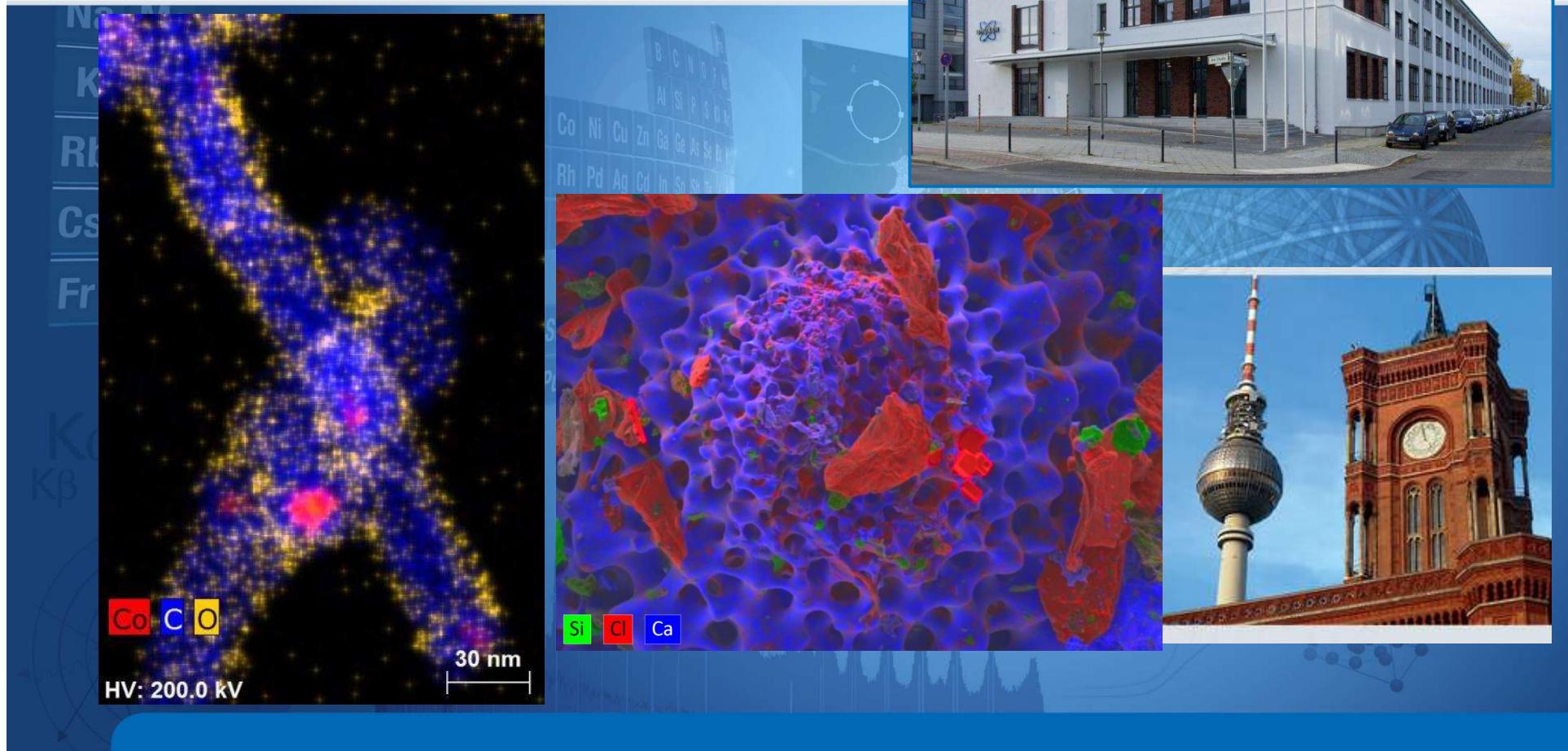


# Element Mapping for Life Science; EDX Analysis of Organic and Soft Materials and their Interface to Minerals in the Electron Microscope



Meiken Falke et al., September 2020



# Element Mapping for Life Science; EDX Analysis of Organic and Soft Materials and their Interface to Minerals in the Electron Microscope

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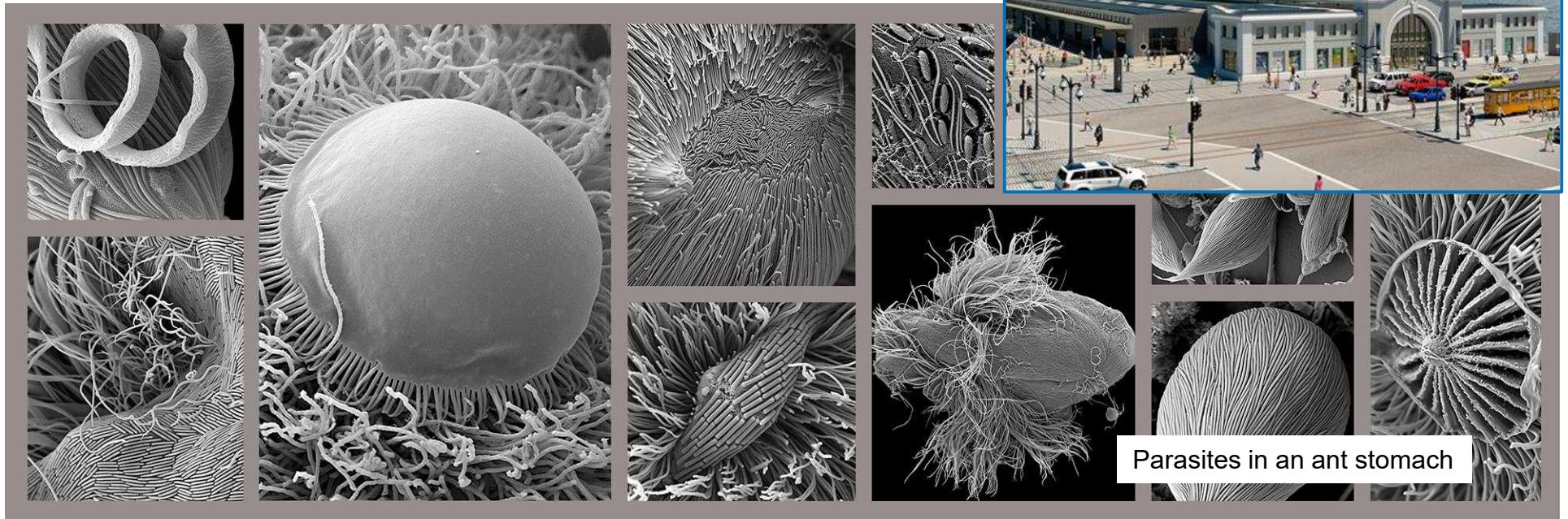


## Dr. Meiken Falke

Product Manager TEM-EDS, Bruker Nano Analytics

# Kevin J. Carpenter, SEM- Microbe Exhibit for Exploratorium

[kevinjcarpenter.com](http://kevinjcarpenter.com)

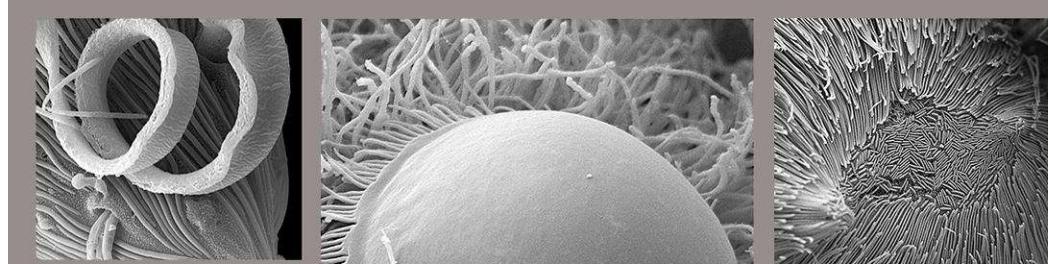


# Kevin J. Carpenter, SEM- Microbe Exhibit for Exploratorium

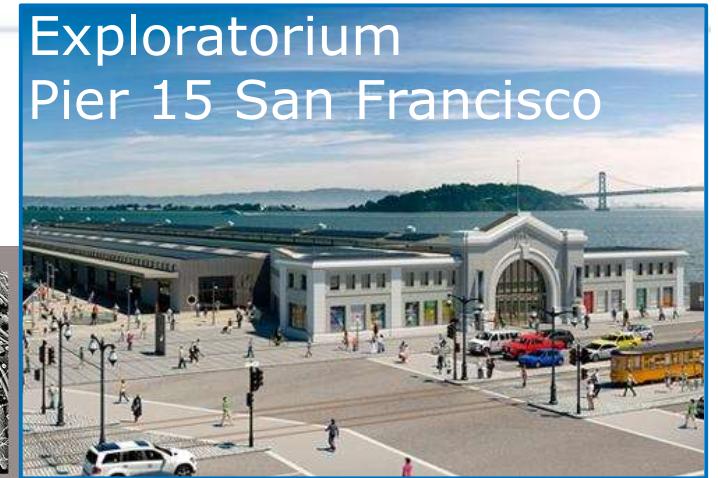
[kevinjcarpenter.com](http://kevinjcarpenter.com)



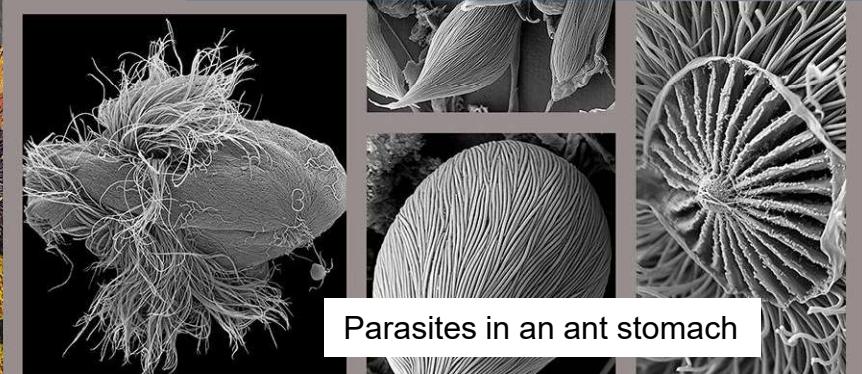
Bruker:  
Add colour using element sensitive detectors!



Exploratorium  
Pier 15 San Francisco



Y. Asisi: Carolas Garden

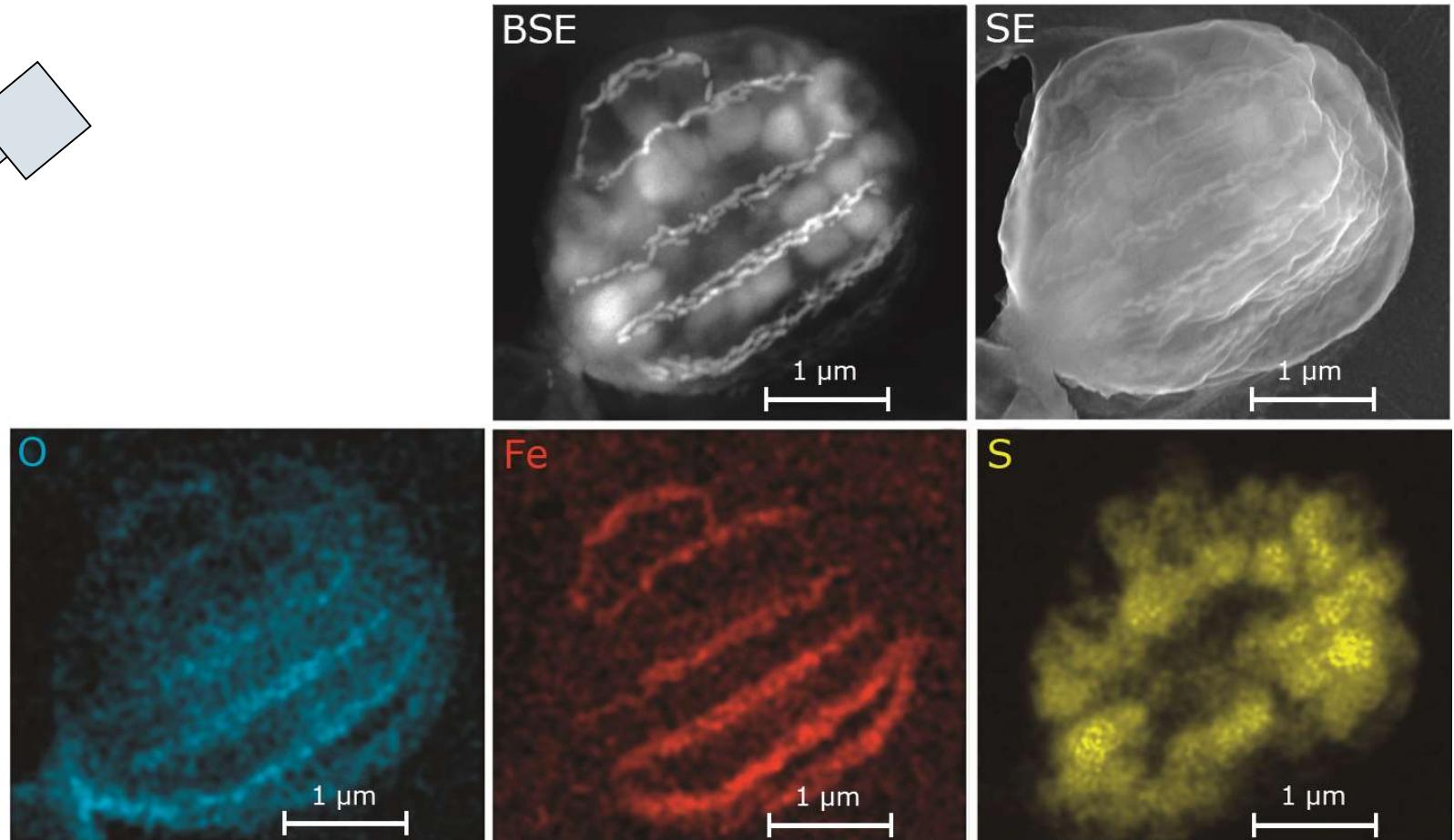
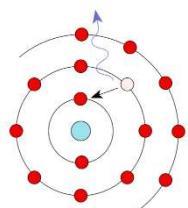
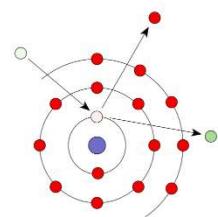
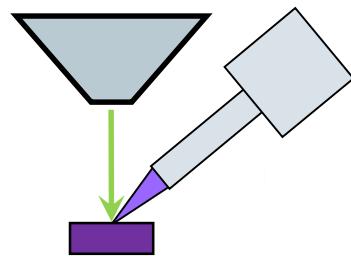
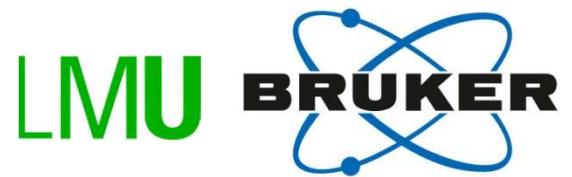


Parasites in an ant stomach



In SEM and TEM ...

# SEM: EDS-Double Detector System; Electron microscopy in **color**: Magnetotactic Bacteria

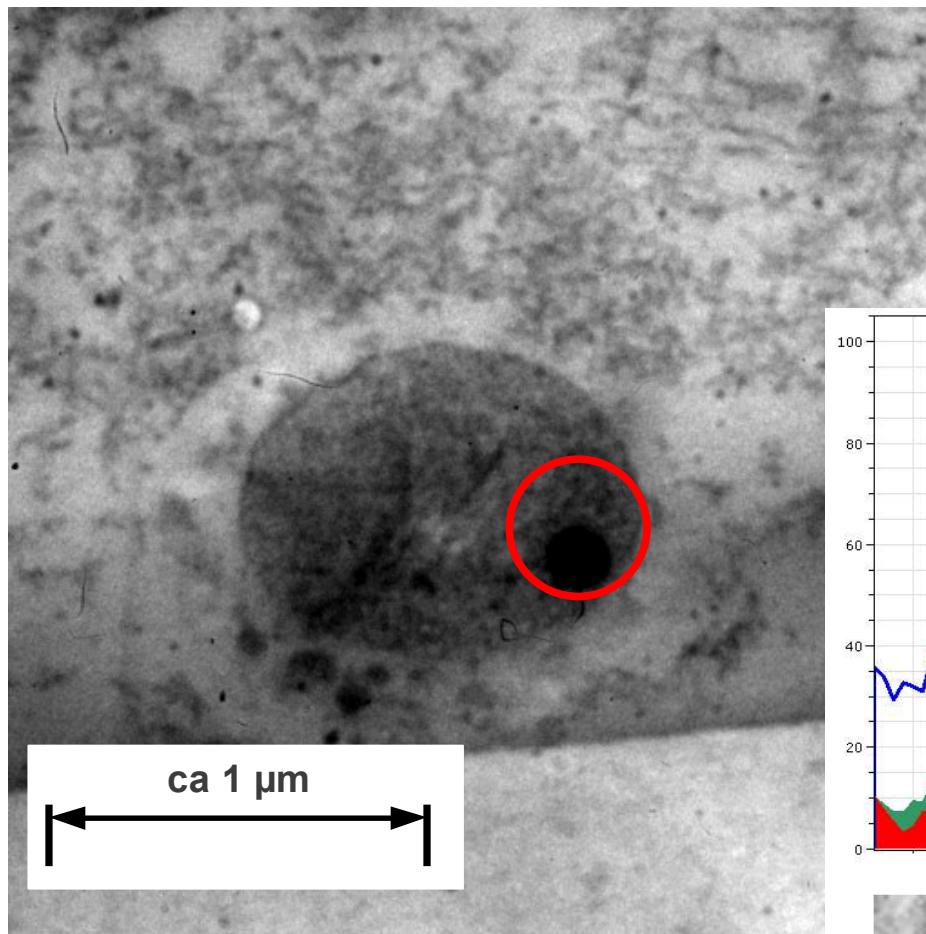


# SDD-EDS in (S)TEM Fe oxidising bacteria ~0.1sr

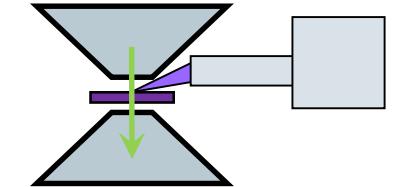


Universität  
Regensburg

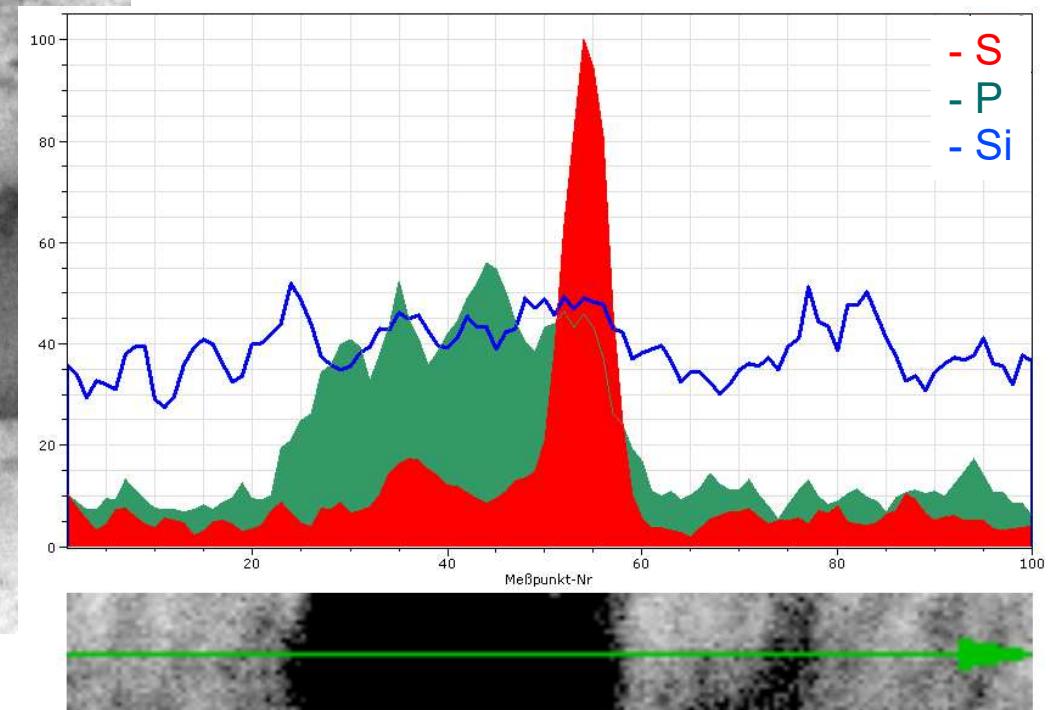
Φ(R)  
Uni Regensburg

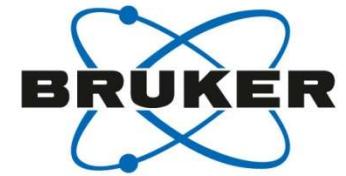


TEM/EDS: Prof. J. Zweck,  
University of Regensburg  
Sample: Thomas Heimerl  
Group Prof. Rachel



Fe particle in Phosphorus close to cell membrane,  
with sulfur. Work in progress





# Outline

---

- Intro:
  - EDS for life science?
  - EM: TEM, STEM and T-SEM EDS
- Available Technology
- EDS Data Acquisition, Quantification and Display of Results
  - Various examples demonstrating ESPRIT implementation of qualitative and quantitative analysis for different specimen types
- Element Mapping in Liquid, Ice, *in-situ*
- Complementary Techniques

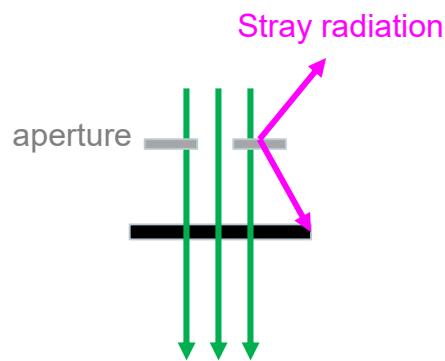


# EDS for Life Science; Suitable Specimens

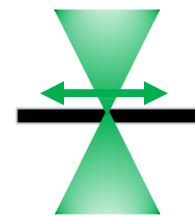
- Bio-mineralization and related:
  - Bio-minerals (bone, teeth, Fe etc. in tissue, magnetotactic bacteria, crustaceans, egg shells, ...)
  - Bio-mimetics (sea urchin, spider web, wood, collagen ... tissue engineering)
  - Nanoparticles > nanotoxicity vs
  - Useful nanoparticles; cell uptake (drug delivery)
  - Distinguish immunolabels, labels for CLEM
- SDDs have become so sensitive in the low energy region, that the detection of small amounts of bio-relevant light elements (N, P, S, O, ...) is no problem anymore!
- Mapping of element and light element distribution in cells and organelles >
- Mapping of nearly **the whole periodic system!**
- CLEM (labels), in liquid cells or ice?

# Electron Microscopy

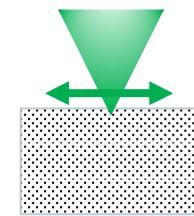
TEM



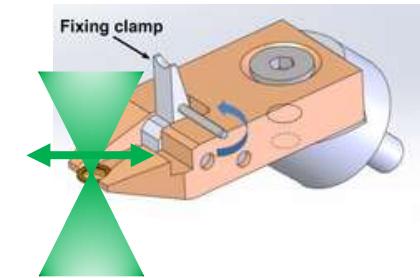
STEM  
Scanning TEM



SEM  
Scanning EM



SEM: „T-SEM“



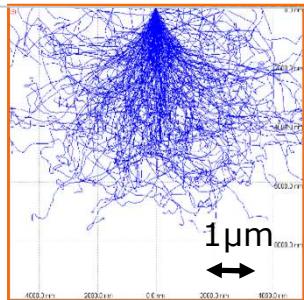
- TKD patented holder
- Commercial STEM holders
- Home made versions

# Spatial Resolution and Cs-correction

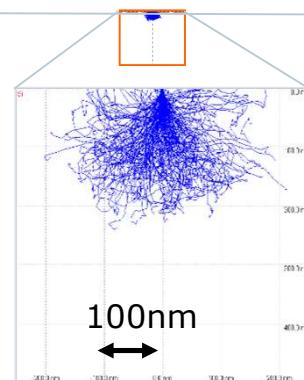


SEM: bulk

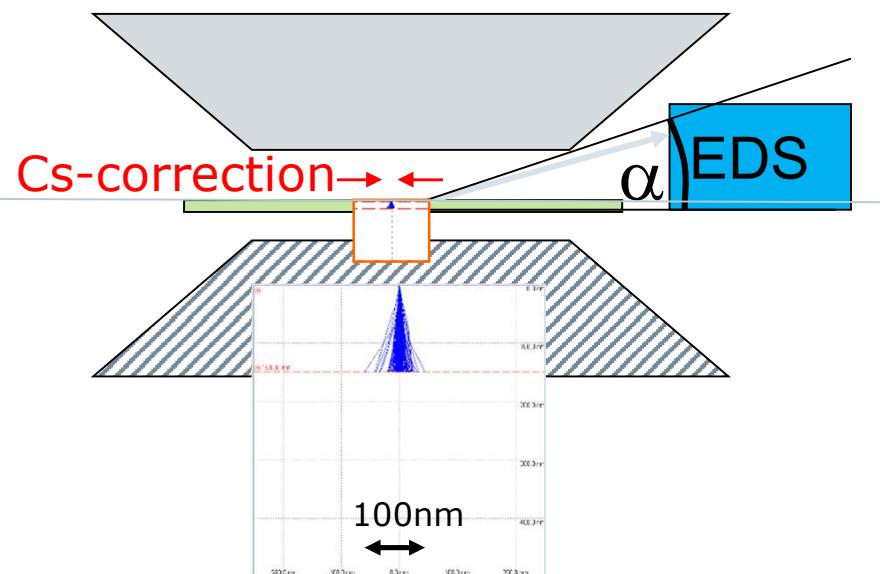
High voltage  
30kV



Lower voltage  
4kV



S/TEM, T-SEM: thin specimen,  
small probe

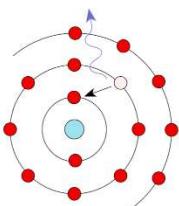


# (TEM) EDS Quantification; R. Egerton

1994, line intensity for a particular element line / transition



$$I_x = N_A \sigma_A \omega_A (\Omega/4\pi) \varepsilon N_e = n_A t \sigma_A \omega_A (\Omega/4\pi) \varepsilon N_e$$



$I_x$

number of X-ray photons in a characteristic peak of species A

$N$

number of atoms per unit volume

$n t$

number of atoms per unit area times thickness

$\sigma$

ionization cross section (Casnati et al., 1982, Bote et al., 2009)

$\omega$

fluorescence yield (Hubbell et al., 1994, Krause, 1979)

$\Omega/4\pi$

solid angle / geometrical collection efficiency

$\varepsilon$

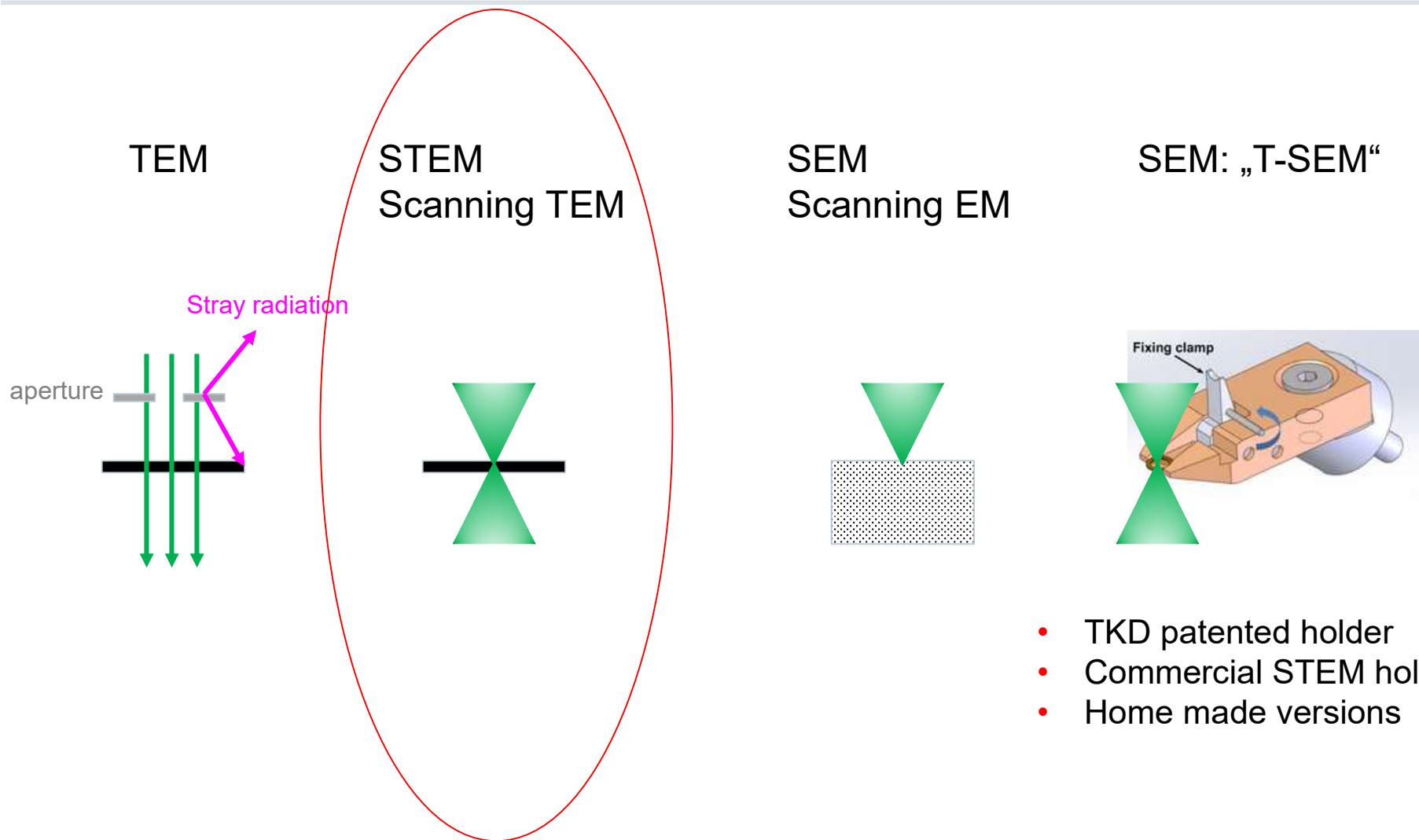
detection quantum efficiency (window: SLEW or no window or other)

$N_e$

number of incident electrons

+ absorption, fluorescence, other effects...

