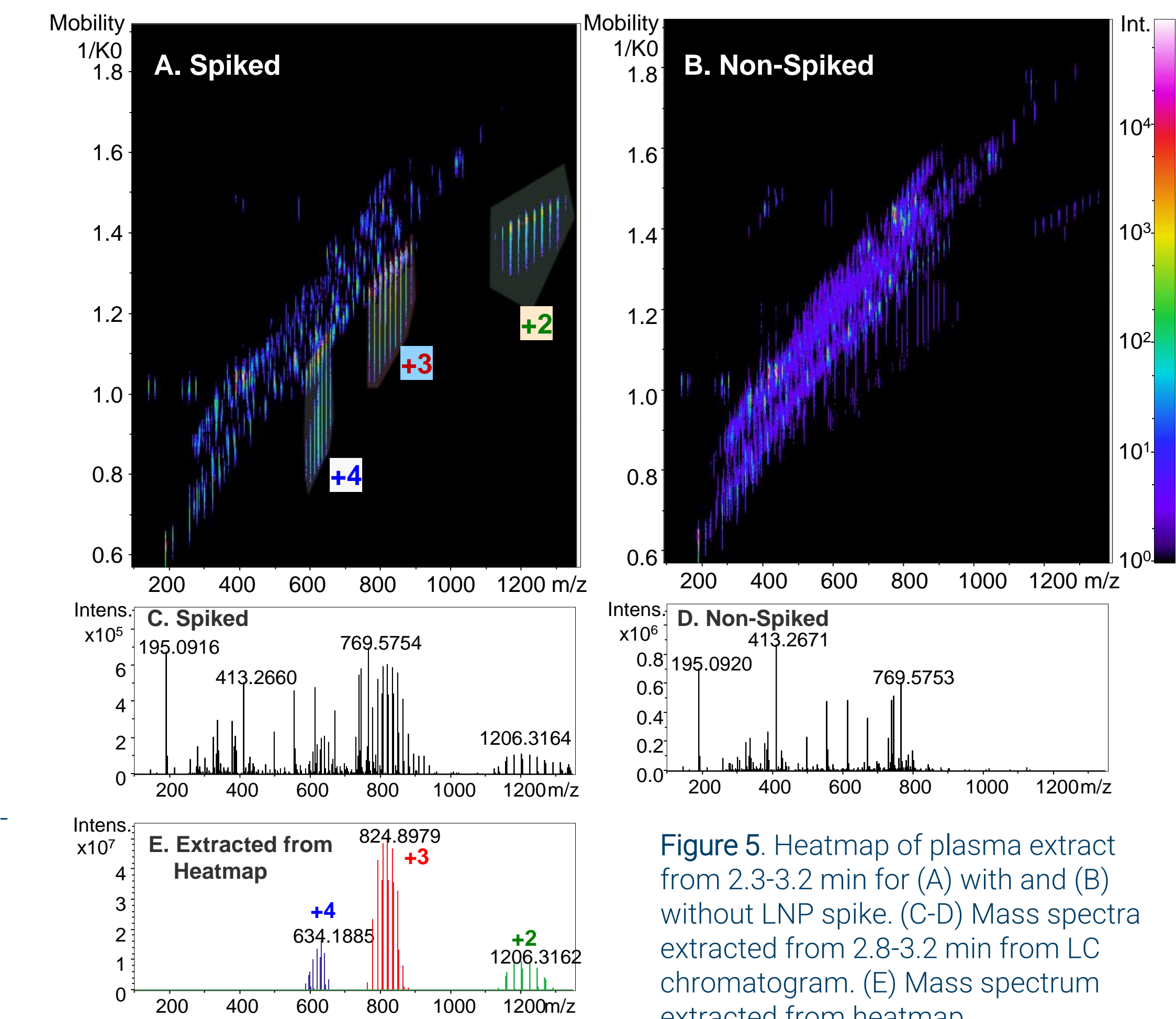


Figure 5. Heatmap of plasma extract from 2.3-3.2 min for (A) with and (B) without LNP spike. (C-D) Mass spectra extracted from 2.8-3.2 min from LC chromatogram. (E) Mass spectrum extracted from heatmap.

## Summary

- LNP components are readily separated and detected by reverse phase UHPLC-MS. VIP-HESI source improved ion signals by 5-25 folds (Fig. 2-3).
- CCS values of all components are measured by TIMS which provides additional dimension of separation for characterization (Fig. 3).
- PEGylated lipids are co-eluting with endogenous species when LNP mixture is spiked into plasma extract. They can be well separated in the TIMS dimension (Fig.4-5).

## Conclusion

- Rapid analysis of LNP components is provided by UHPLC-TIMS-HRMS. The VIP-HESI source demonstrates significant sensitivity enhancement for ionizing LNP materials.
- The additional dimension of ion mobility spectrometry provides CCS information for LNP compound identification and can separate PEGylated lipids from complex plasma samples.

4D-Lipidomics Elute-timsTOF Pro 2