# Electron Microscopy

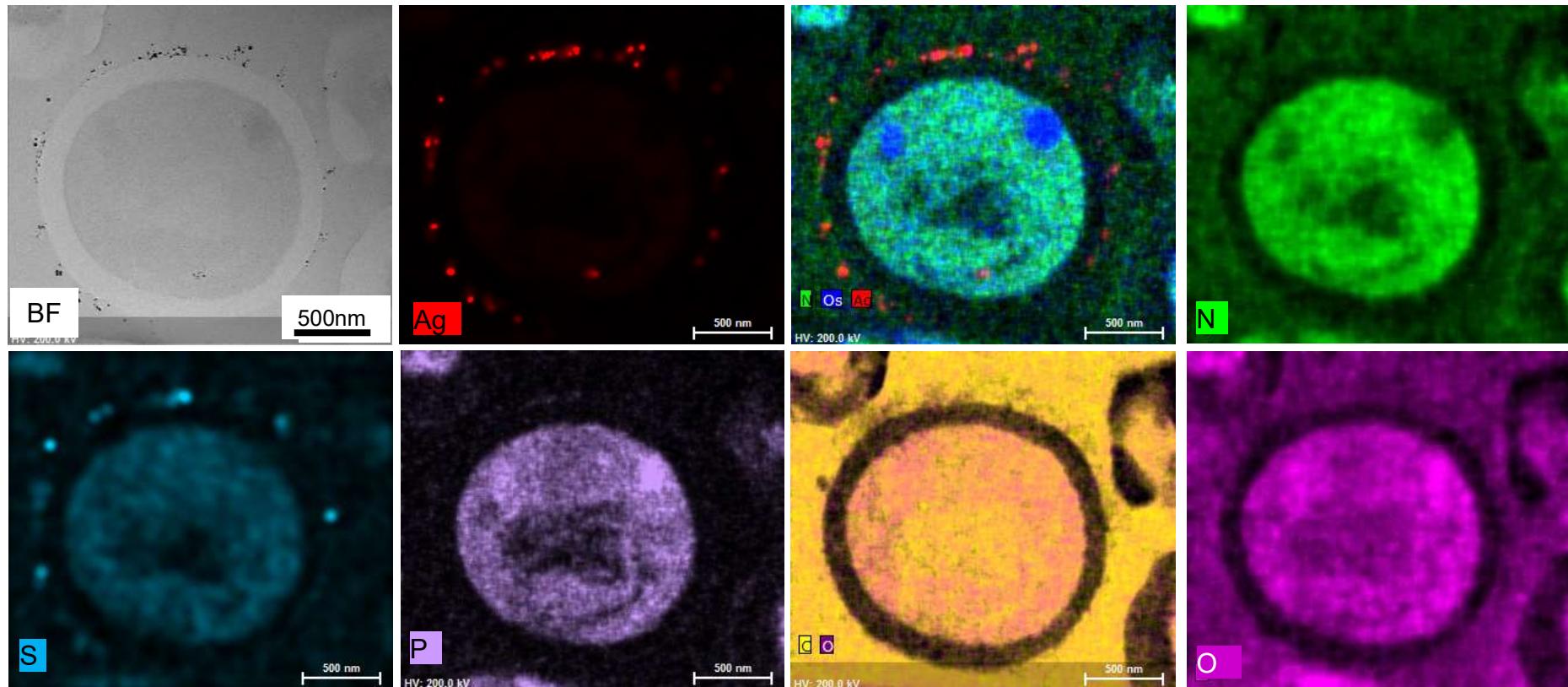


- TKD patented holder
- Commercial STEM holders
- Home made versions

# TEM EDS for Life Science

Yeast cell: Element mapping of protein (Ag) labels and light (N, O) and heavy (Os, Ag) elements

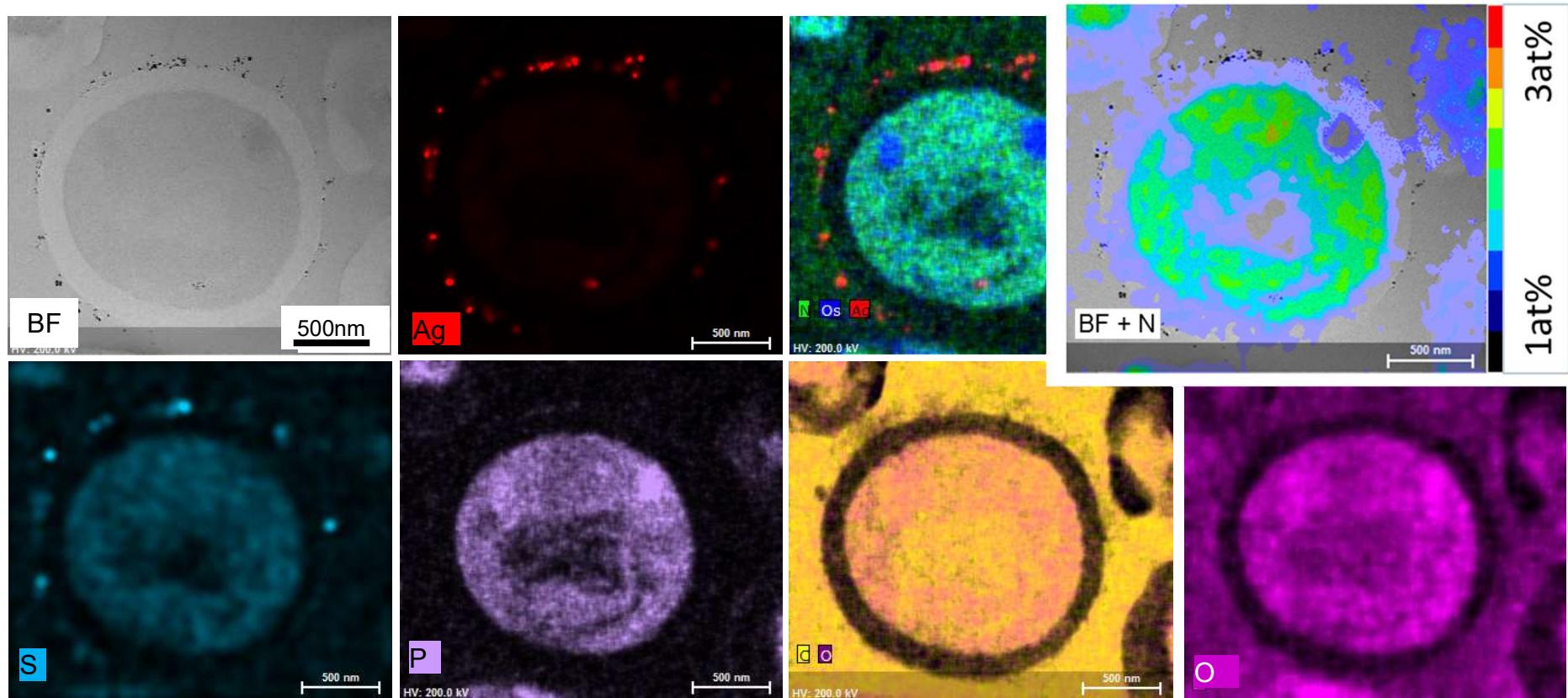
30 mm<sup>2</sup>, 0.12 sr (Standard EDS); Conventional STEM

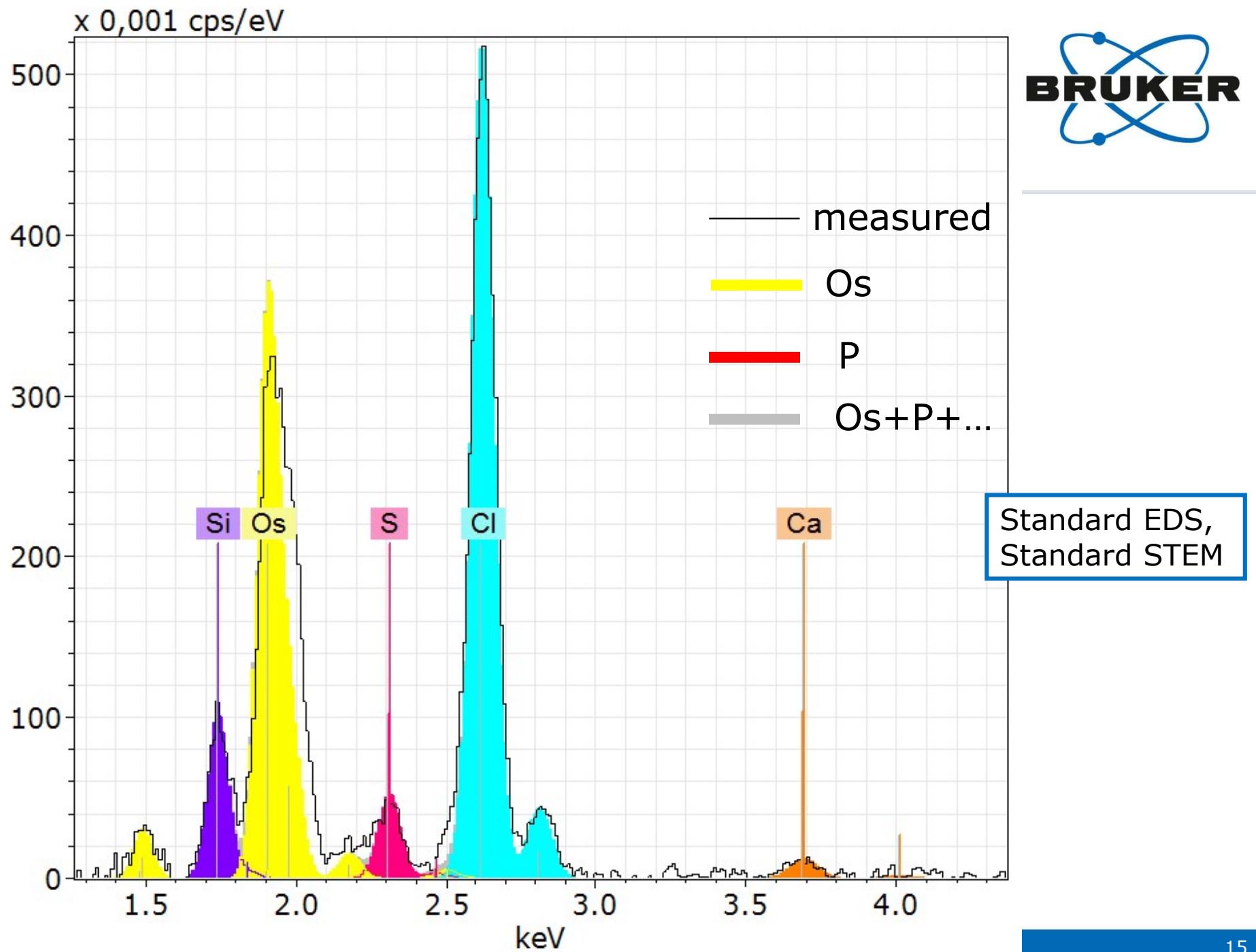


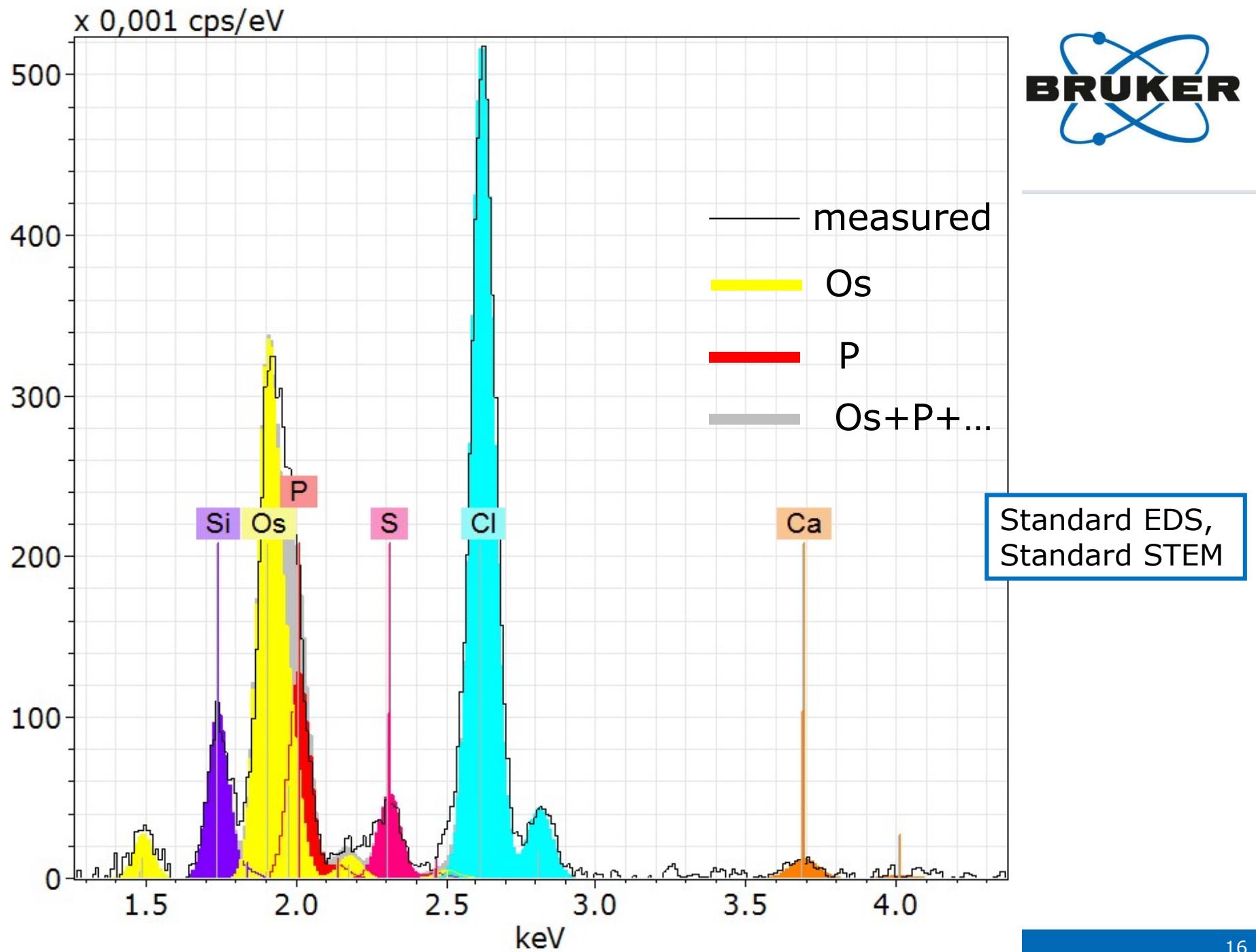
# TEM EDS for Life Science

Yeast cell: Element mapping of protein (Ag) labels and light (N, O) and heavy (Os, Ag) elements quantitatively

30 mm<sup>2</sup>, 0.12 sr (Standard EDS); Conventional STEM







# TEM-EDS for cell analysis at 0.1sr

Malaria parasite: *Plasmodium* in erythrocyte  
(red blood cell) treated with Chloroquine

Standard EDS, Standard STEM

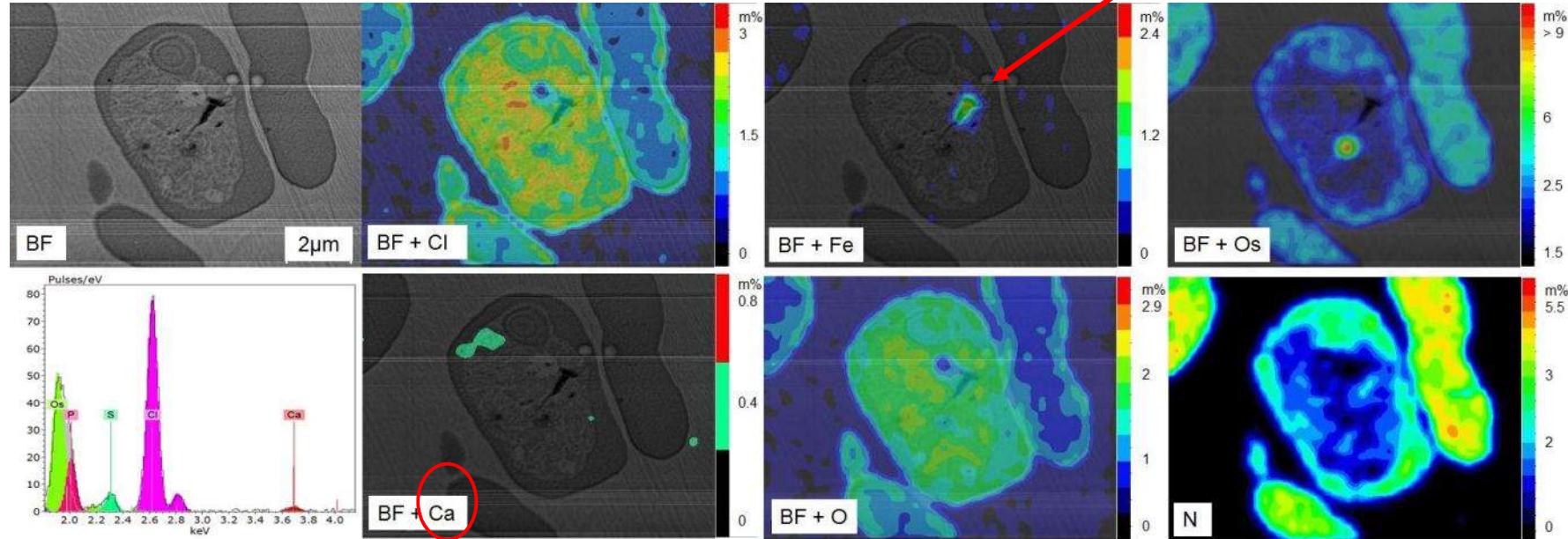


Anopheles mosquito,  
spreader of *Plasmodium falciparum*, the cause of Malaria

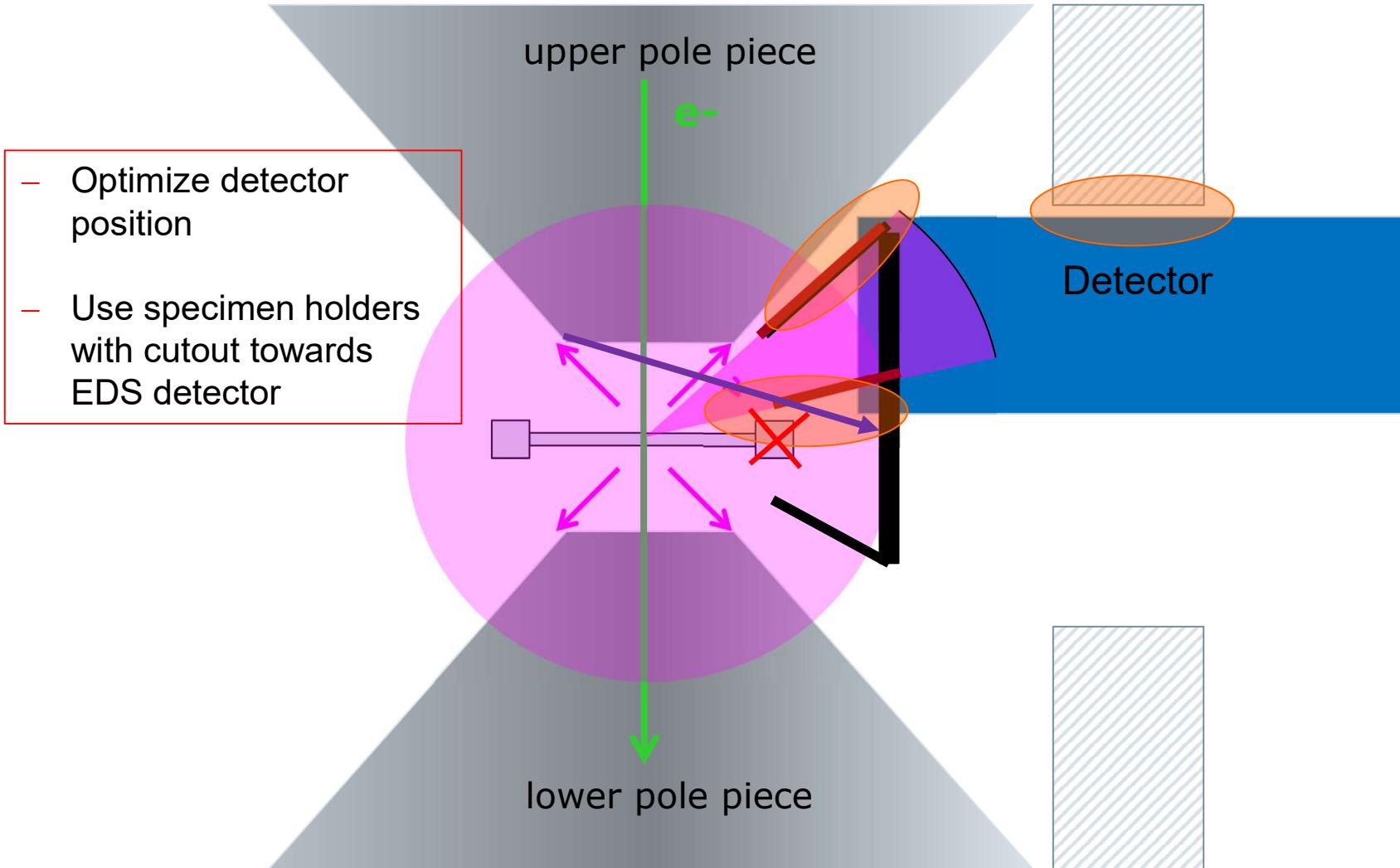
The parasite multiplies by destroying red blood cells



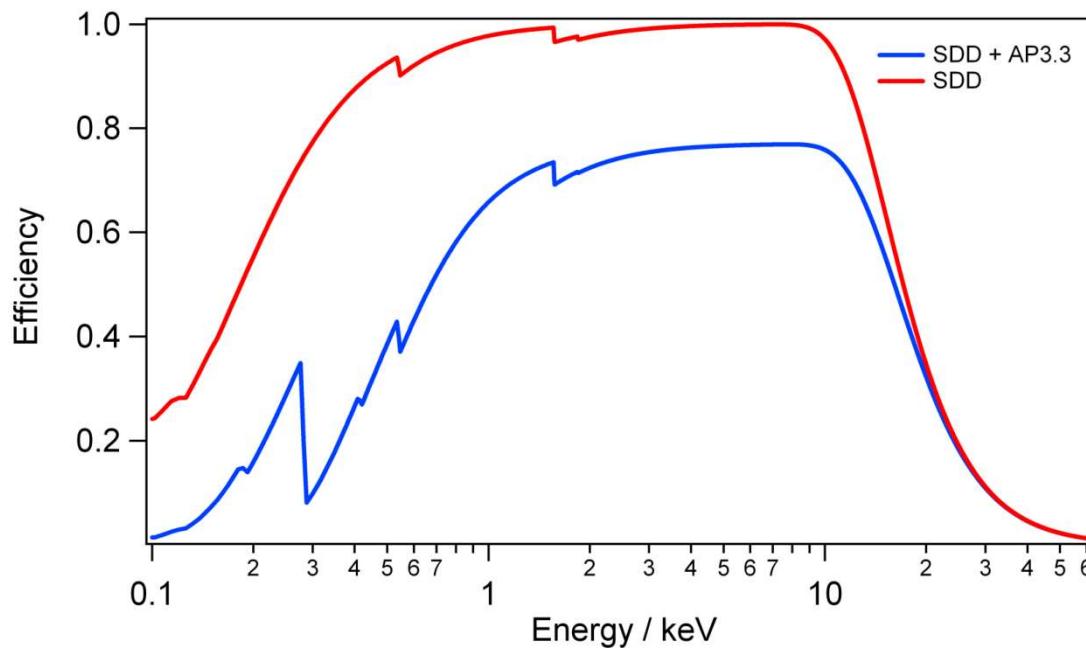
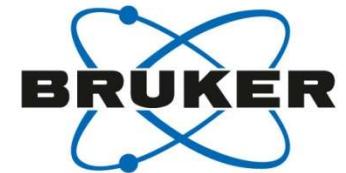
The parasite digests haemoglobin, hence the iron intake in its food vacuole



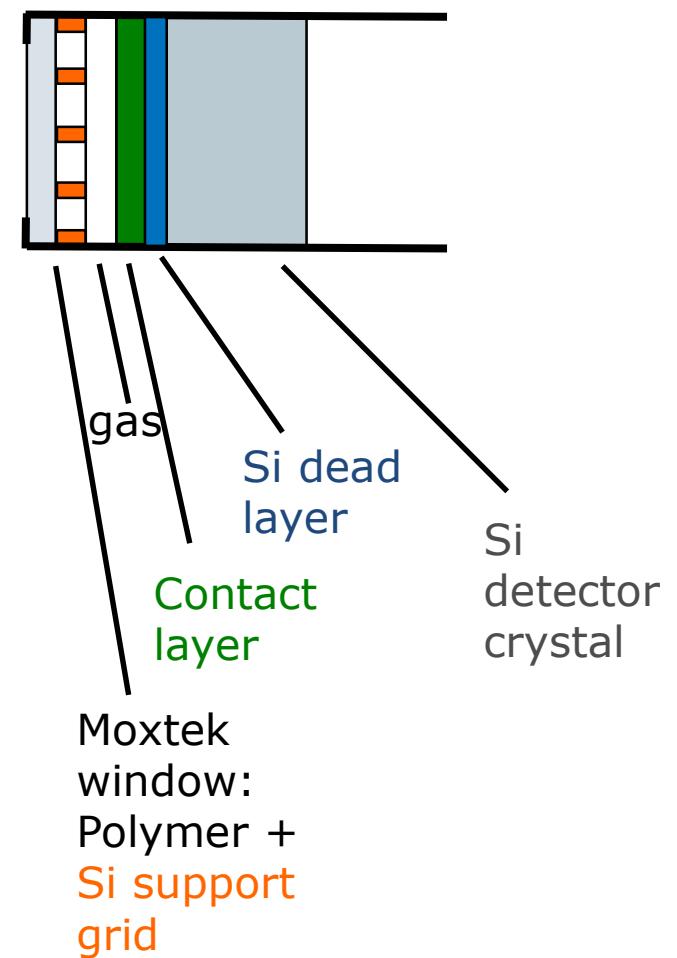
# Geometric Limitations



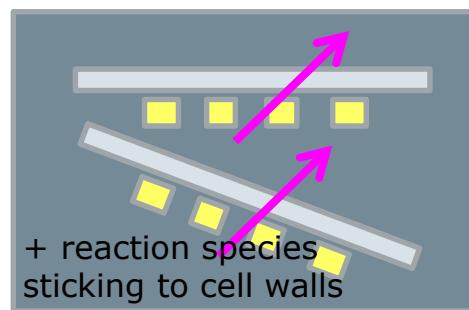
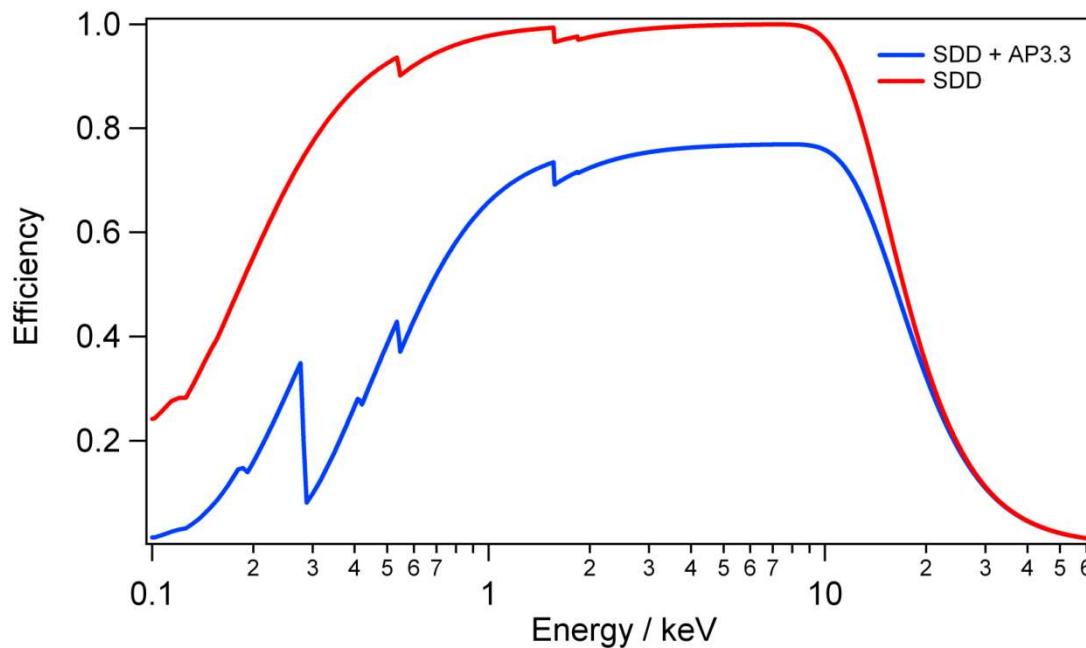
# Detector (window) quantum efficiency and windows of in situ reaction cells



Detector w/wo window

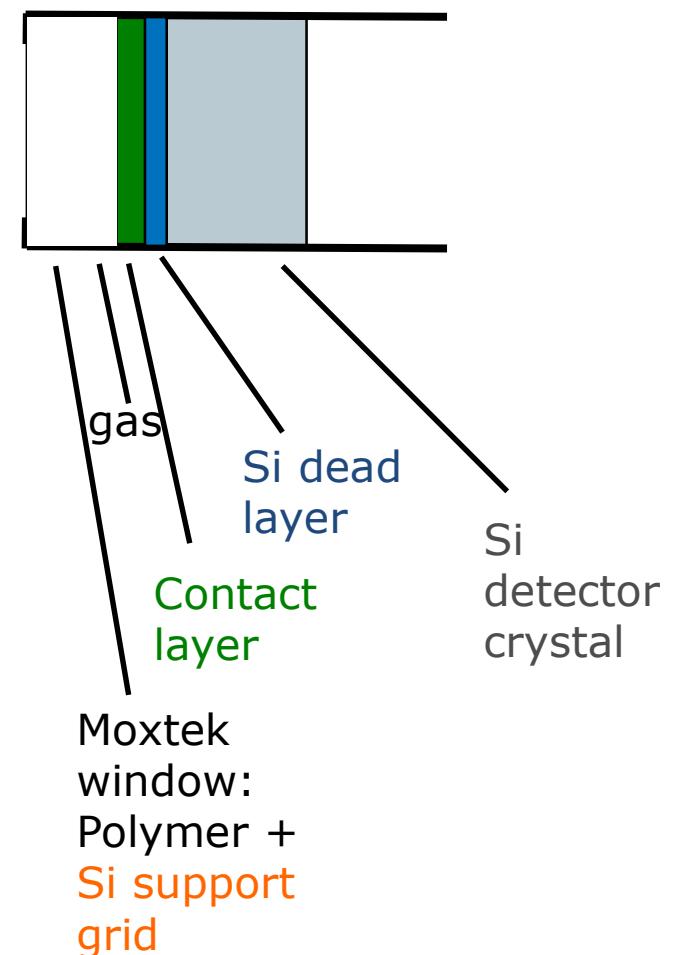


# Detector (window) quantum efficiency and windows of in situ reaction cells

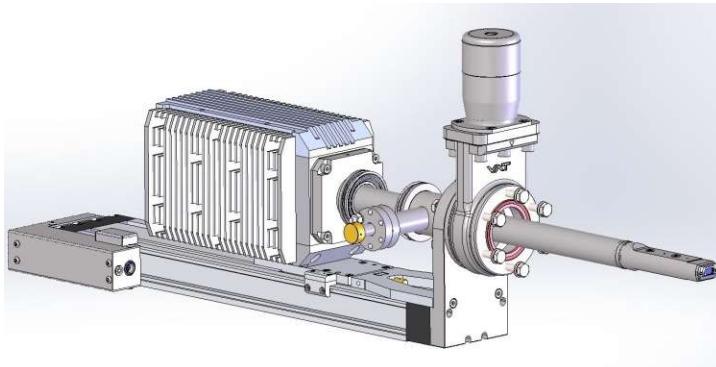


Window of reaction cell

Detector w/wo window



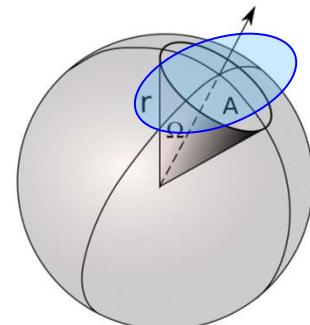
EDXS with 100 mm<sup>2</sup> windowless oval detector area;  
Nion UltraSTEM, Cs-corrected, high brightness source



EDXS at ~0.7 sr. This is the real solid angle for a flat SDD (see wiki below).

Wrong:  $100\text{mm}^2 / (10.5\text{mm})^2 = 0.91\text{sr}$ .

TOA: 13.4°

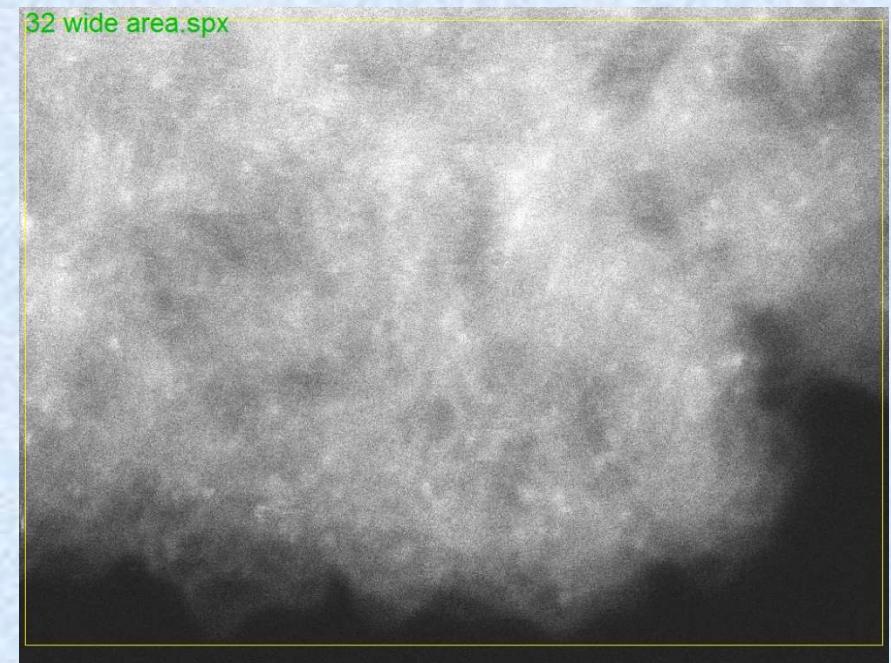
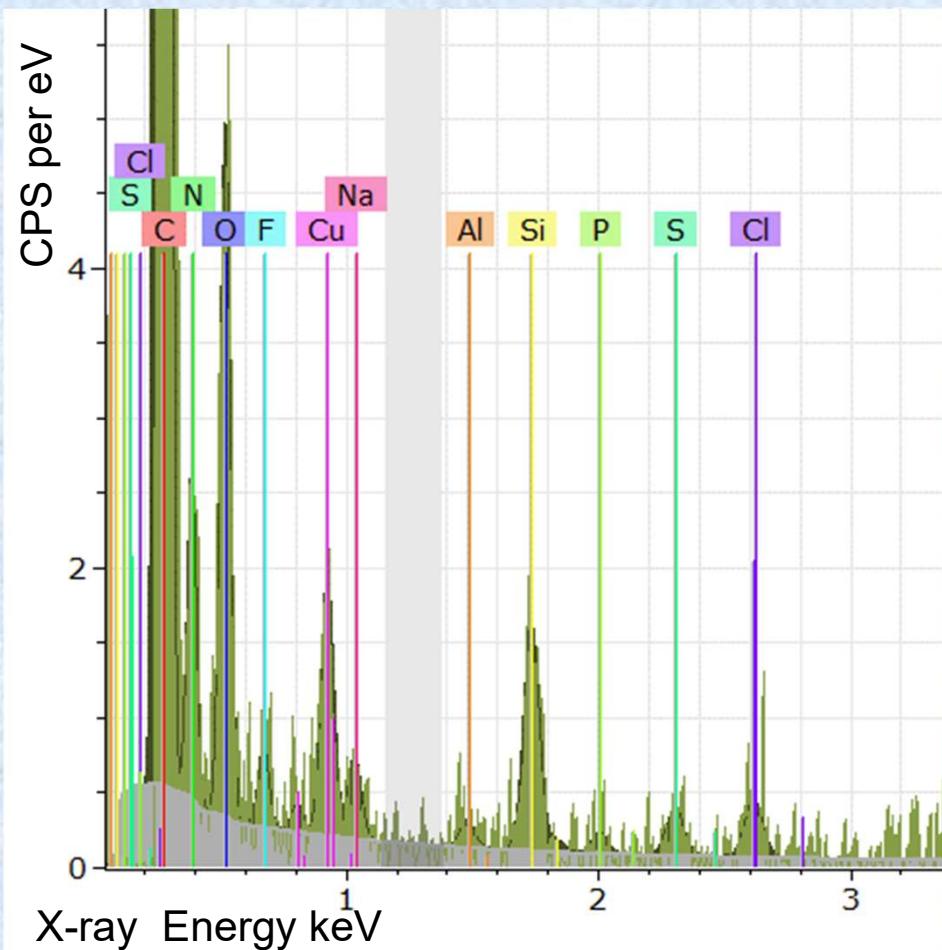


wikipedia

Used to analyse dust from space!  
See next slides ...

# EDXS detects single atoms and concentrations ~0.01%

C (at.%)	N	O	F	Cu	Na	Al	Si	P	S	Cl
96.75	0.84	1.45	0.16	Sys.	0.11	0.05	0.4	0.04	0.08	0.11



NRL UltraSTEM200 with Bruker X-flash detector, 60 kV.  
Concentrations as low as 0.01 atomic % can be explored.  
Courtesy Rhonda Stroud, NRL.



STEM: single atom ID in carbonaceous material, here: nano-diamonds from space; 0.65sr using 100mm<sup>2</sup> oval detector  
<http://creativecommons.org/licenses/by/4.0/>

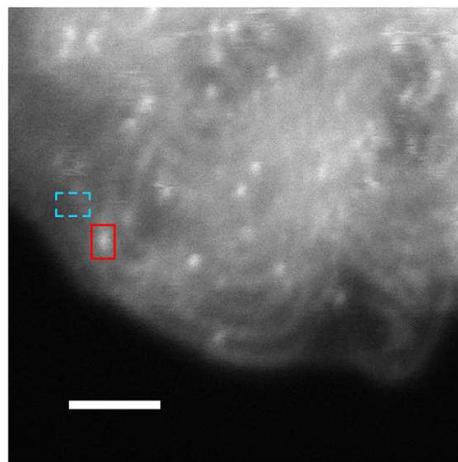
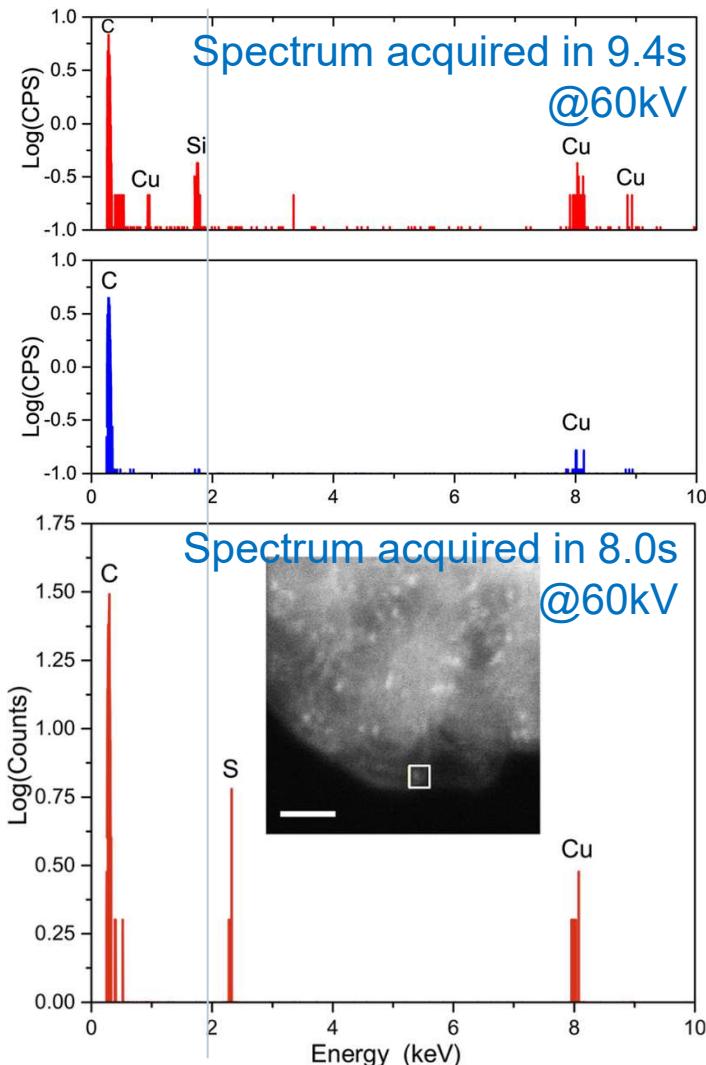
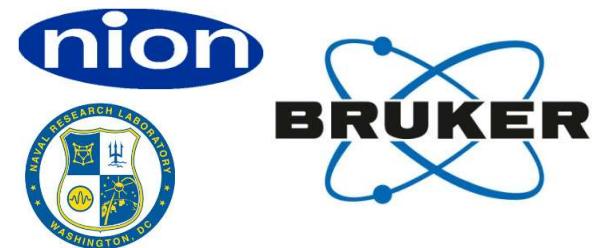
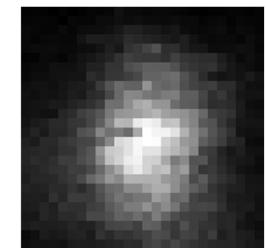


FIG. 1. High-angle ADF image and EDX spectra on (red solid line box) and off (blue-dashed line box) an Si atom on amorphous carbon. The scale bar is 1 nm.

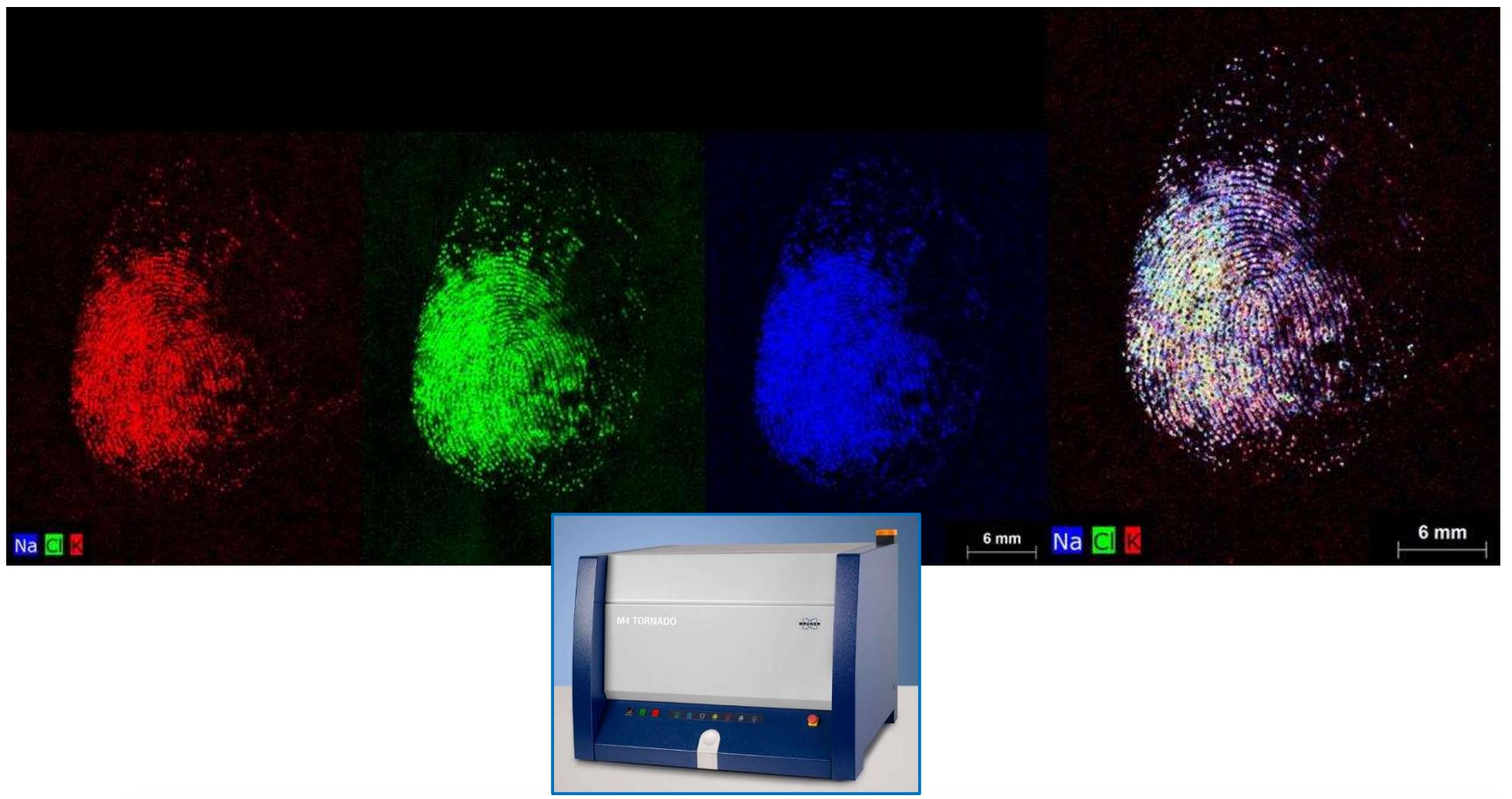


Trackingwindow for EDS of single atom.

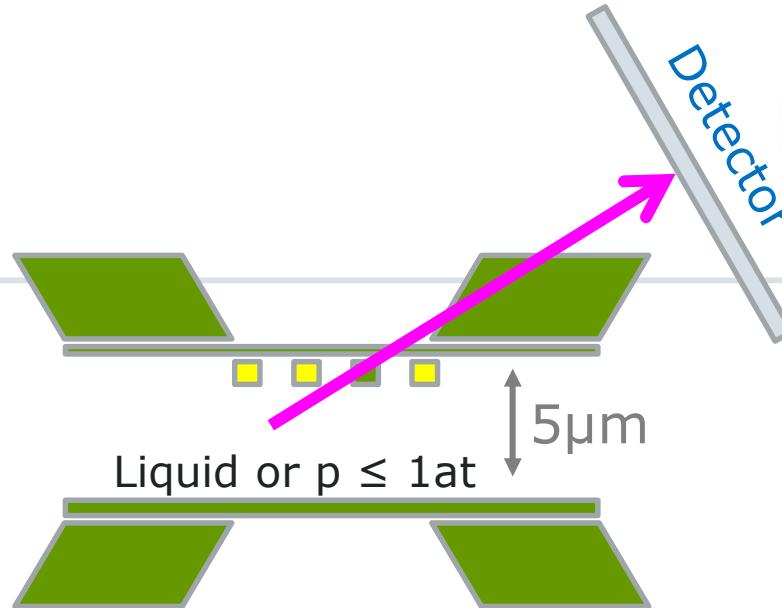
FIG. 2. High-angle ADF image and EDX spectrum of an S atom on amorphous carbon. The scale bar is 1 nm.

R. M. Stroud et al., *APL* **108**, 163101 (2016) open access  
T. C. Lovejoy et al., *APL* **100**, 154101 (2012): 30mm<sup>2</sup>, 0.1sr

# Sources of foreign signals? Fingerprint on Paper; X-Ray fluorescence in the M4 Tornado



# Reaction Cells



- Shadowing
- Gas, liquid, + heat:
- Windows:
  - Graphene, Si<sub>3</sub>N<sub>4</sub> or alpha SiNx
  - Thickness,
  - Support (grid): Si, polymer film, SiC (heating), Au spacers O-rings
  - Al film (light tight))
- Reaction species sticking to cell walls

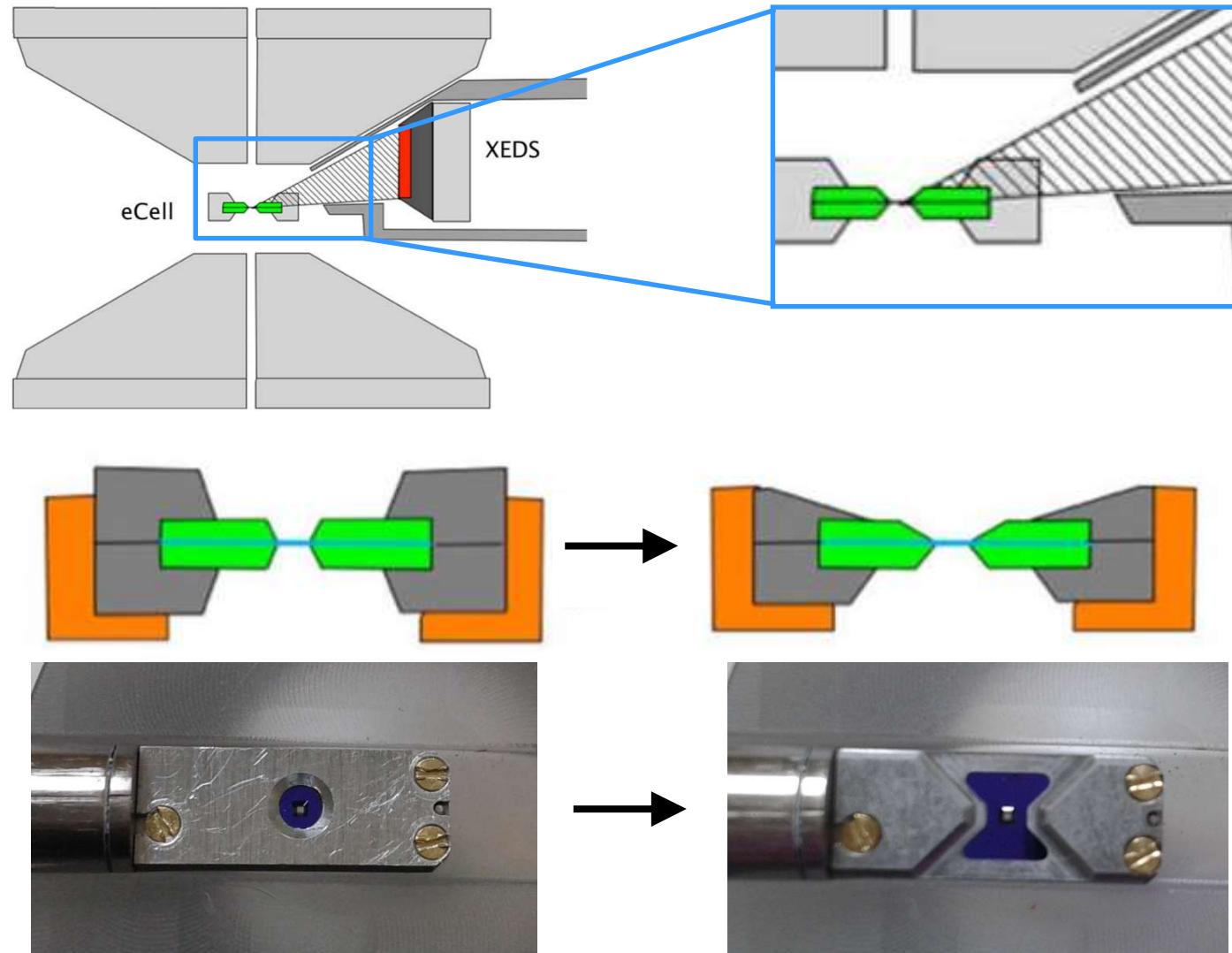
➤ **Effects:**

- Windows etc. > Reduced transmission / quantum efficiency
- Cell Materials > System peaks
- Heat > enlarged peak width and spectrum background / noise

calculate transmission depending on thickness/materials here:

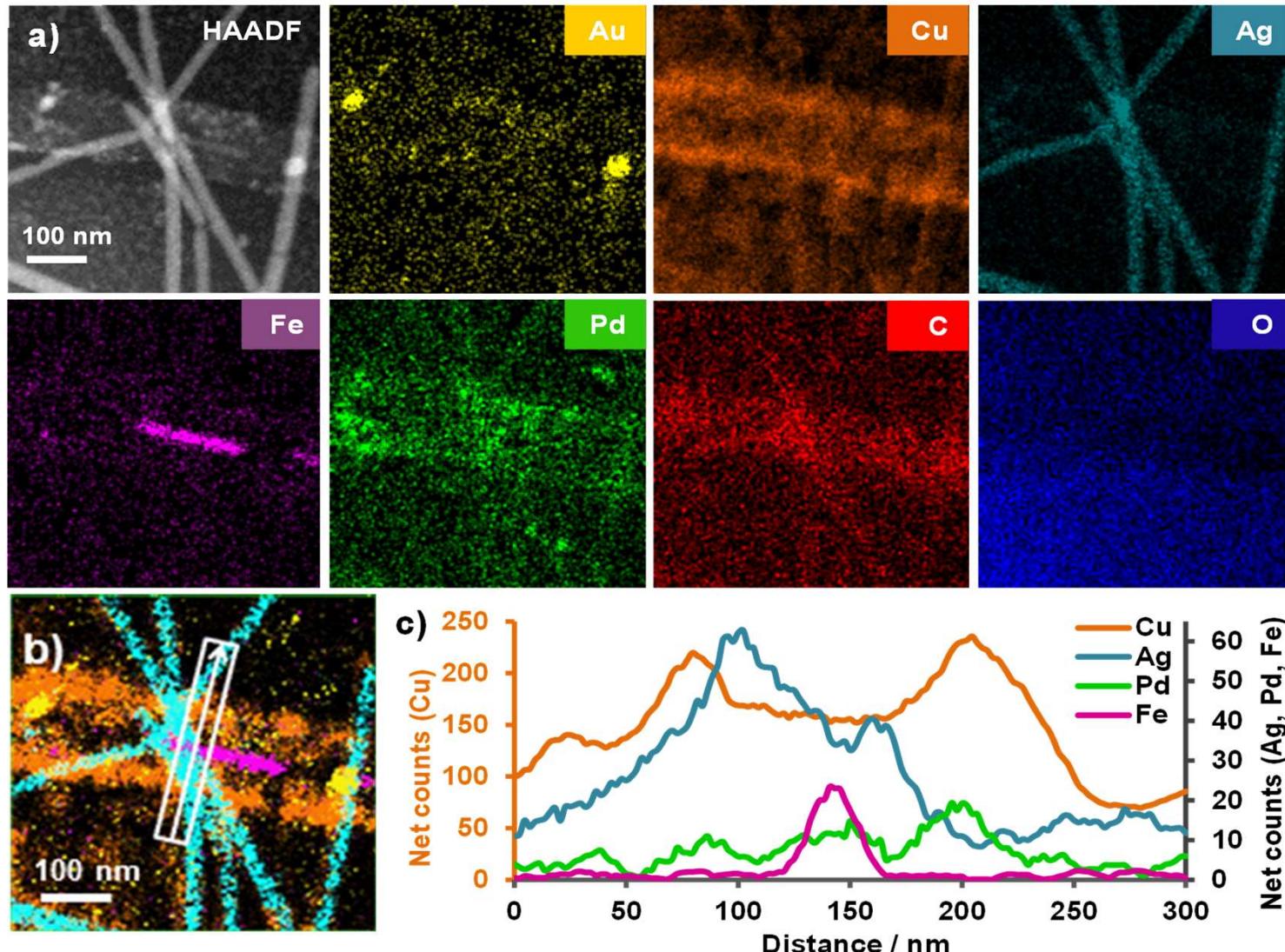
[http://henke.lbl.gov/optical\\_constants/filter2.html](http://henke.lbl.gov/optical_constants/filter2.html)

## EDXS compatible environmental cell design



Zaluzec *et al.* Microsc. Microanal. (2014) 20 p. 323

# EDXS Elemental mapping in liquids



# Cryo-STEM-EDS: M&M 2018 with N.J. Zaluzec and TFS

ThermoFisher  
SCIENTIFIC

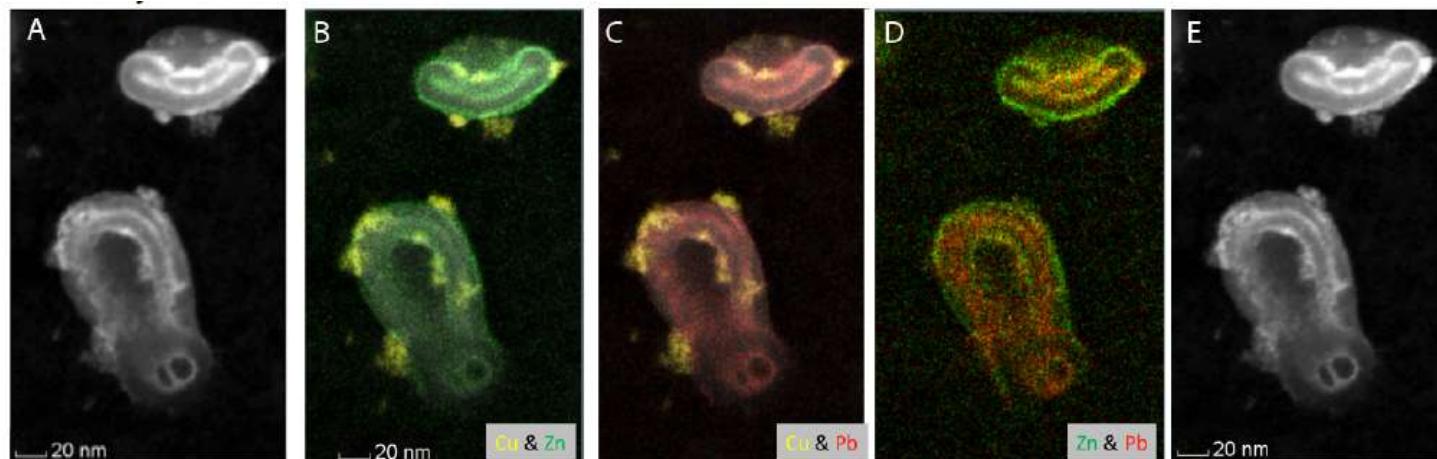


Figure 1.) Hyperspectral images of site specific heavy metal localization to folds of polyamide film.  
A) HAADF preXEDS, B) Cu & Zn, C) Cu & Pb, D) Zn & Pb, E) HAADF postXEDS measurement (333 Frames)

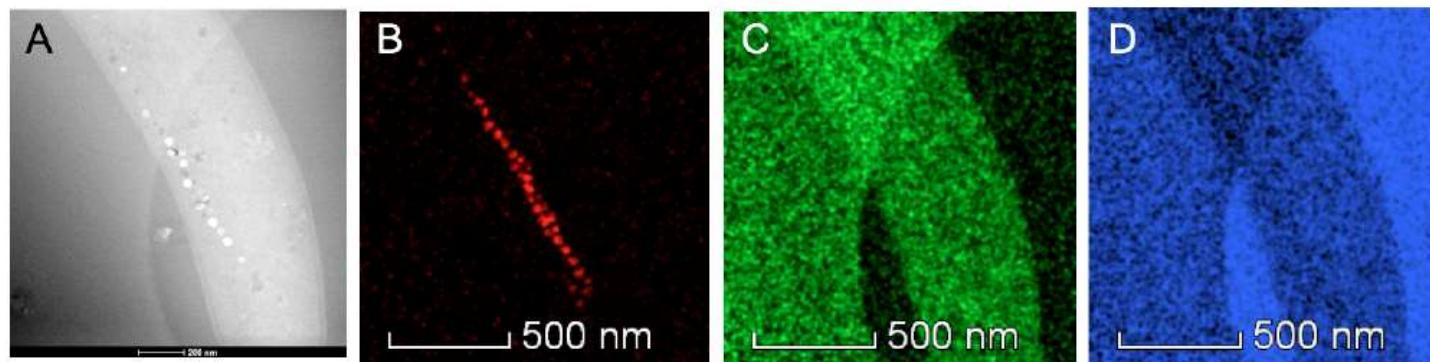
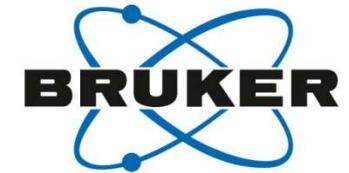


Figure 2.) Hyperspectral images of frozen magnetotactic bacteria,  $E_0:200$  kV, 5 pA, Dual X detector.  
A) HAADF image, B) Fe , C) Carbon, D) Oxygen elemental distributions.

# Cryo-STEM-EDS



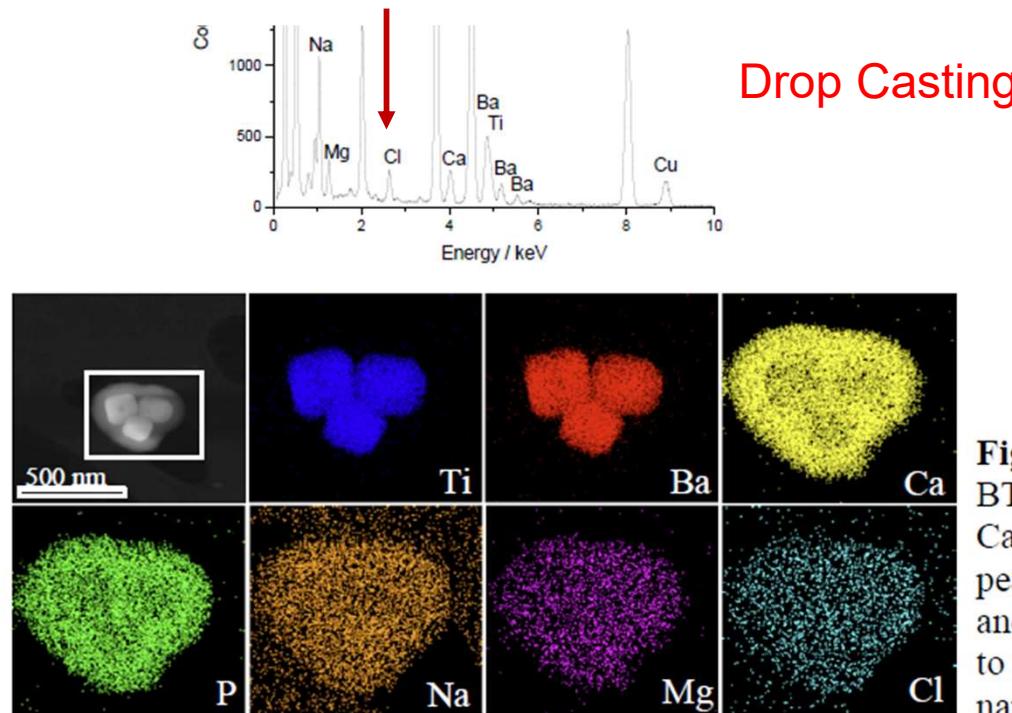
Drying artefacts for drop casted NP in CCCM,  
overcome by plunge-freezing, confirmed by EDS.

IOP Conf. Series: Journal of Physics: Conf. Series **902** (2017) 012006

## Cryo-STEM-EDX spectroscopy for the characterisation of nanoparticles in cell culture media

M Illett<sup>1</sup>, F Bamiduro<sup>1</sup>, O Matar<sup>1</sup>, A Brown<sup>1</sup>, R Brydson<sup>1</sup> and N Hondow<sup>1</sup>

<sup>1</sup> School of Chemical and Process Engineering, University of Leeds, Leeds LS2 9JT,  
UK



**Figure 2:** STEM-EDX data of DC BT in CCCM. In the EDX spectrum Ca and P are the most prominent peaks but also present were Cl, Mg and Na, all of which spatially match to the coating imaged around the BT nanoparticles.

# Cryo-STEM-EDS



Drying artefacts for drop casted NP in CCCM,  
overcome by plunge-freezing, confirmed by EDS.

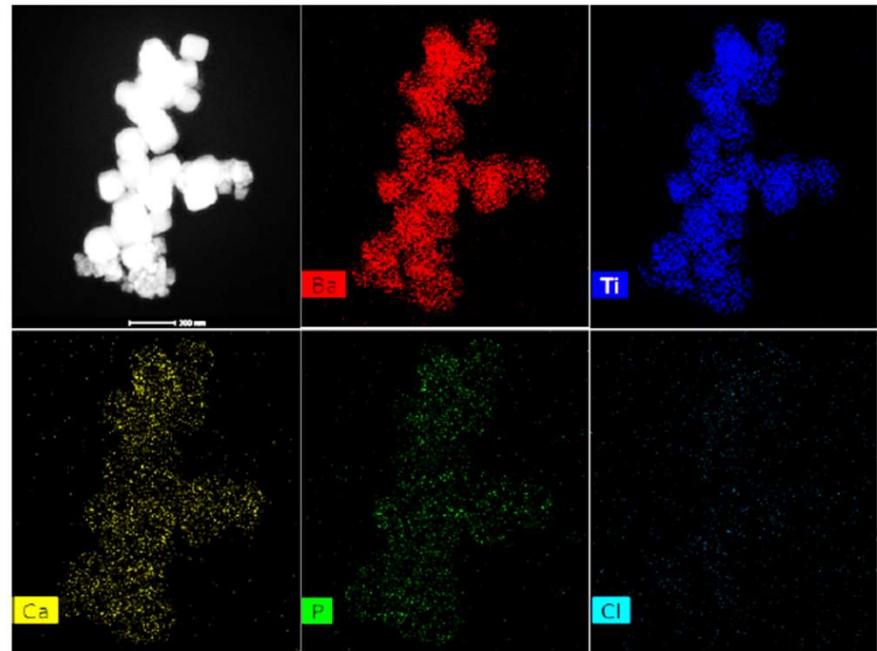
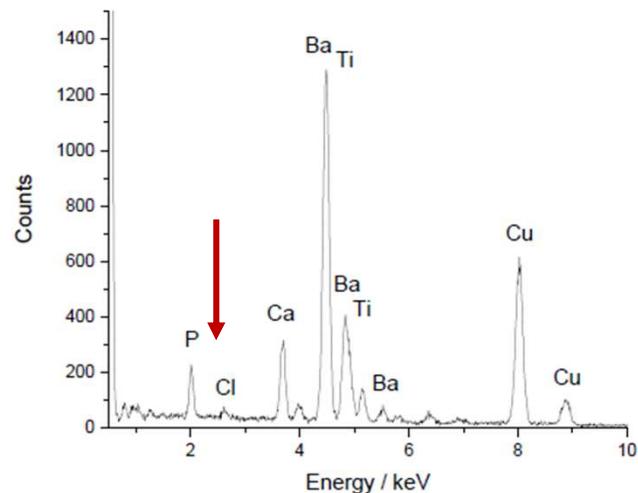
IOP Conf. Series: Journal of Physics: Conf. Series **902** (2017) 012006

**Cryo-STEM-EDX spectroscopy for the characterisation of  
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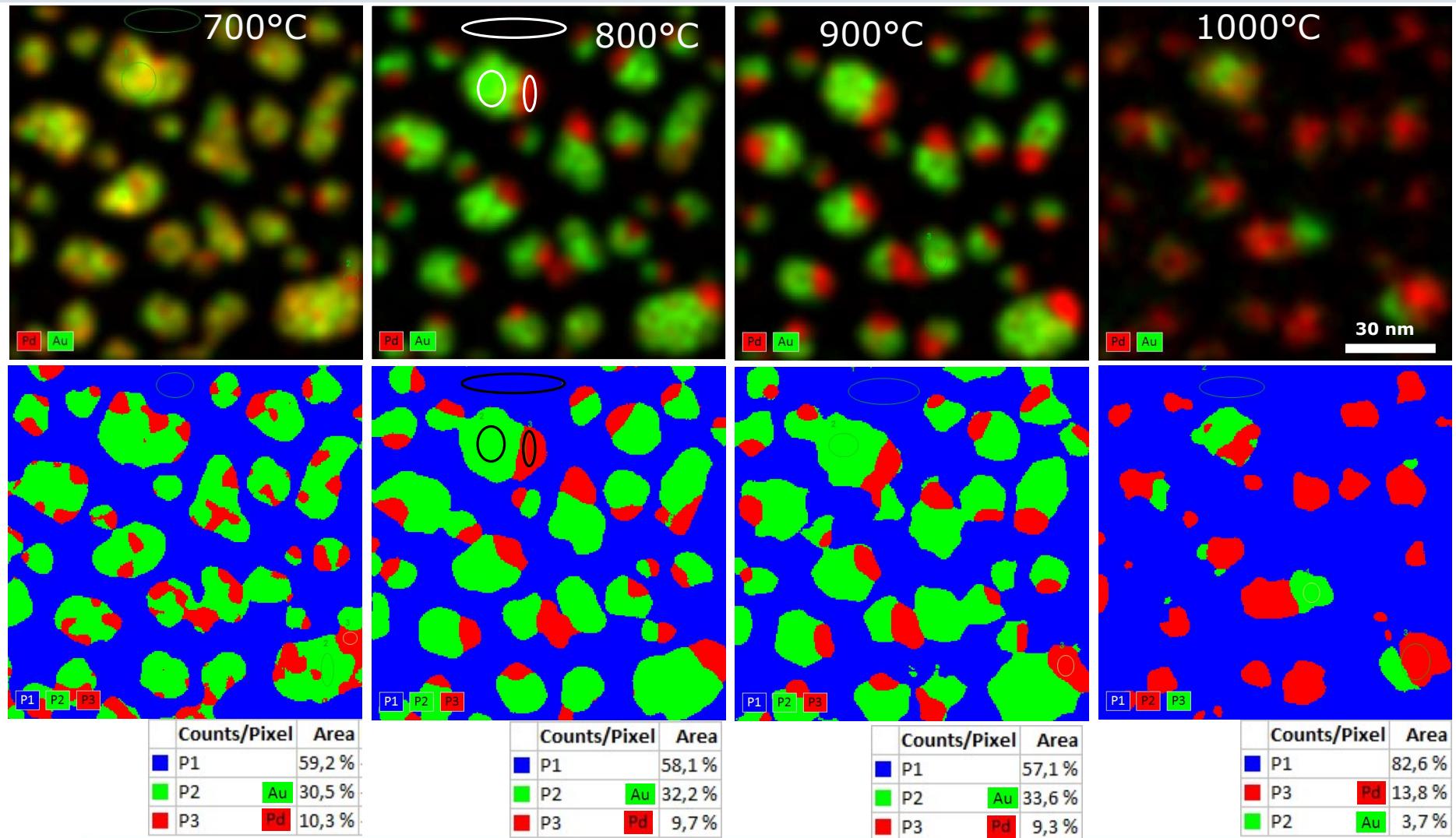
<sup>1</sup> Schoc  
UK

## Plunge-Freezing

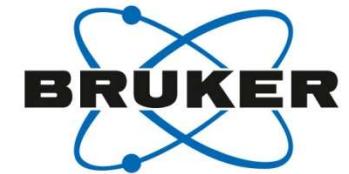


**Figure 3:** Cryo-EDX spectrum and corresponding EDX maps for BT dispersed in CCCM. Ca and P are present in the spectrum but through cryo analysis no visible Na and Mg peaks were present and the Cl peak was only just distinguishable. Analysis was carried out using a probe current of 40 pA.

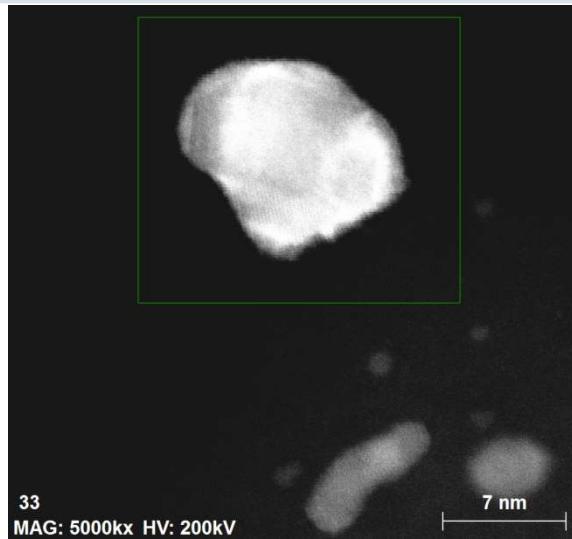
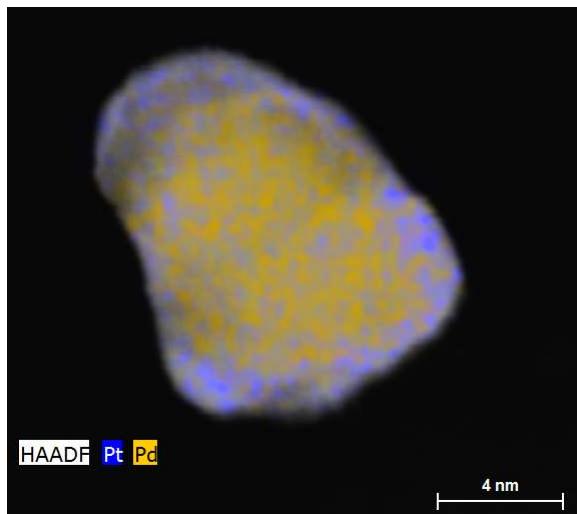
# Phase Analysis (PCA-based) and (*in situ*) monitoring of particles



# EDS for Catalysis, Quantification Pt-Pd Core Shell Particles

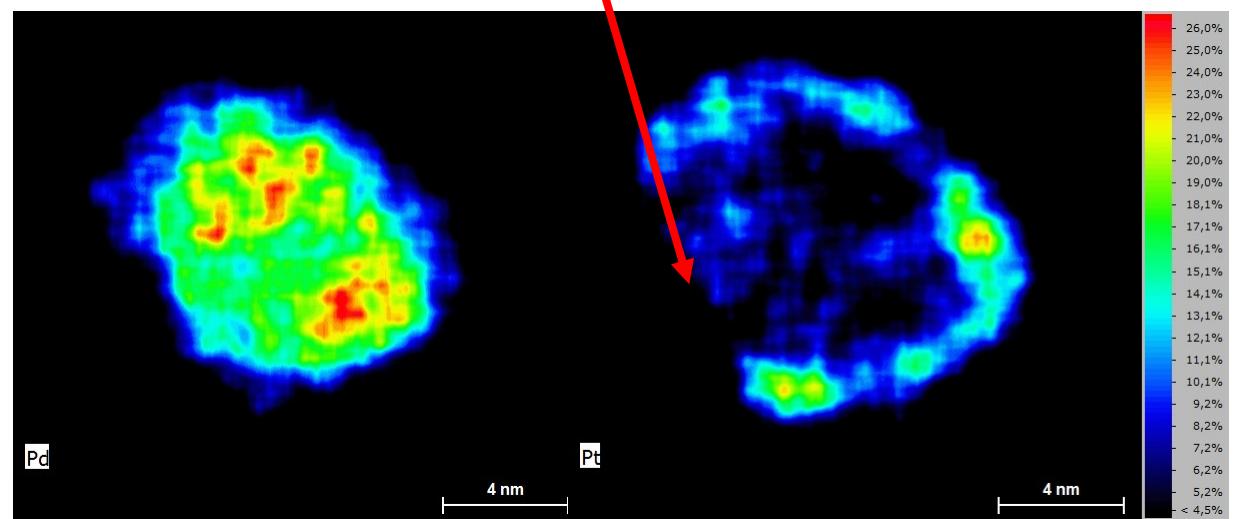
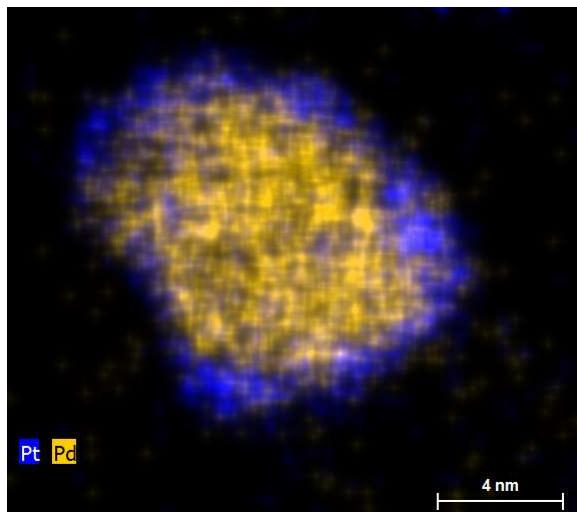


mass%, 30 mm<sup>2</sup>, 0.12 sr (Standard EDS); Cs-corr. STEM



Data courtesy: Dogan Ozkaya,  
Johnson Matthey  
Technology Center

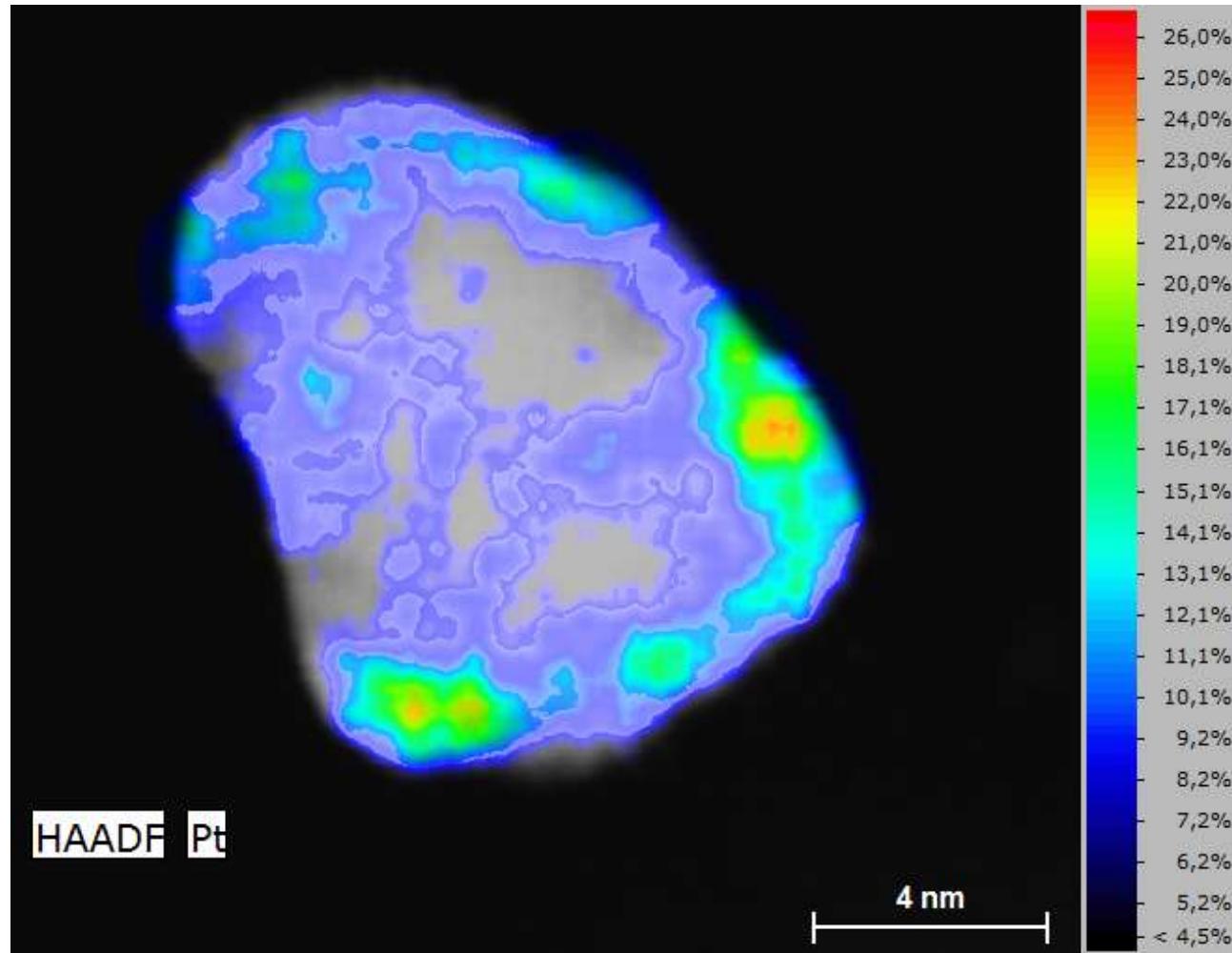
Pt shell not closed  
due to fabrication procedure



# EDS for Catalysis, Quantification Pt-Pd Core Shell Particles

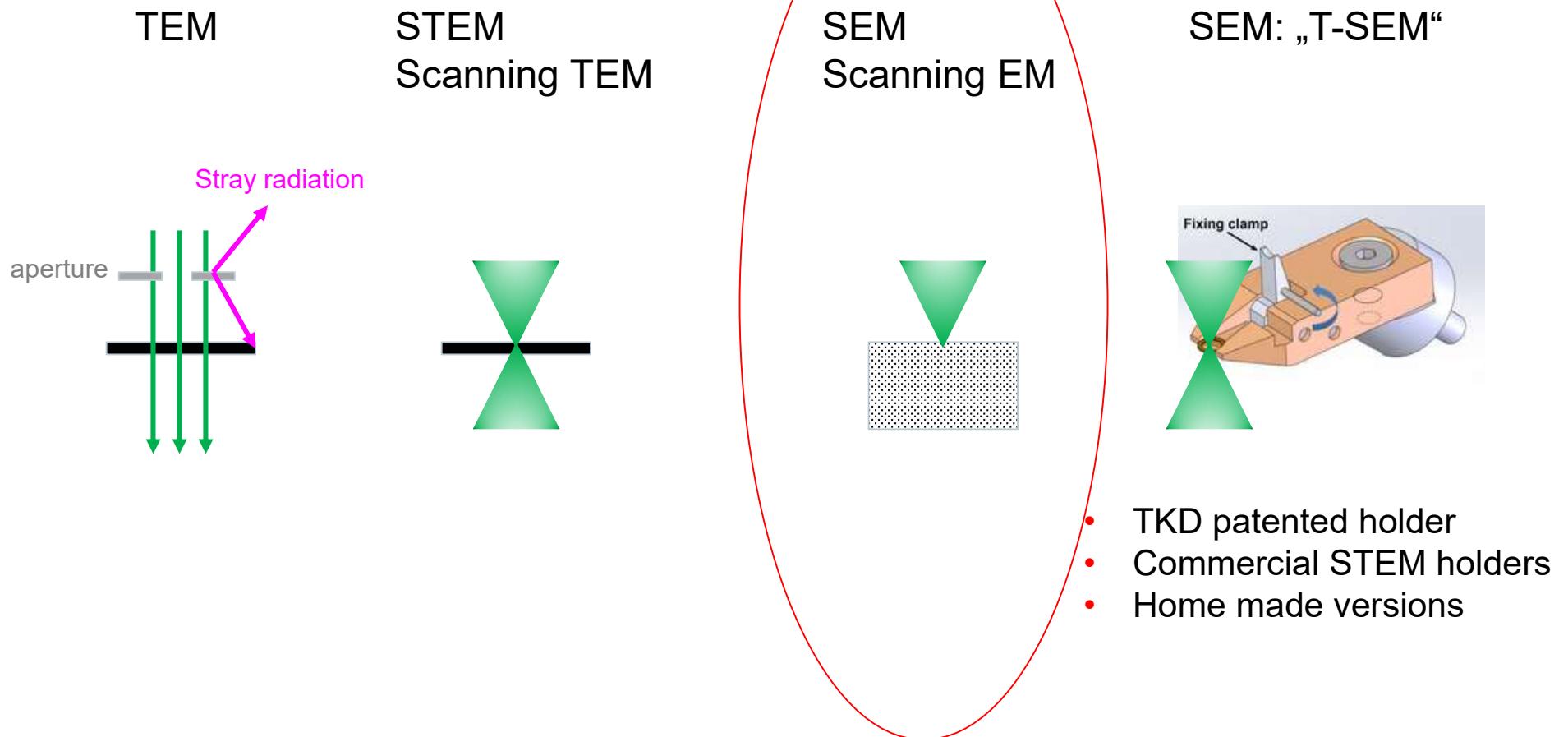


mass%, 30 mm<sup>2</sup>, 0.12 sr (Standard EDS); Cs-corr. STEM



Data courtesy:  
Dogan Ozkaya,  
Johnson Matthey  
Technology Center

# Electron Microscopy

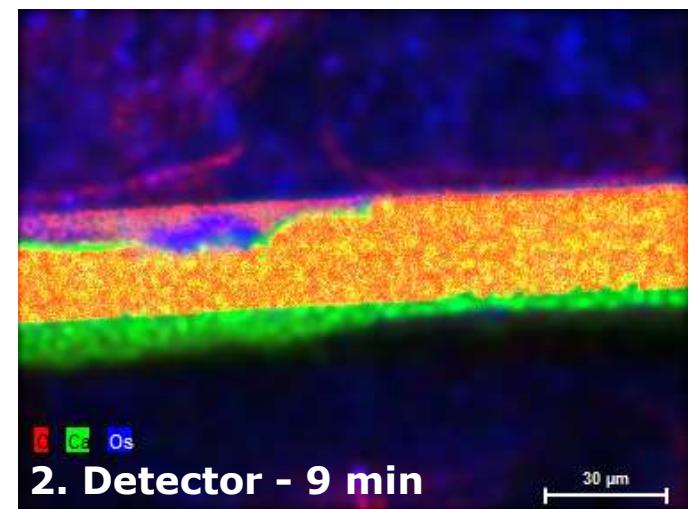
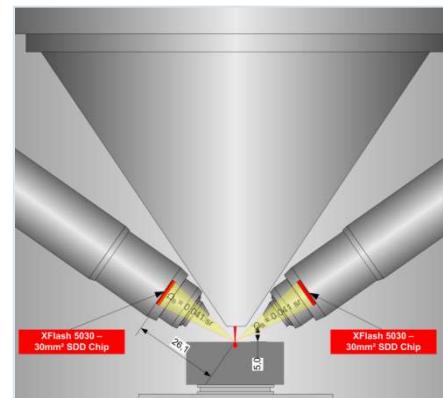
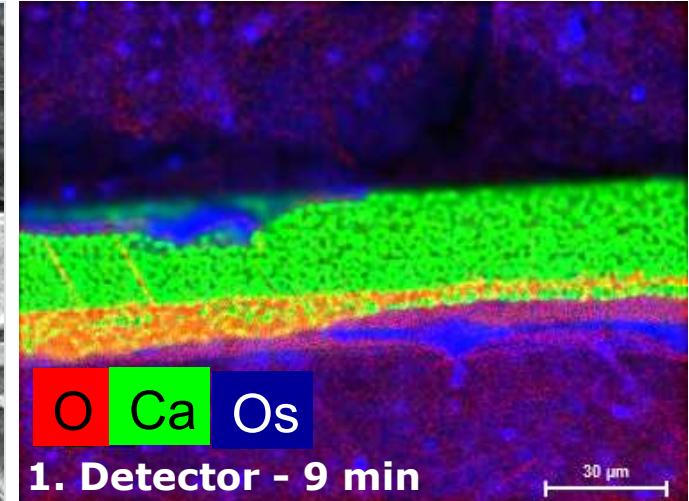
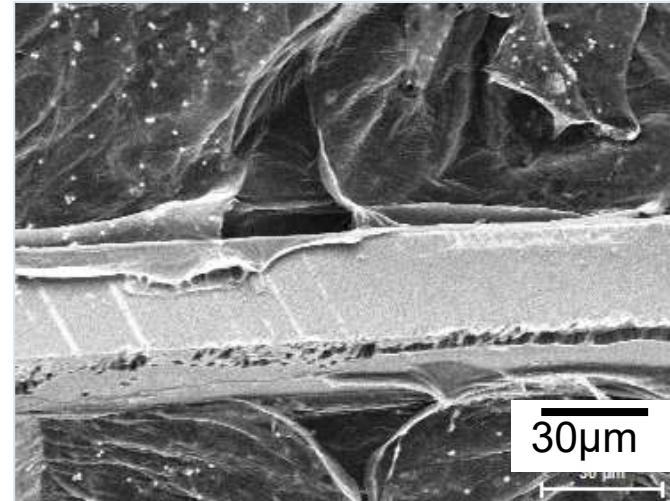


# Detector geometries for EDS Agave Leaf



Agave:

Ca-oxalate ( $\text{CaC}_2\text{O}_4$ )  
styloid crystal, iching



Analytical  
conditions:

- 8 kV
- ~0.5 nA
- ~3-5 kcps  
(each detector)
- 400 x 300 pixel

# Detector geometries for EDS Agave Leaf

LMU

BRUKER

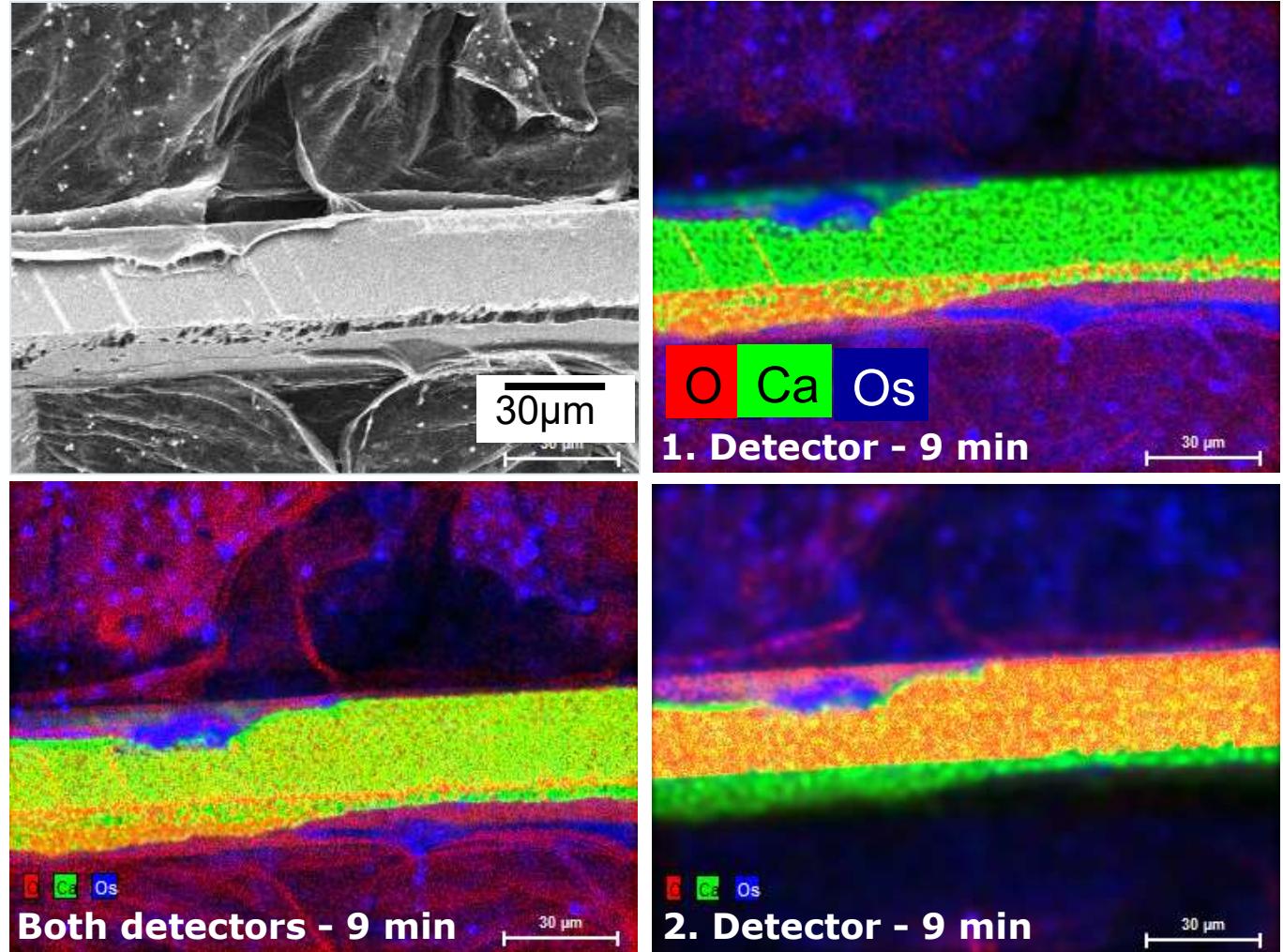


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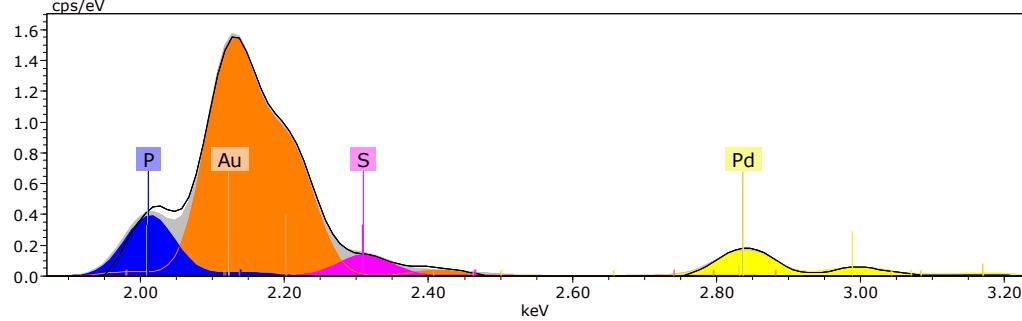
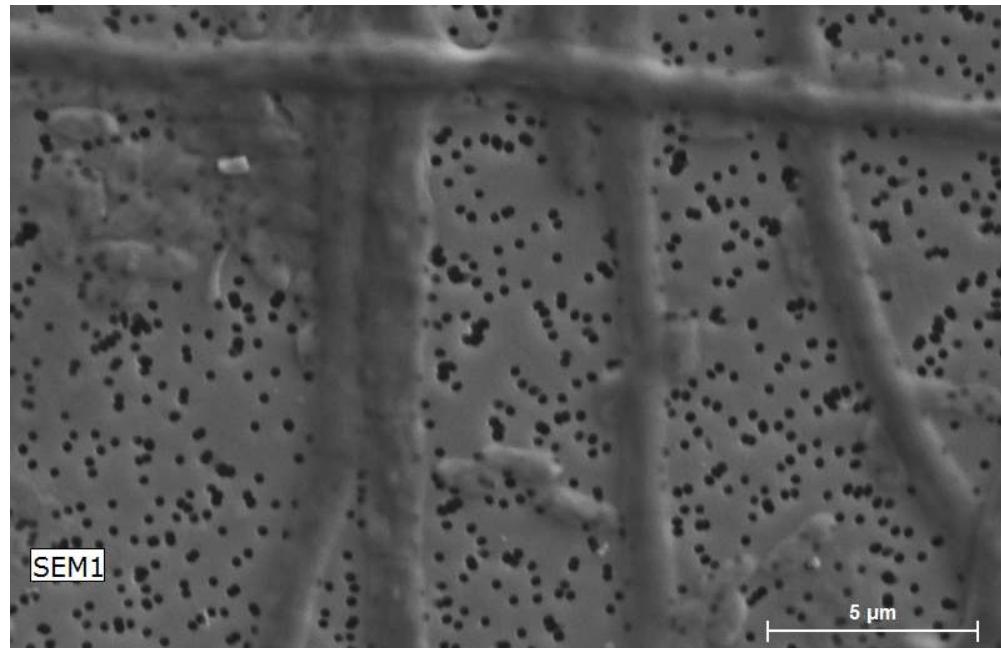
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- $\sim 0.5$  nA
- $\sim 3\text{-}5$  kcps  
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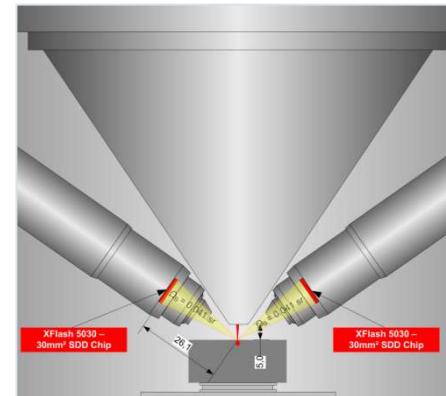
# Double detector system

## Cell analysis: *Beggiatoa alba* on Au/Pd coated polycarbonate filter

2x30mm<sup>2</sup>, 5 kV, 415 pA, 12 kcps, 86 min



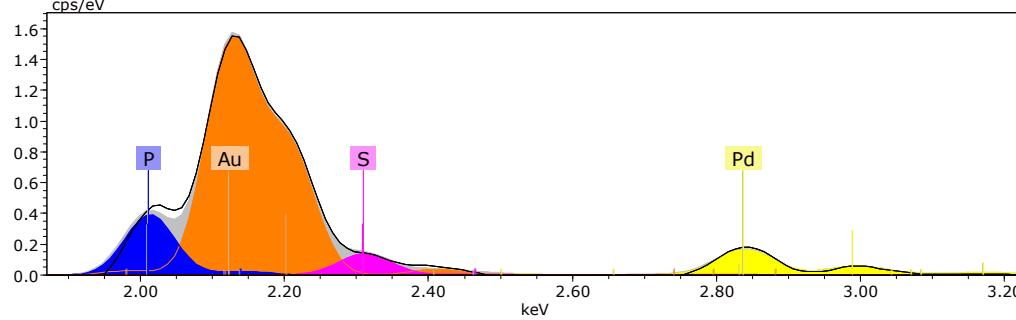
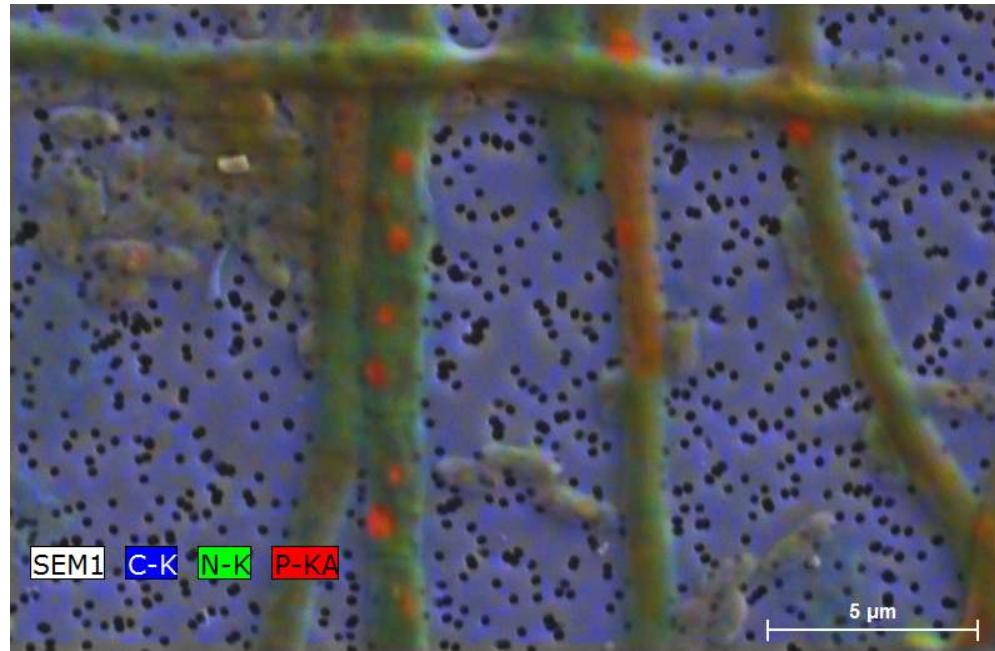
- Accumulation of nitrate in massive vacuoles
- Oxidize organics to CO<sub>2</sub> for biosynthesis
- Store sulfur
- Peak deconvolution reveals distribution of S and P



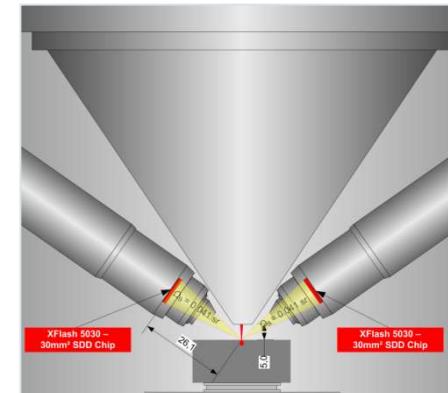
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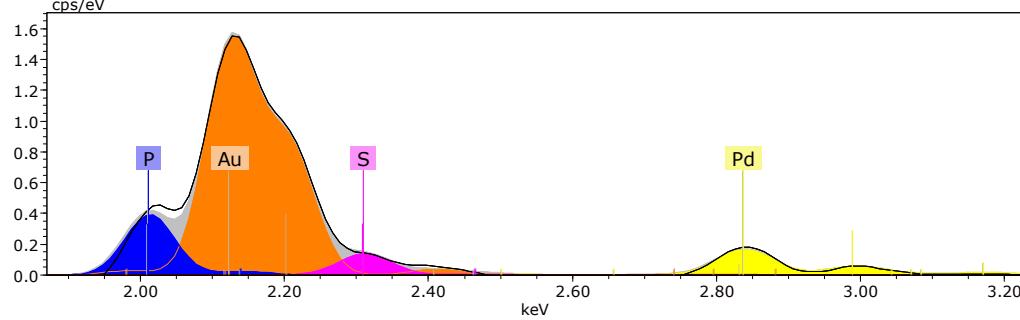
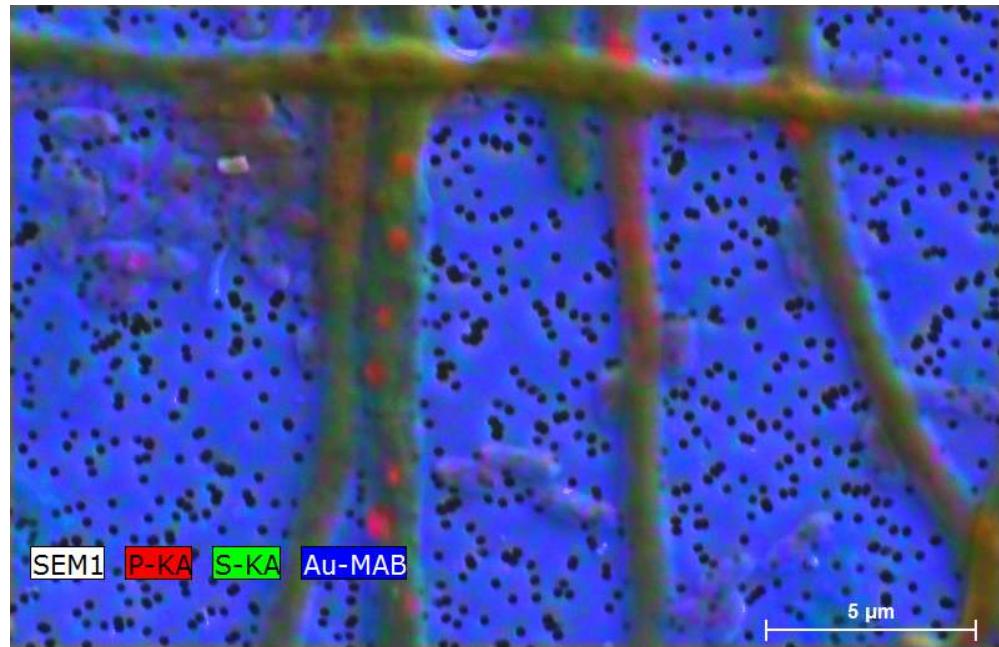
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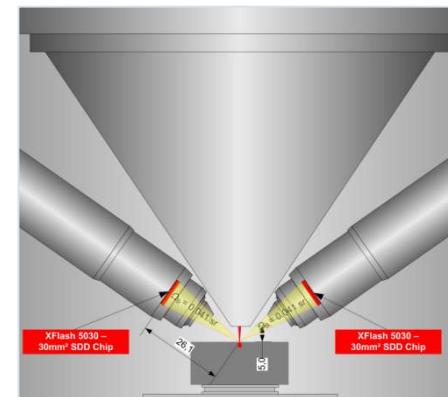
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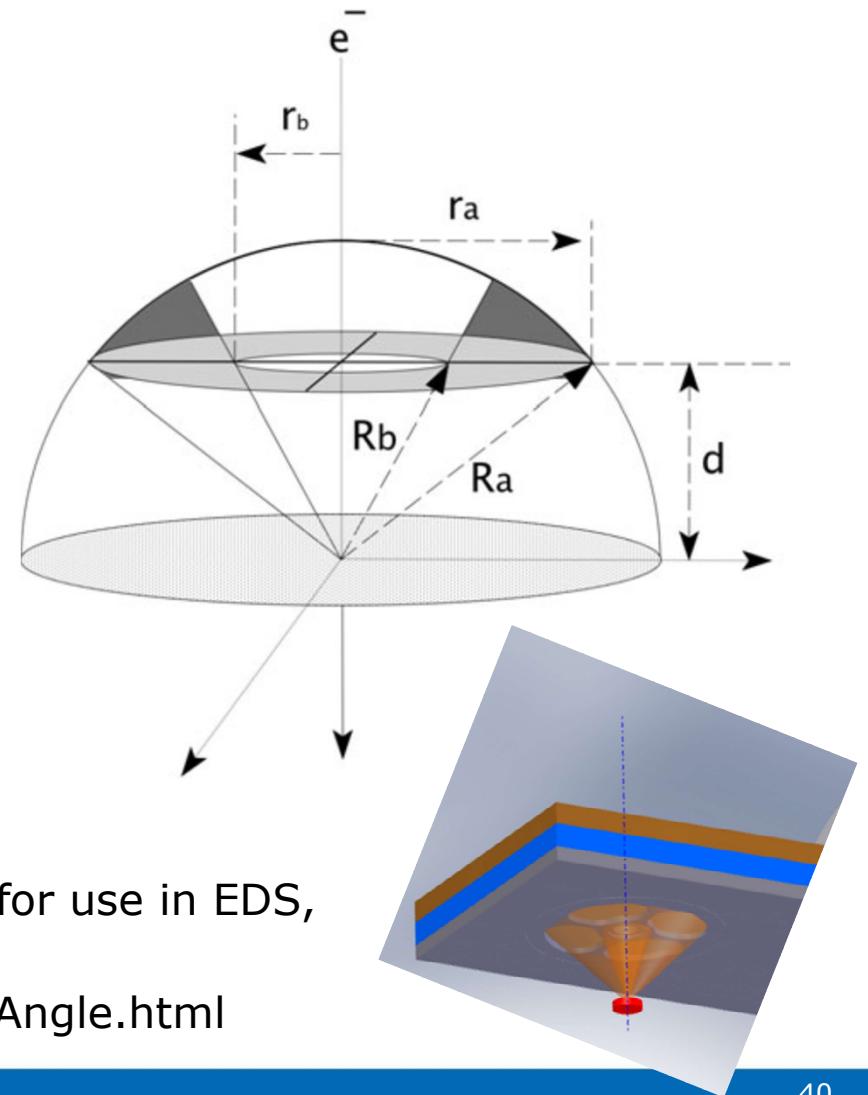
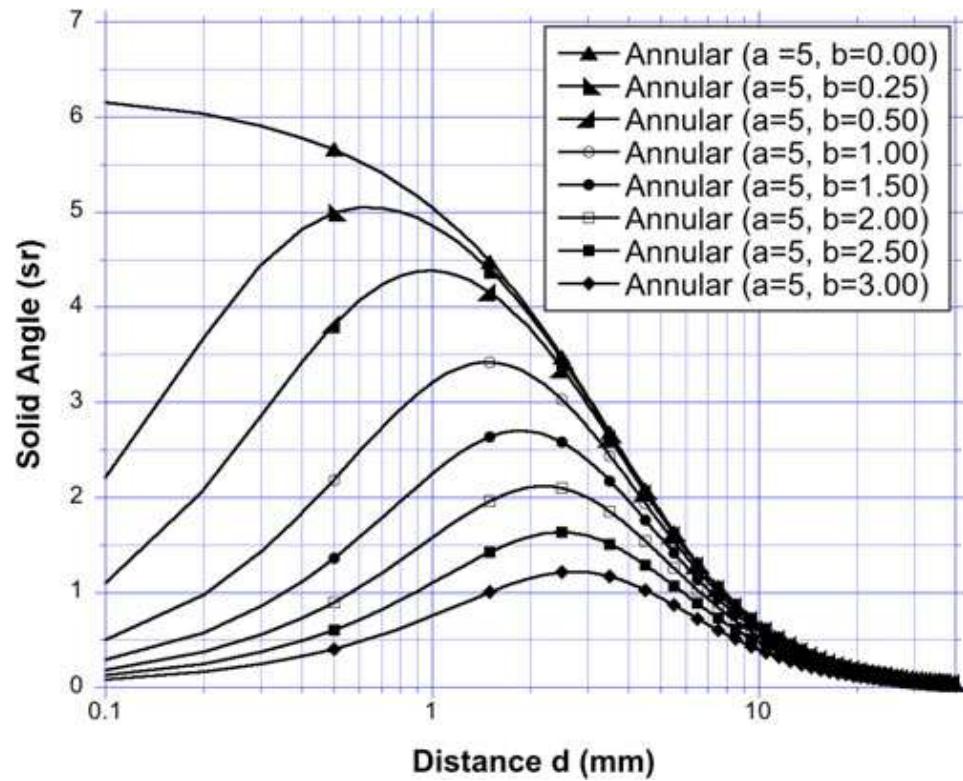
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# Annular EDS for T-SEM Flat Quad XFlash® 5060FQ



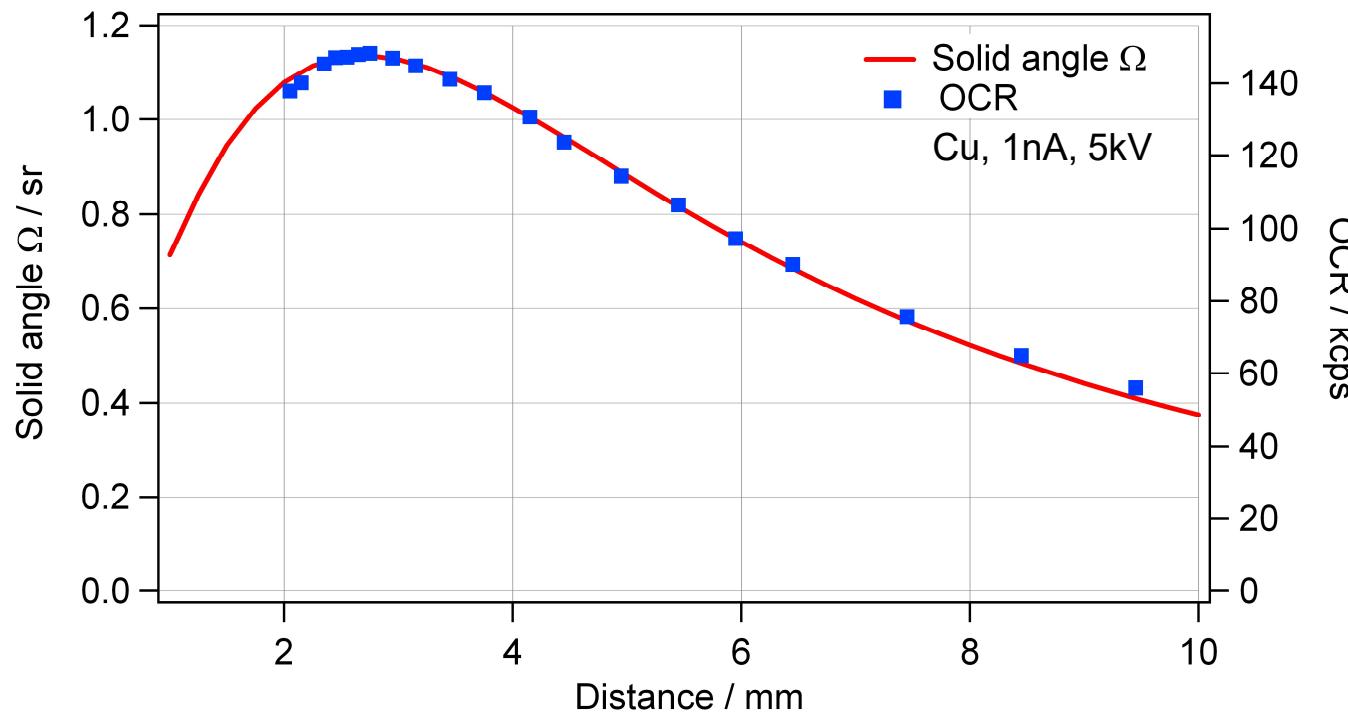
N. J. Zaluzec, Detector solid angle Formulas for use in EDS,  
Microsc. Microanal., 15 (2009) 93

<http://tpm.amc.anl.gov/NJZTools/XEDSSolidAngle.html>

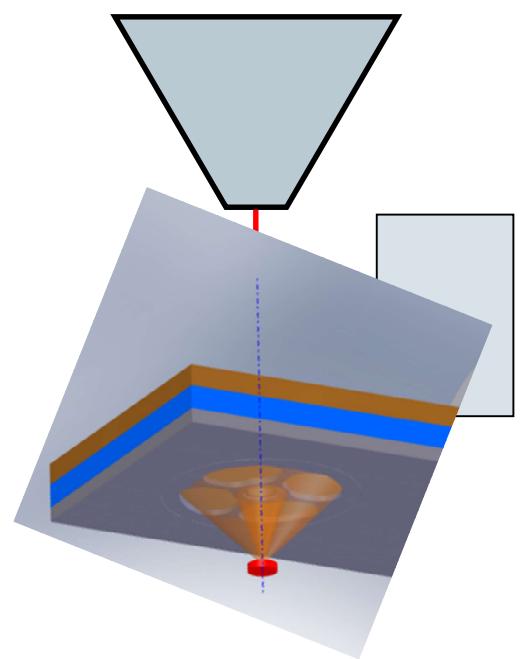
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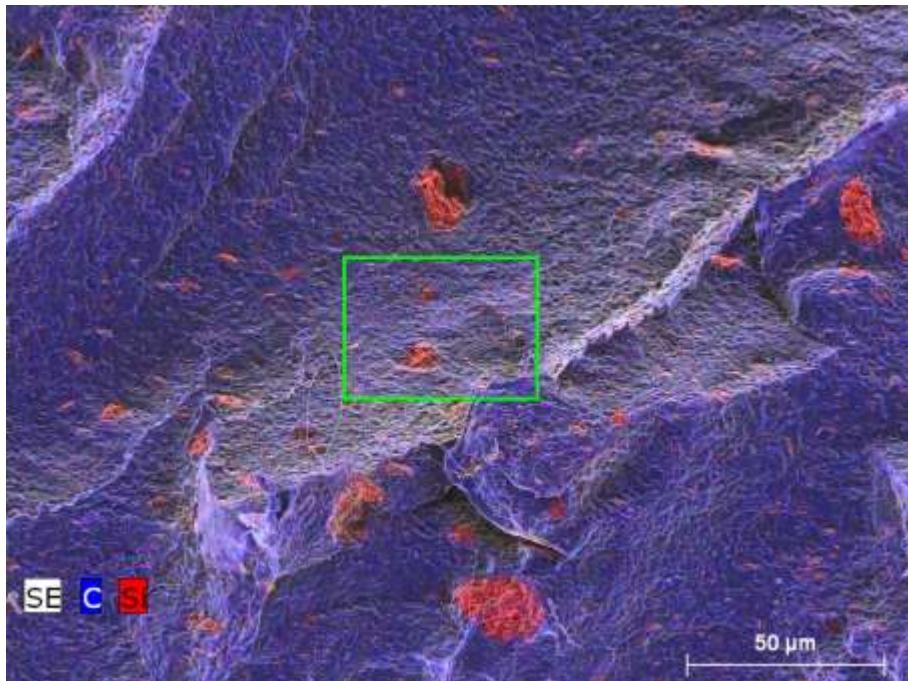
## Solid angle and OCR vs distance d



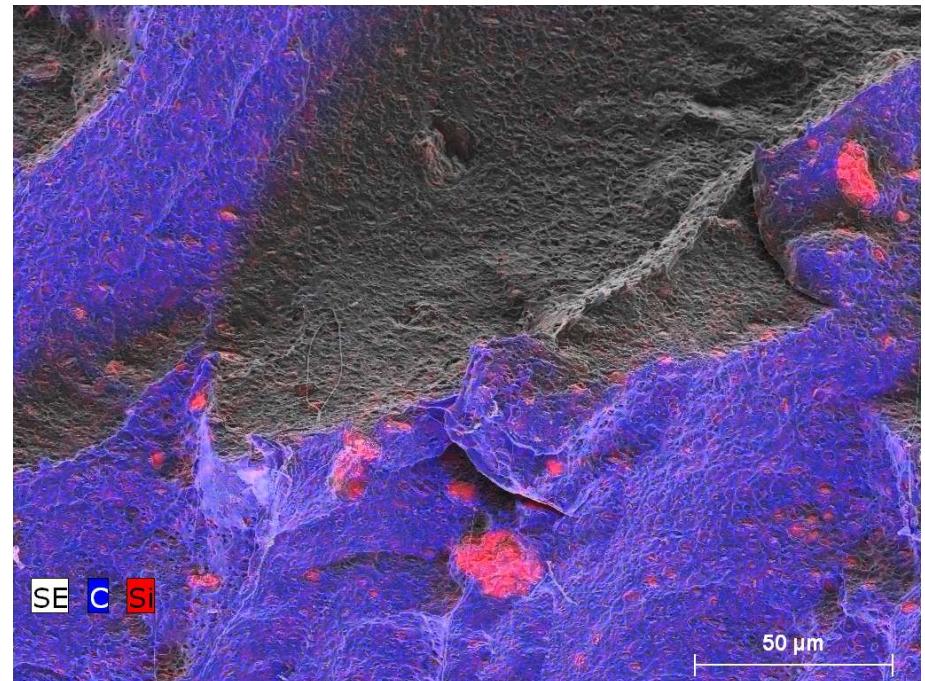
max solid angle at  
 $d = 2.5\text{mm}:$   
 $\Omega > 1.1 \text{ sr}$



# XFlash QUAD vs Single detector in SEM: Polymer composite containing organo clay



**XFlash Flat QUAD detector**  
3 kV, 220pA, 10 kcps,  
320 s, 1024x768 pixel

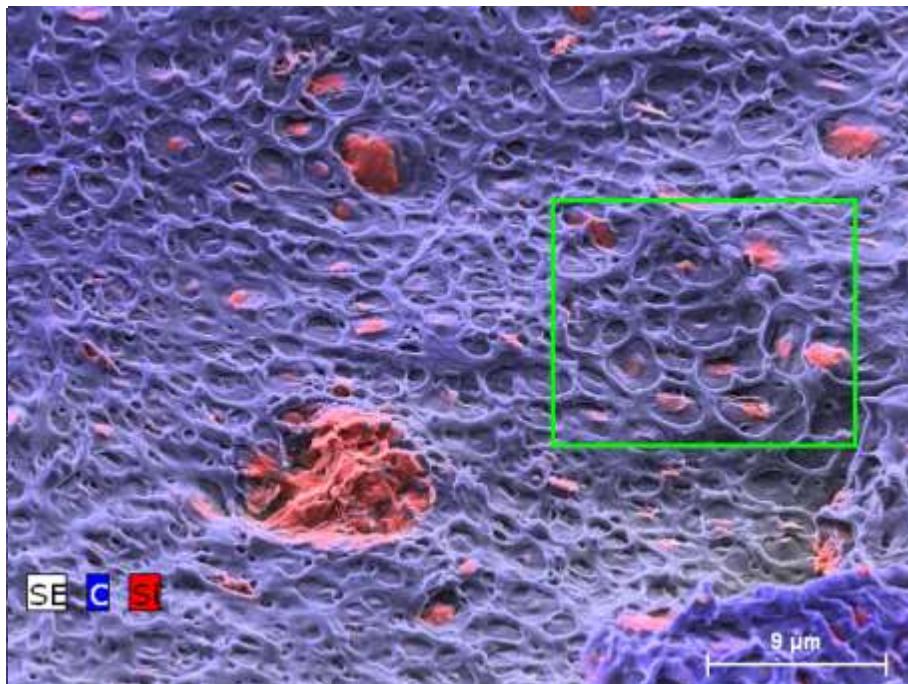


**Single 30mm<sup>2</sup> XFlash**  
3 kV, 220pA, 0.8 kcps,  
320 s, 1024x768 pixel  
Shadow effects due to rough surface

Sample courtesy by Dalto et al., Universidade Federal do Rio de Janeiro,  
Data courtesy T. Salge (Bruker / NHM, London);

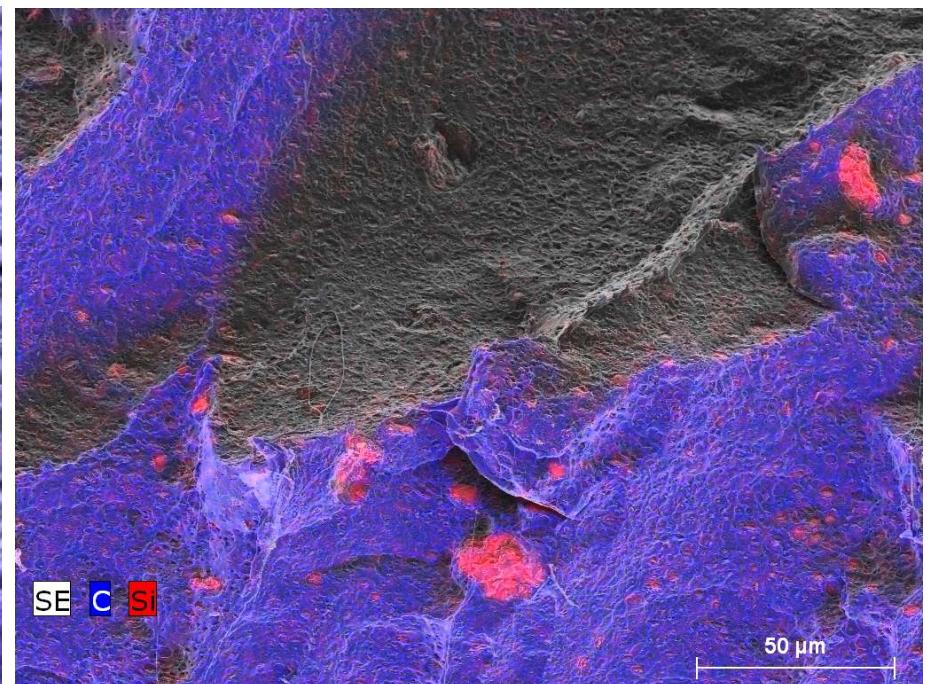
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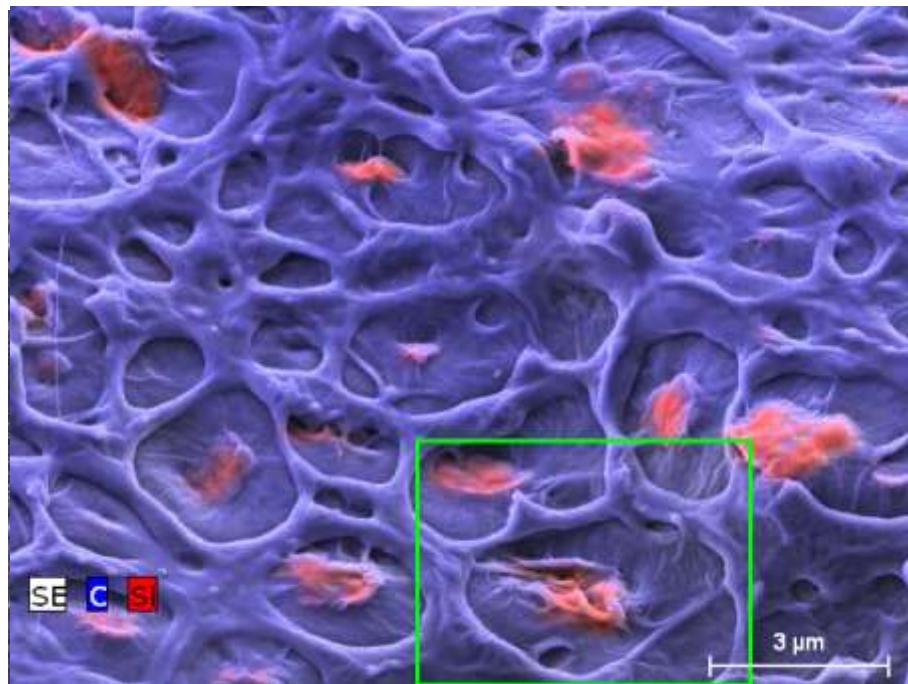
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Sample courtesy by Dalto et al., Universidade Federal do Rio de Janeiro,  
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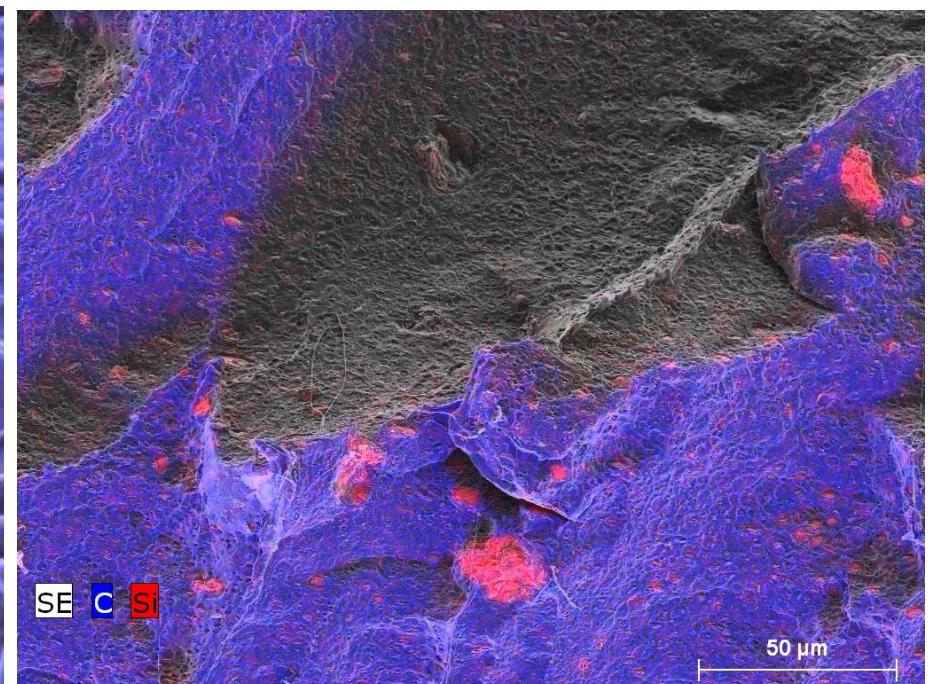
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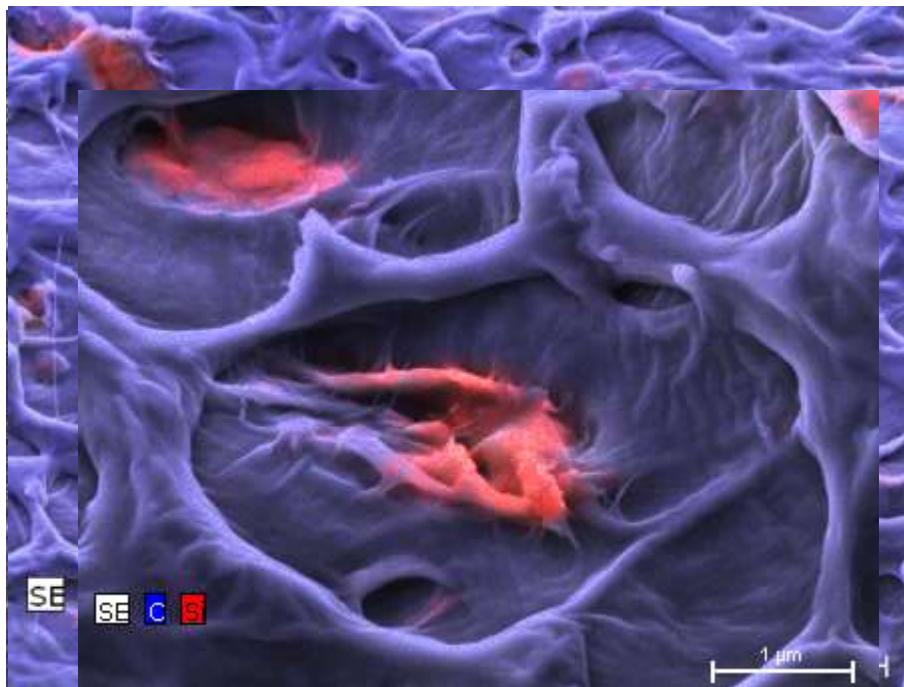
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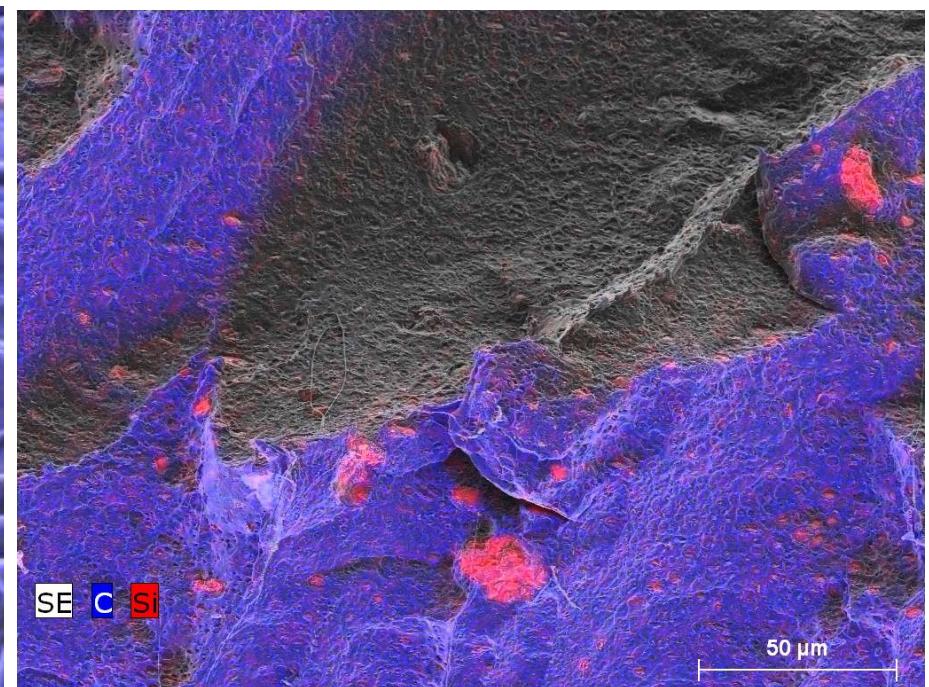
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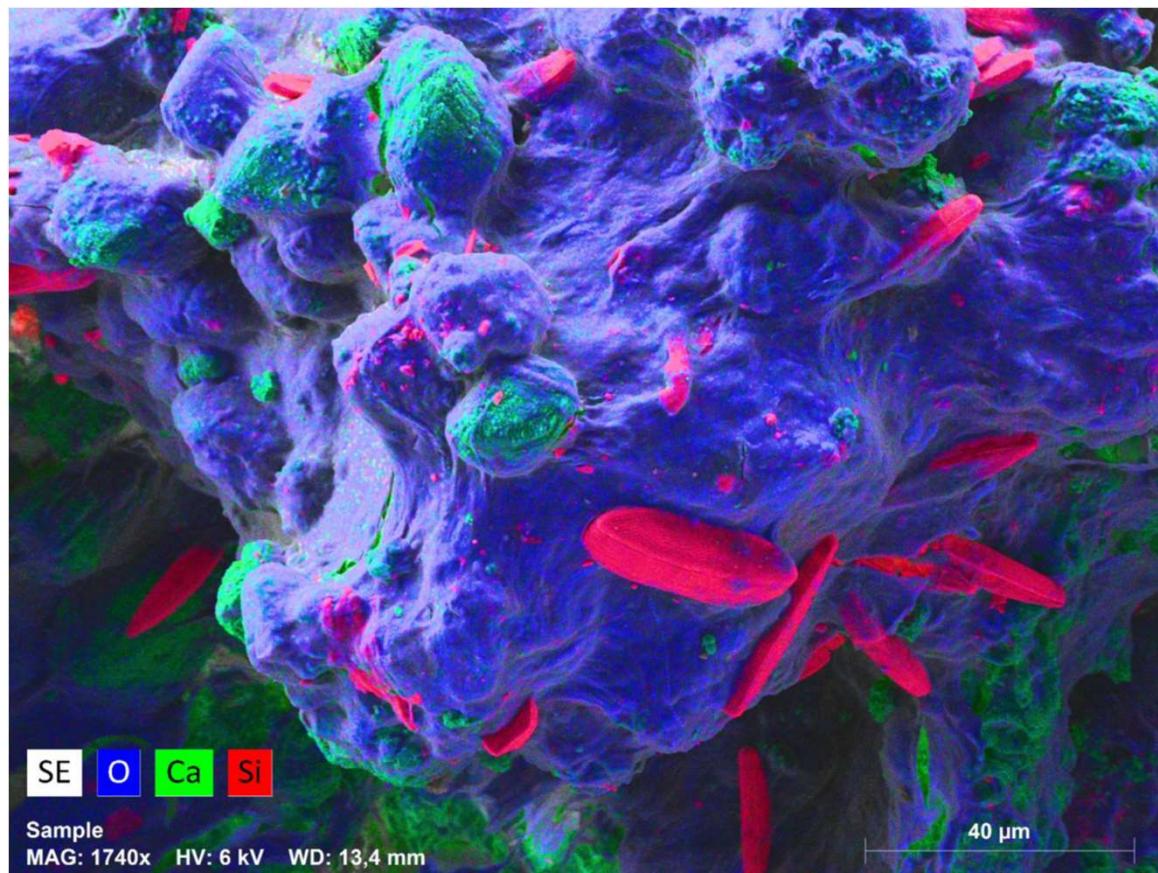


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# Microbial mat with diatoms, Specimen taken from Hot spring



Reddish ovals are diatoms sticking to the bluish biofilm surface

Carbonates (green) are embedded within the biofilm matrix

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Carbonates (green) are embedded within the biofilm matrix

# Low vacuum analysis Parasitoid wasp *Monolexis fuscicornis*



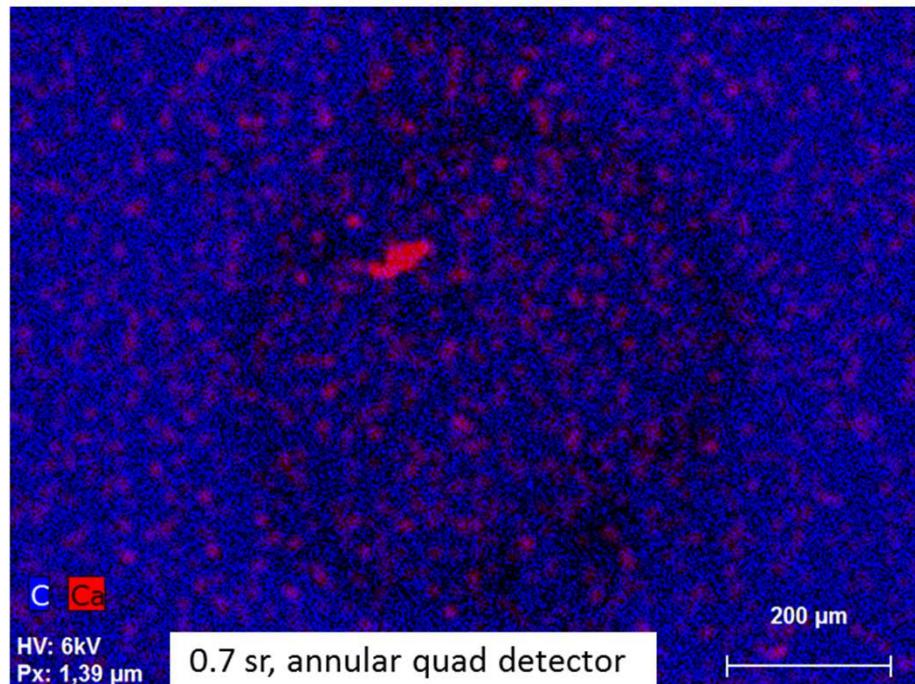
NATURAL  
HISTORY  
MUSEUM

BRUKER

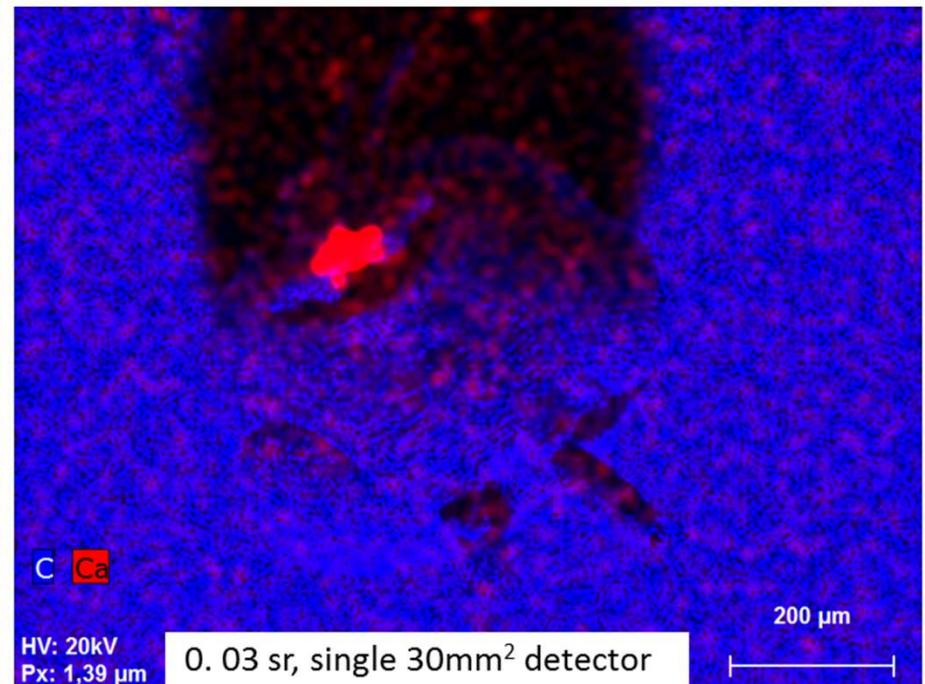
1.2sr

Wasps head with tooth

Flat Quad (**annular**)



**Single** 30mm<sup>2</sup> detector



Low vacuum (20 Pa), 6kV, 240 sec, 800 x 600 pixel

# Low vacuum analysis Parasitoid wasp *Monolexis fuscicornis*



NATURAL  
HISTORY  
MUSEUM

BRUKER

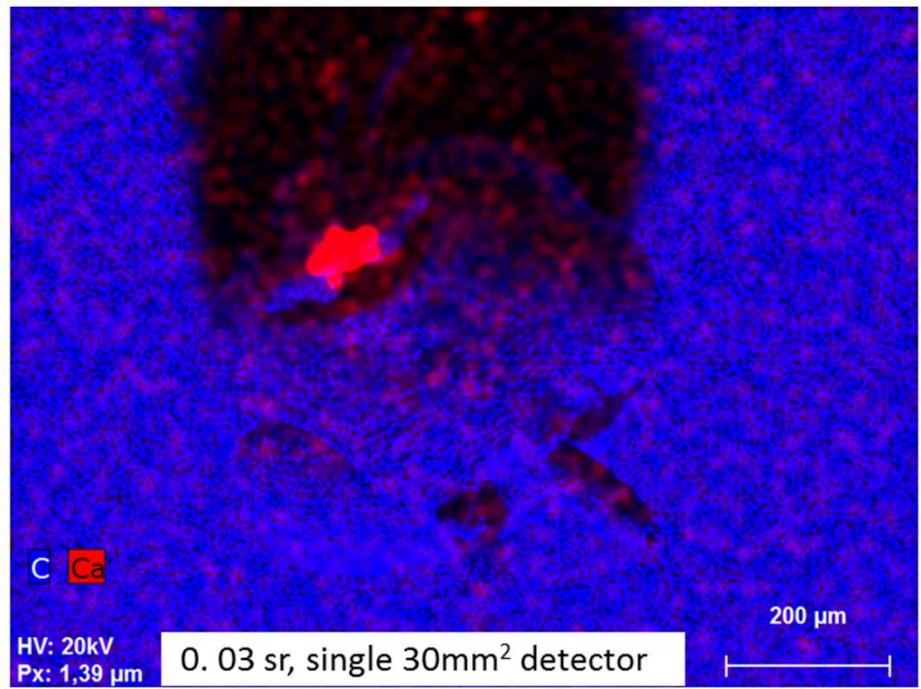
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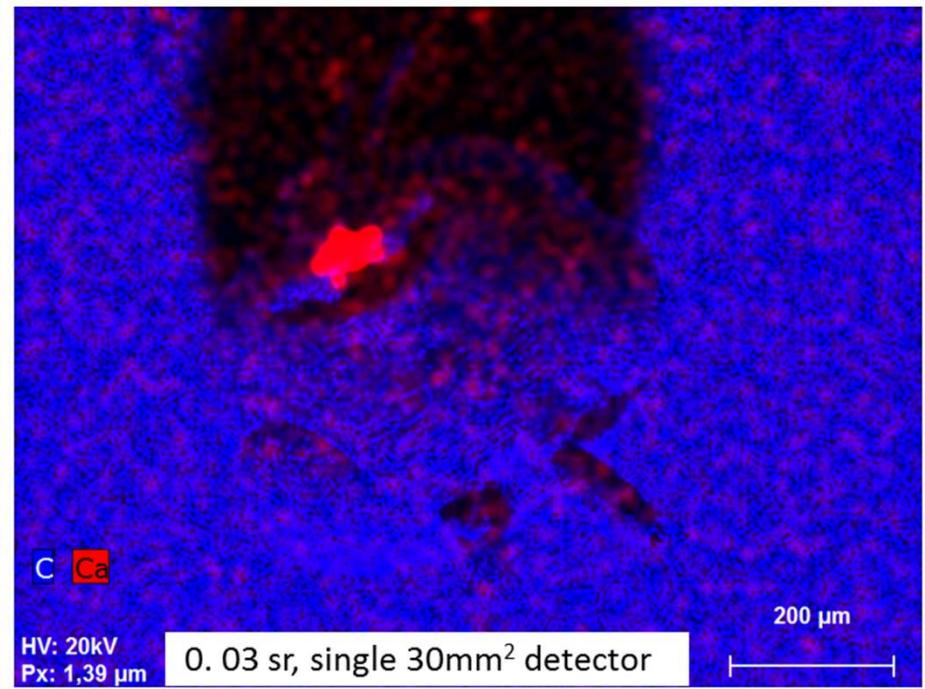
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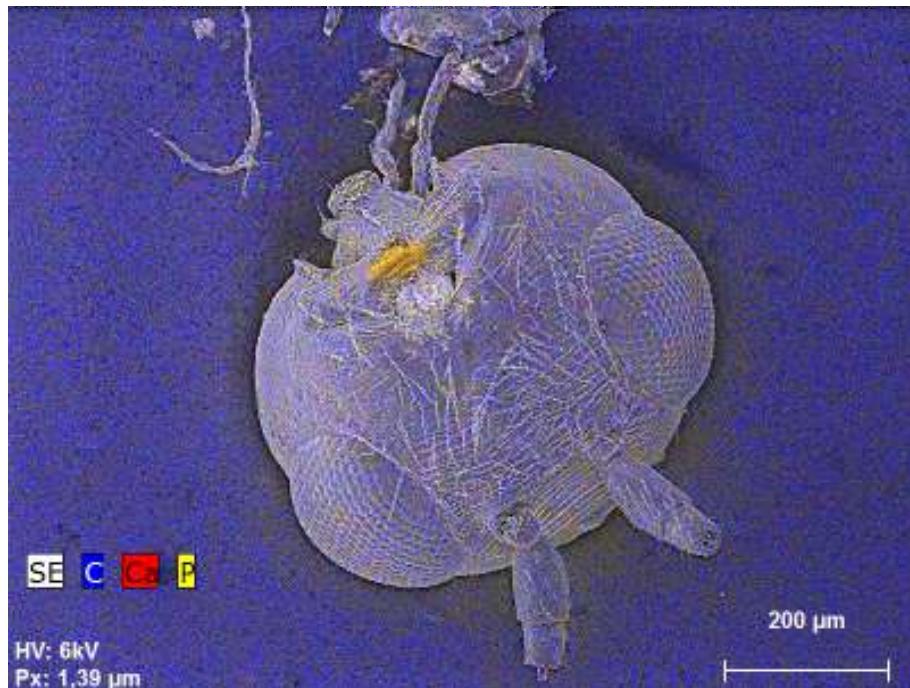


NATURAL  
HISTORY  
MUSEUM

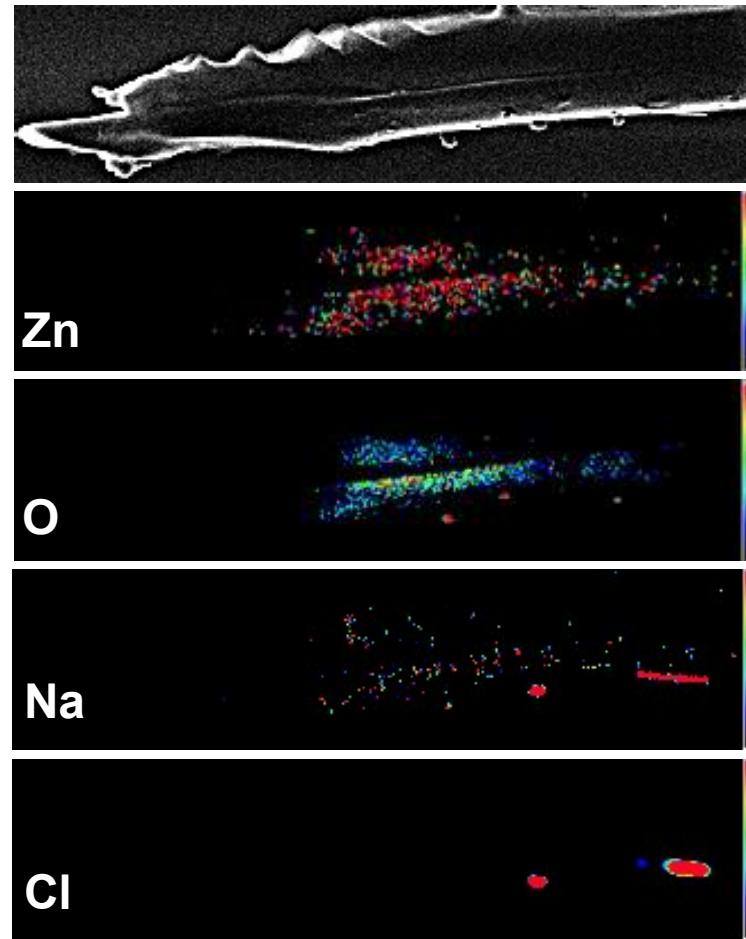
BRUKER

1.2sr

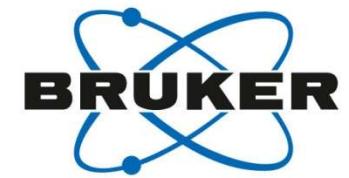
Head



Ovipositor (sting and egg-layer)

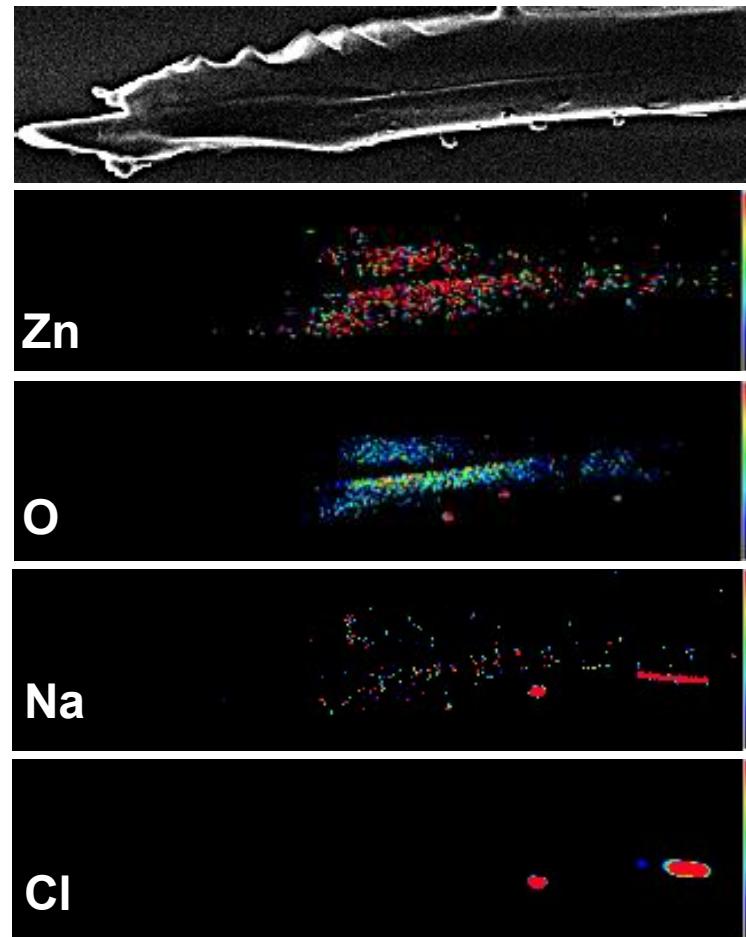
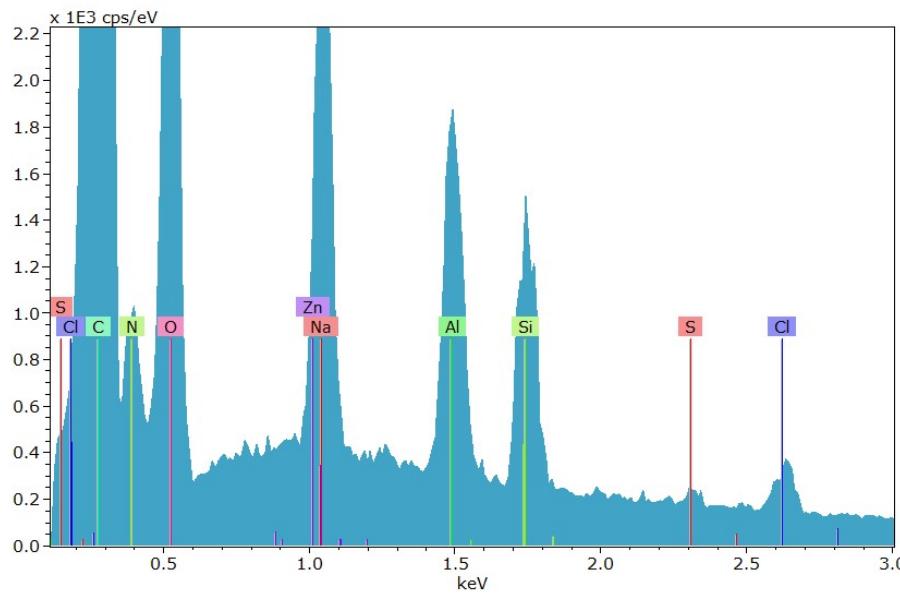


# Low vacuum analysis Parasitoid wasp (*Monolexis fuscicornis*)



## Ovipositor (sting and egg-layer)

- Low vacuum (20 Pa),  
5kV, 30 min, 320 x 240 pixel

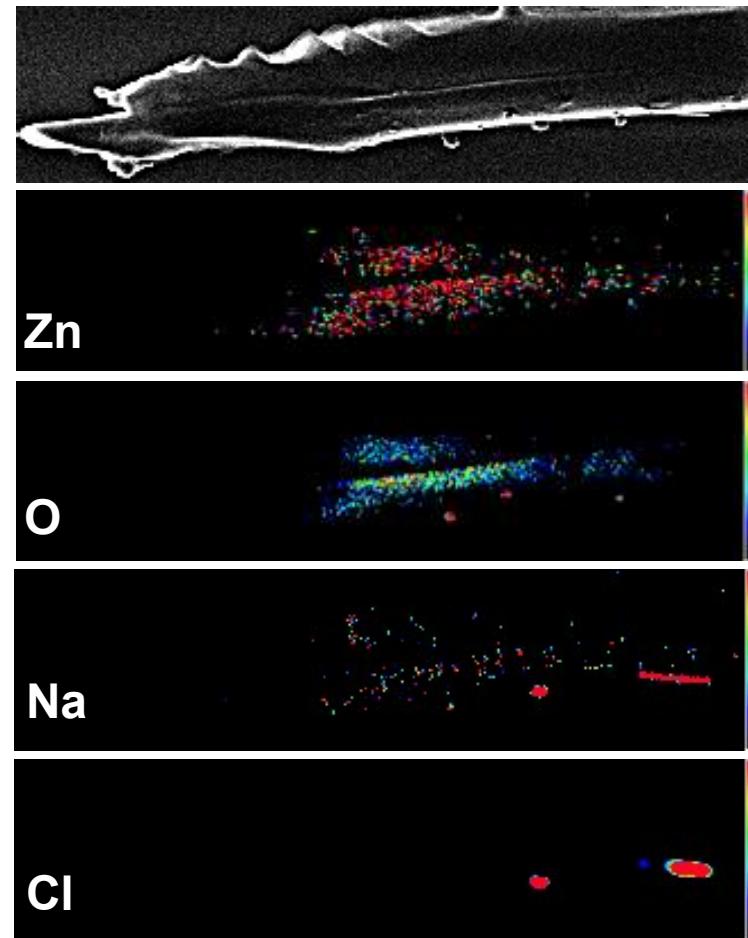
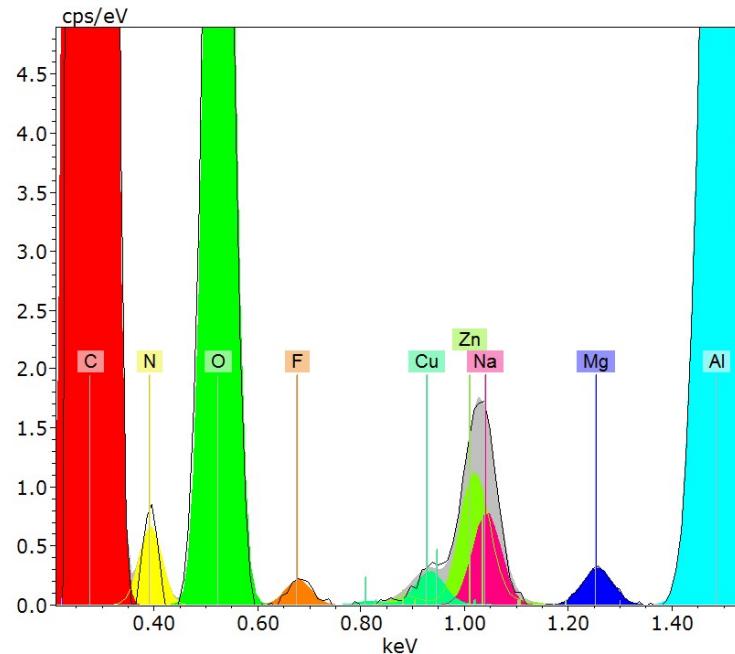


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In cooperation with: A. T. Kearsley & G. R. Broad (Natural History Museum, London)

# A Sea Urchins Spine Tuberclle: bio-mimetics / bio-inspired construction

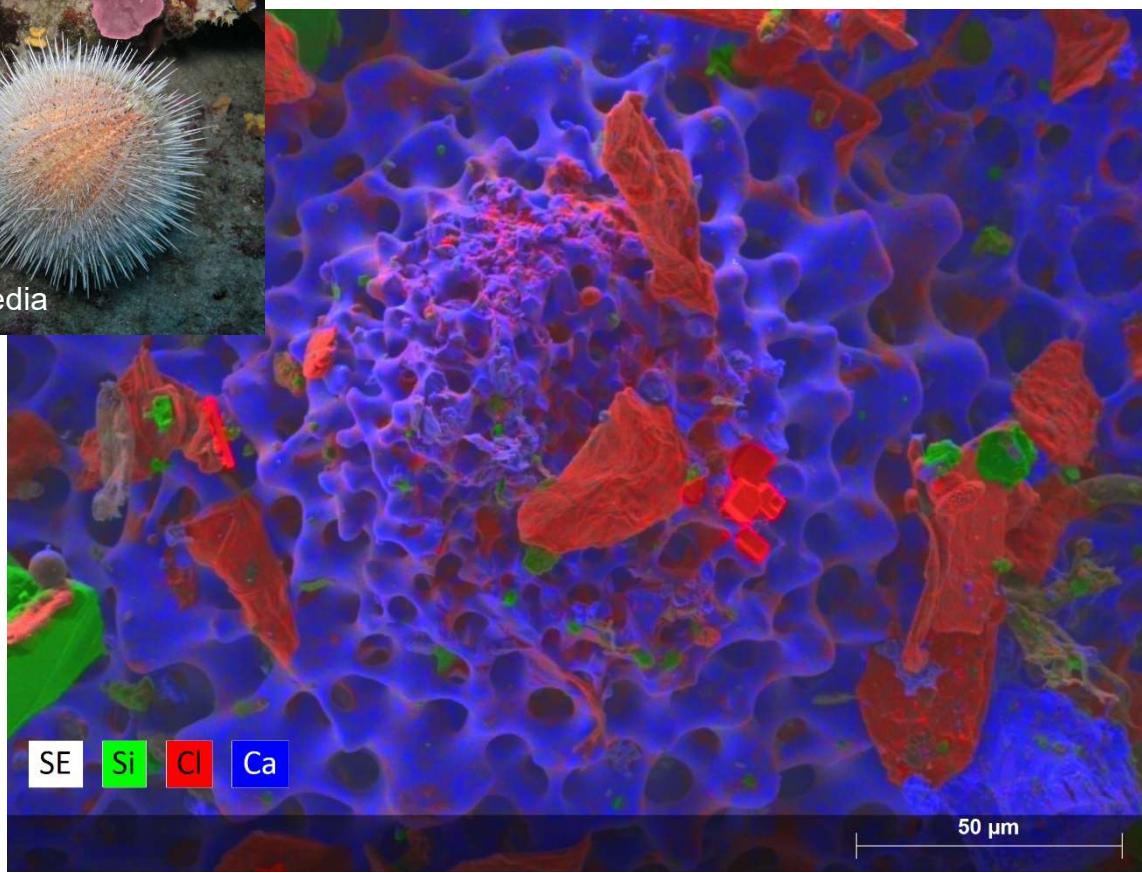
T. B. Grun, J. H. Nebelsick, PLOS, Sep. 27, 2018, <https://doi.org/10.1371/journal.pone.0204432>



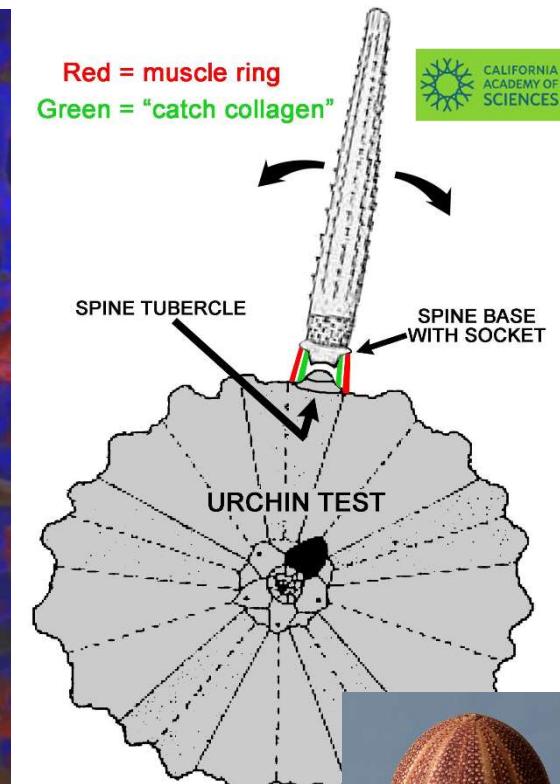
Lightweight hierarchical construction ensures stiffness, heavy load bearing and flexibility



wikipedia



Red = muscle ring  
Green = "catch collagen"



wikipedia

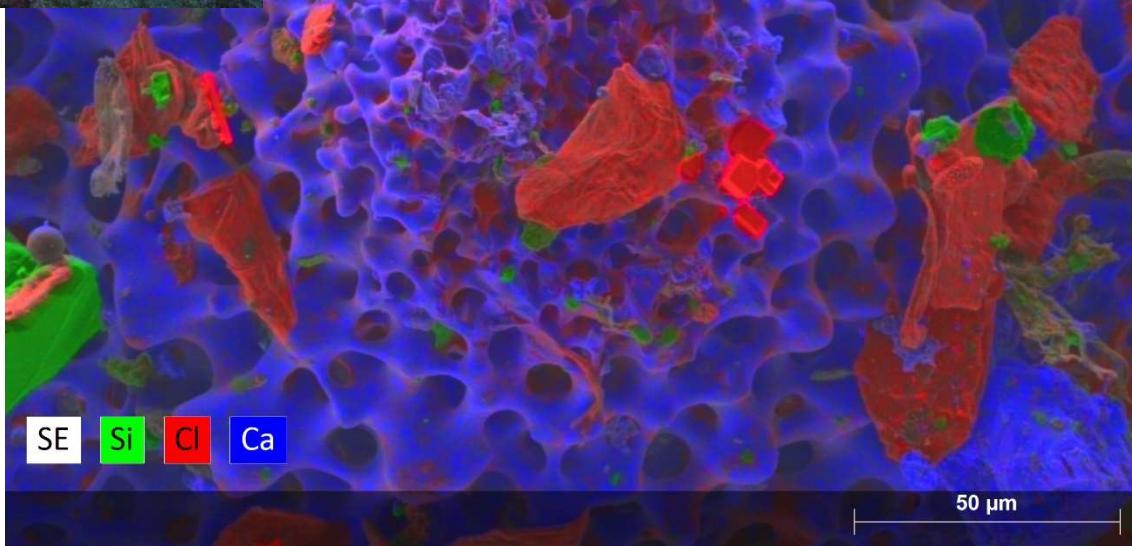
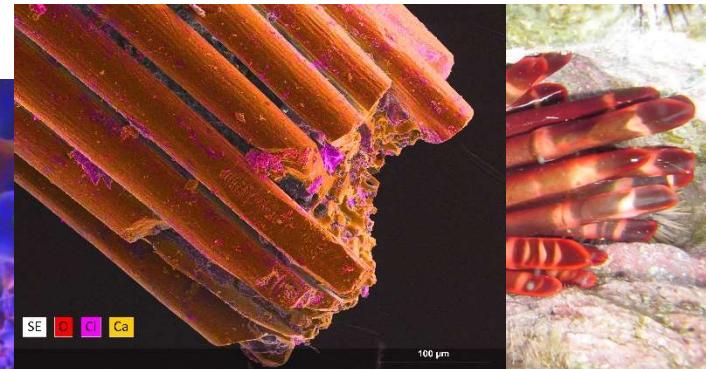
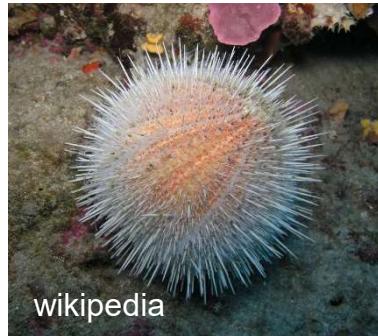
Data courtesy: M. Patzschke (Bruker)

# A Sea Urchins Spine Tuberclle: bio-mimetics / bio-inspired construction

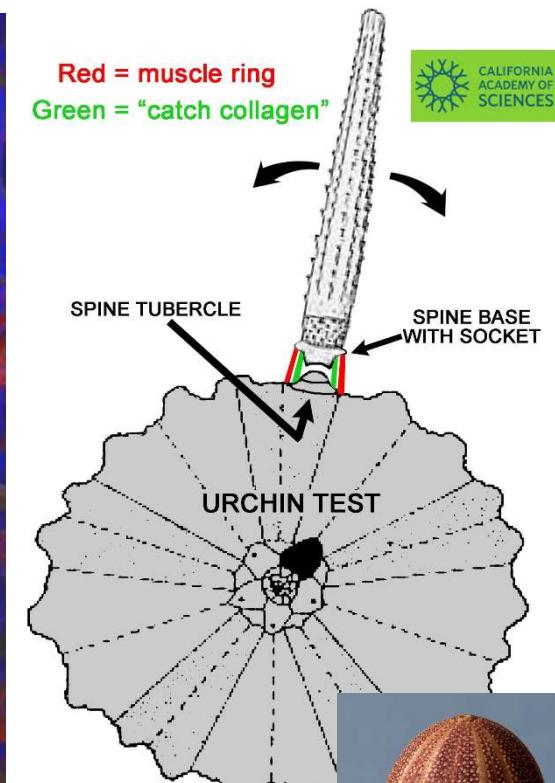


<https://www.calacademy.org/blogs/2011-philippine-biodiversity-expedition/when-it-comes-to-echinoderm-collagen-there-is-always-a>

Lightweight hierarchical construction ensures stiffness, heavy load bearing and flexibility



Red = muscle ring  
Green = "catch collagen"

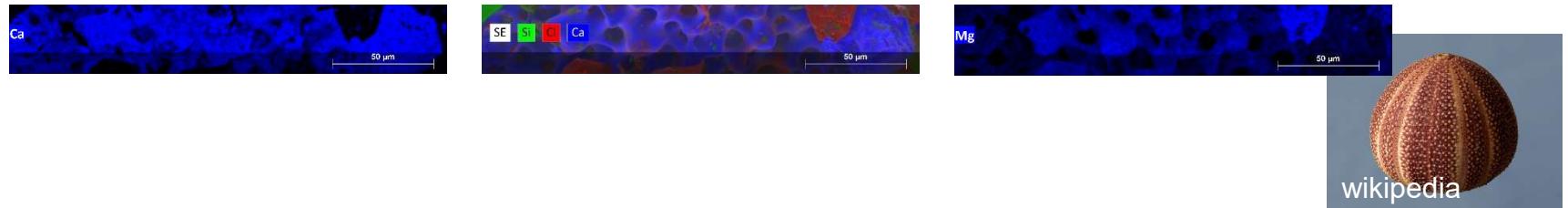
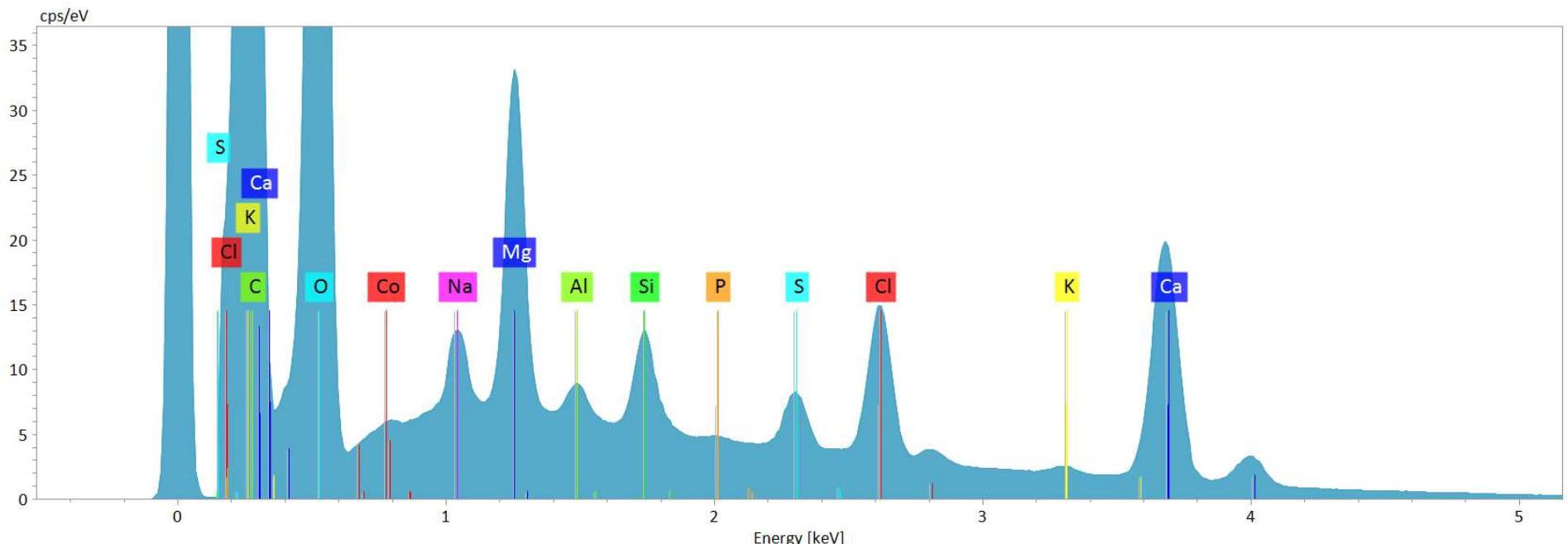


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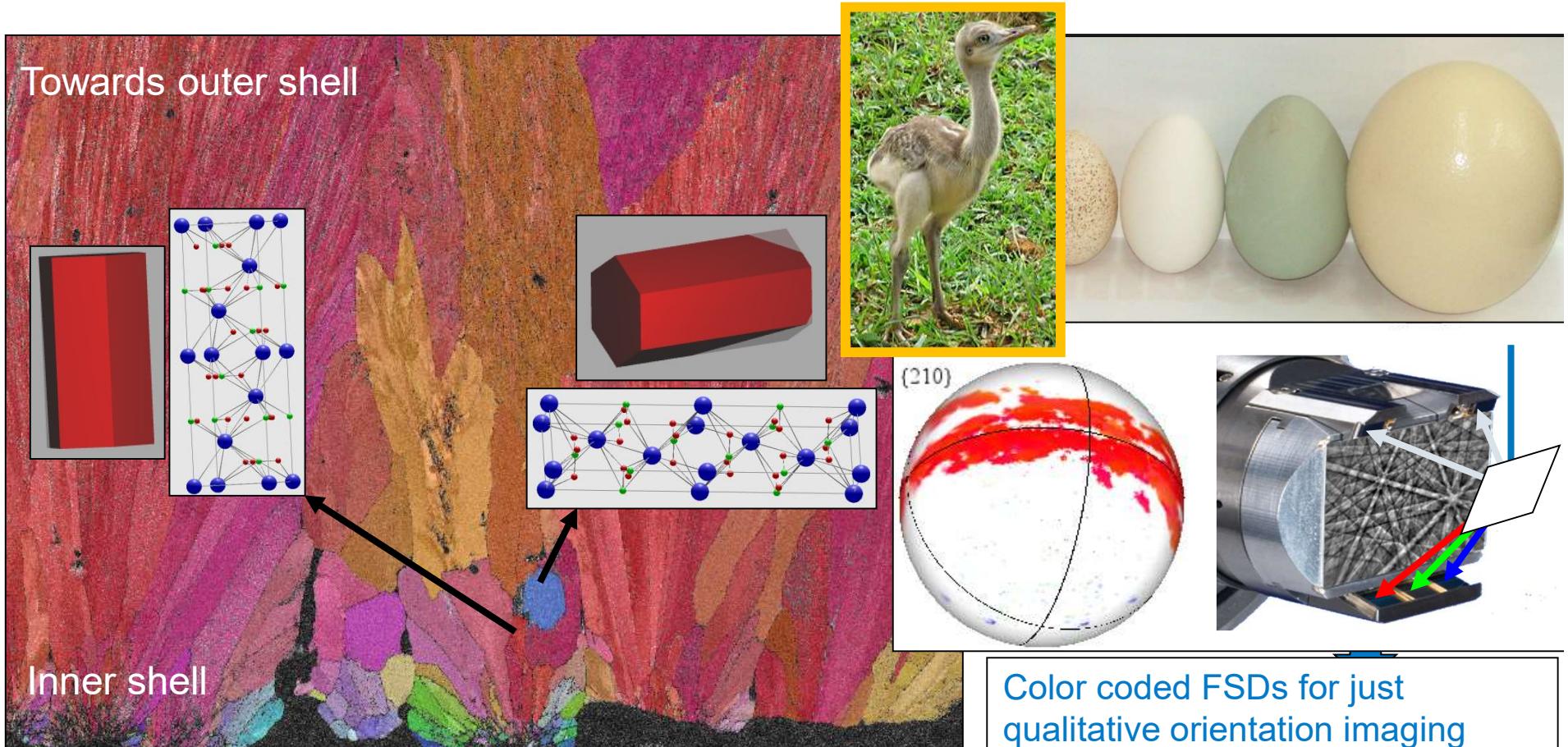


<https://www.calacademy.org/blogs/2011-philippine-biodiversity-expedition/when-it-comes-to-echinoderm-collagen-there-is-always-a>

Lightweight hierarchical construction ensures stiffness, heavy load bearing and flexibility



# EBSD for bio (bone, teeth, crustaceans,...): Egg shells; Ostrich: strongest texture but micro-cracks and disorder for chick to get out



Crystallographic orientation distribution map of the  
Cross section of an Ostrich egg shell (inner part)

# Understanding the early evolution of nervous systems including vision

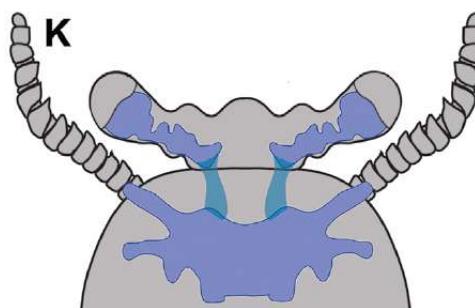


Ma et al., 2015, Current Biology 25, 2969–2975

<http://dx.doi.org/10.1016/j.cub.2015.09.063>

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<http://creativecommons.org/licenses/by-nc-nd/4.0/>

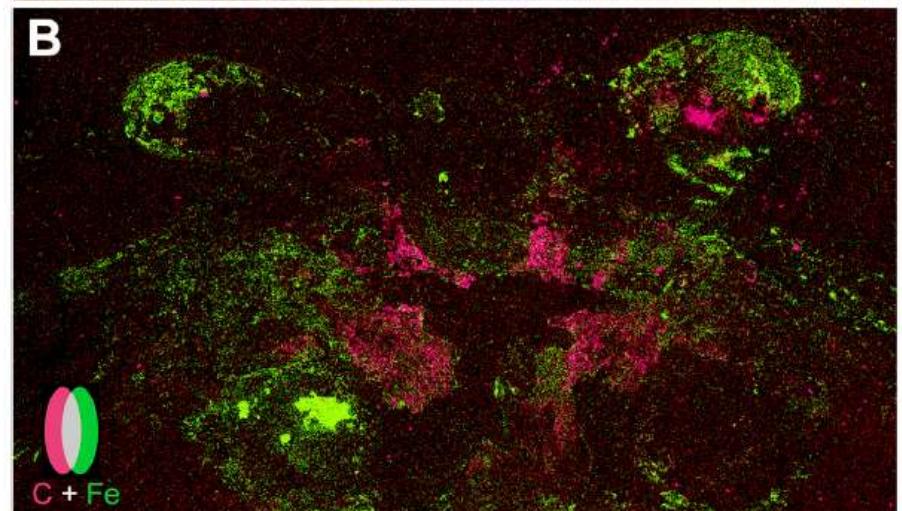


K: interpretive drawing of a Cambrian arthropod fossil

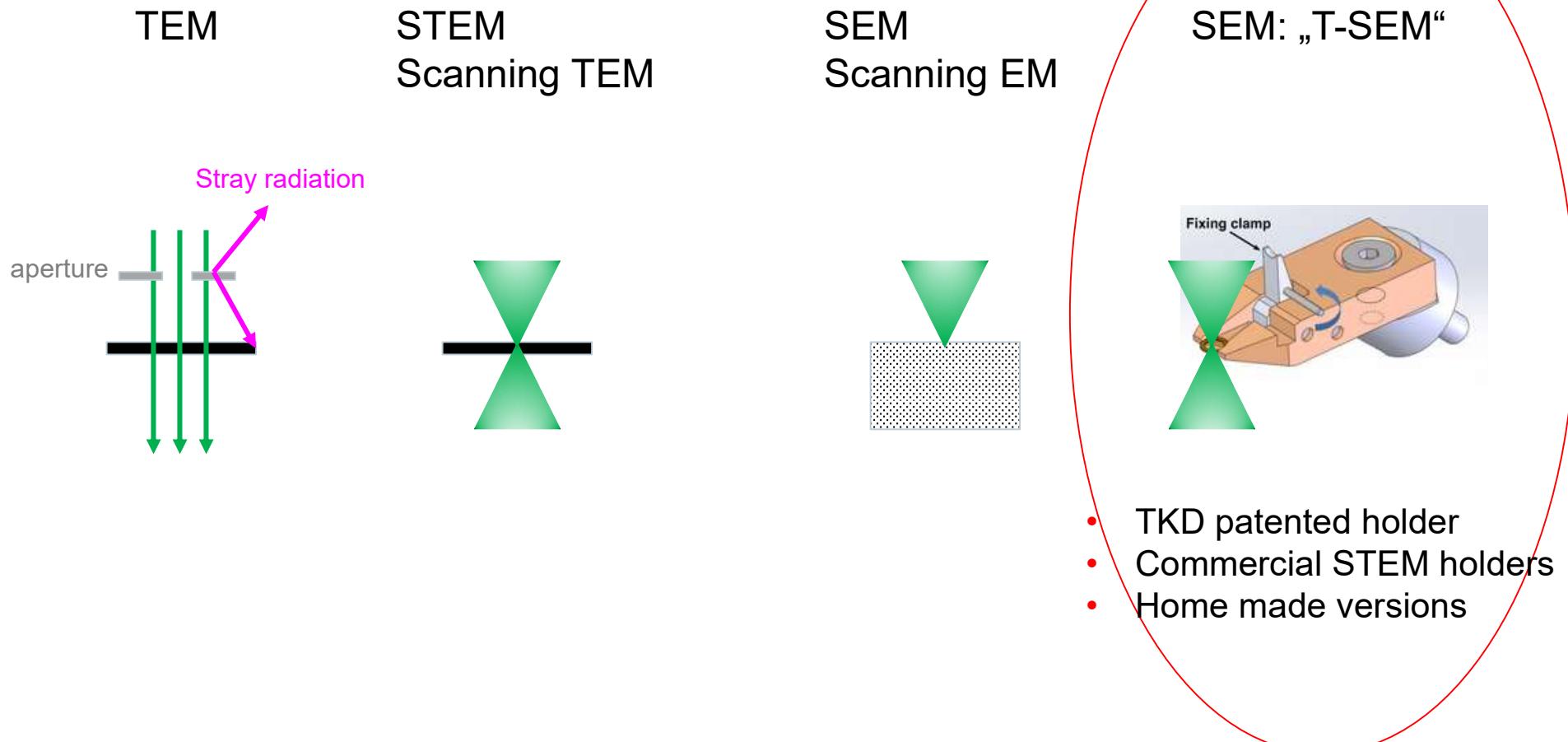
A: Direct illumination, C-traces of preserved neural tissue are shown as black

B: Merged C and Fe EDX maps resolve carbon and iron as entirely non-overlapping

- VP-mode: 30Pa, 10kV
- Stiching 63 fields of view
- each frame 15min, 923nm/p,
- > 500kcps OCR

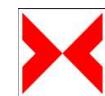
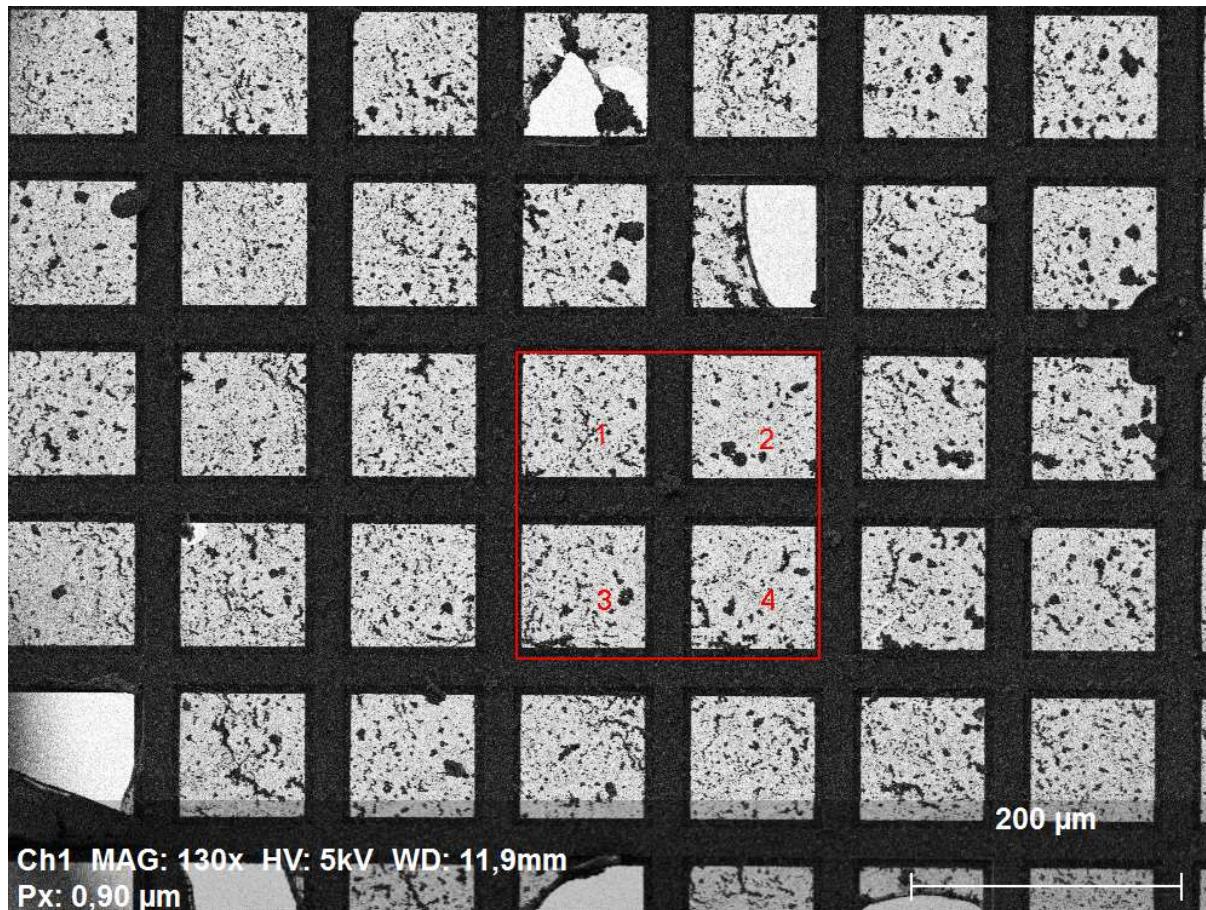


# Electron Microscopy



# T-SEM-EDS of NP

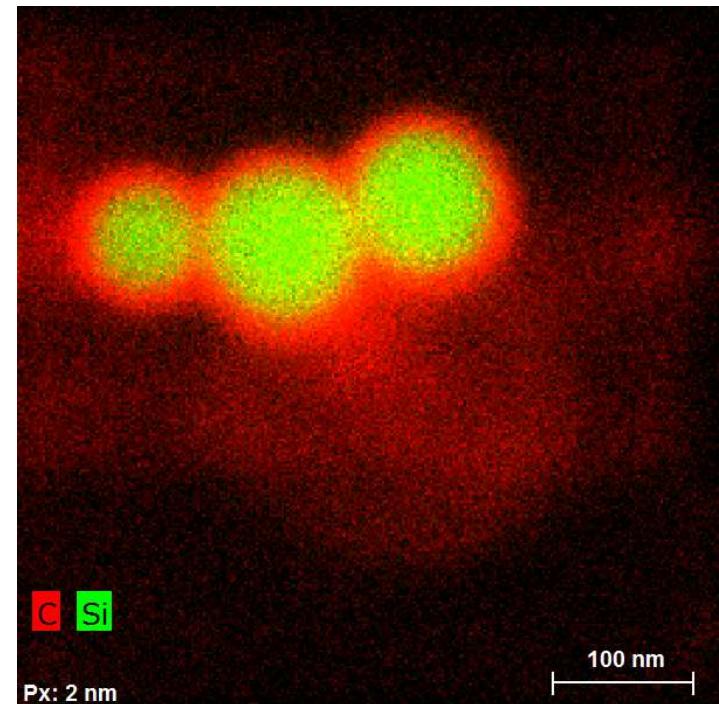
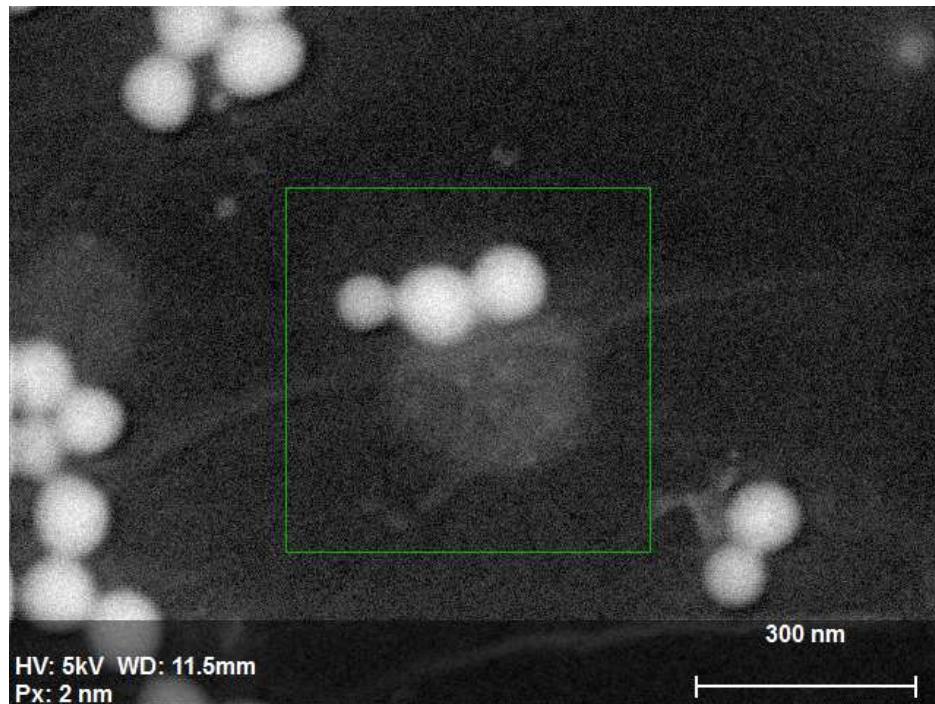
## Typical Overview



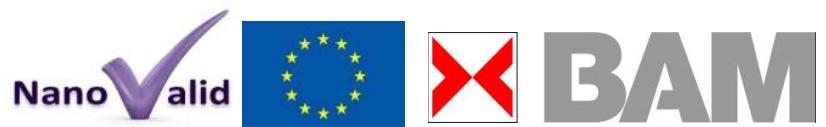
# T-SEM-EDX of fluorescent core shell NP; Silica nanoparticles Alexa<sup>®</sup> dye coated



XFlash FlatQUAD, 5 kV, 520 pA , 22.5 kcps, 250x250 pixel, 2 nm pixel size, 377 s



K. Natte, T. Behnke, G. Orts-Gil, C. Würth, J. F. Friedrich, W. Österle and U. Resch-Genger, J Nanopart Res, 2012, 14, 680

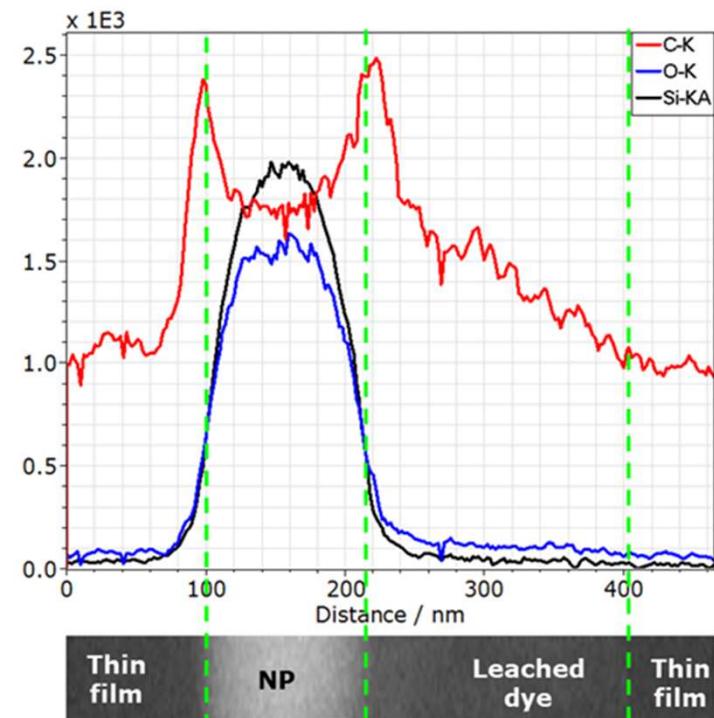
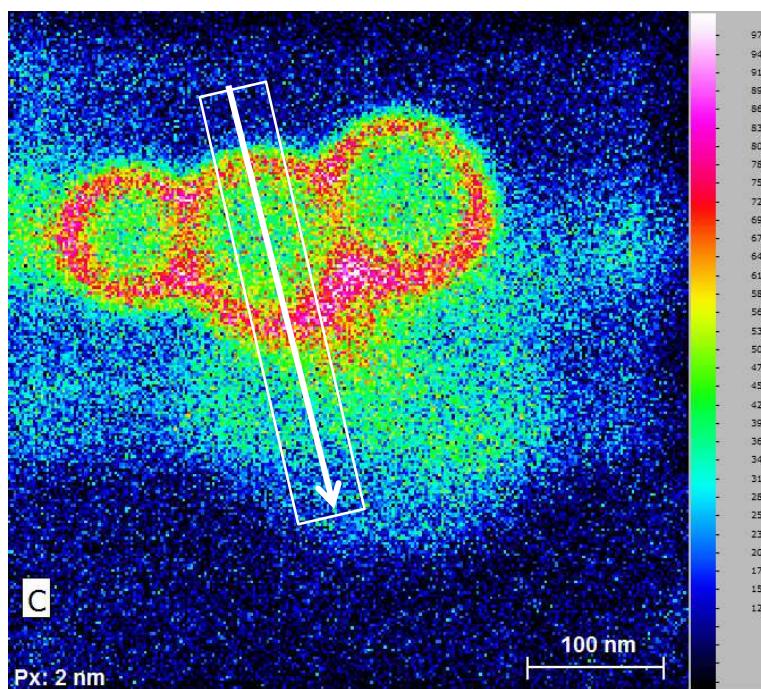


Analysis: T. Salge (Bruker/NHM)

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XFlash FlatQUAD, 5 kV, 520 pA , 22.5 kcps, 250x250 pixel, 2 nm pixel size, 377 s

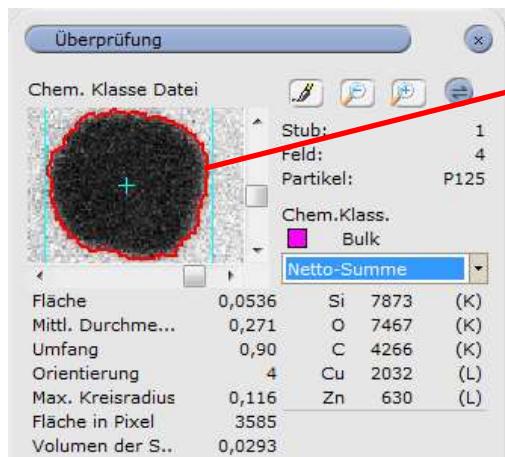


K. Natte, T. Behnke, G. Orts-Gil, C. Würth, J. F. Friedrich, W. Österle and U. Resch-Genger,  
J Nanopart Res, 2012, 14, 680;

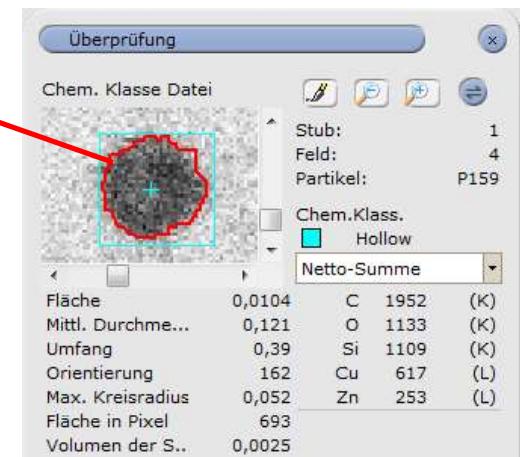
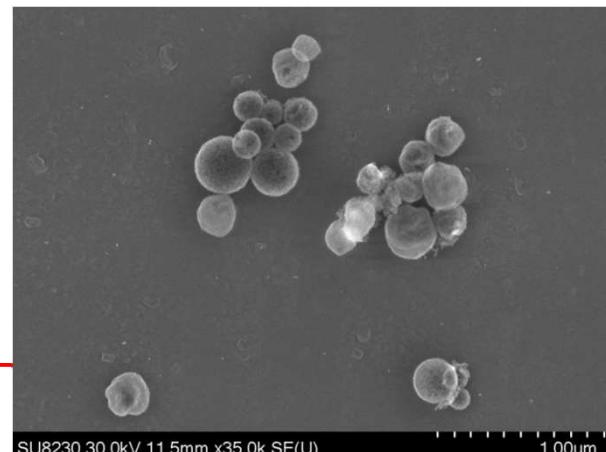
Analysis: T. Salge, M. Falke; Hitachi SEM

# T-SEM-EDX of NP

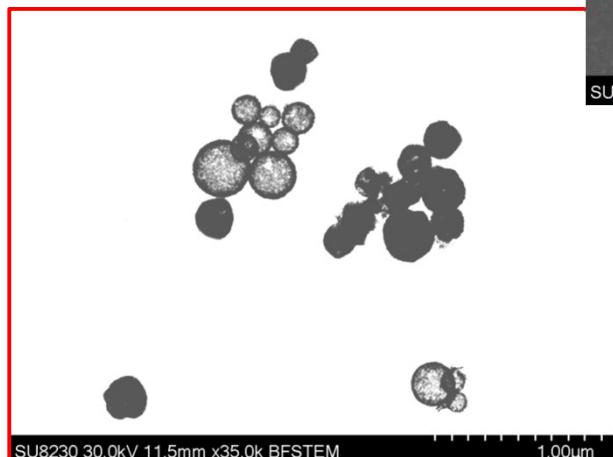
Classification using Si/C ratio



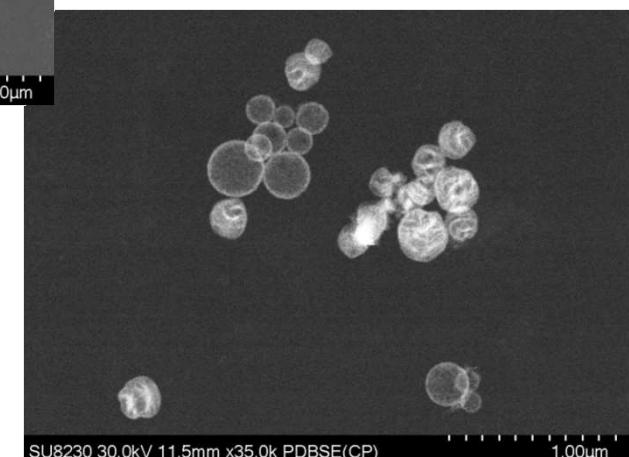
„bulk“ NP



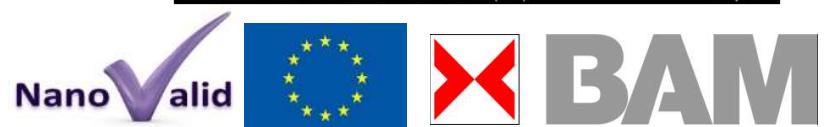
„hollow“ NP



SU8230 30.0kV 11.5mm x35.0k SE(U) 1.00µm

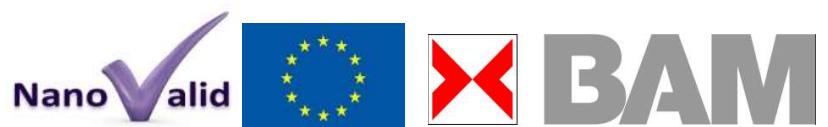
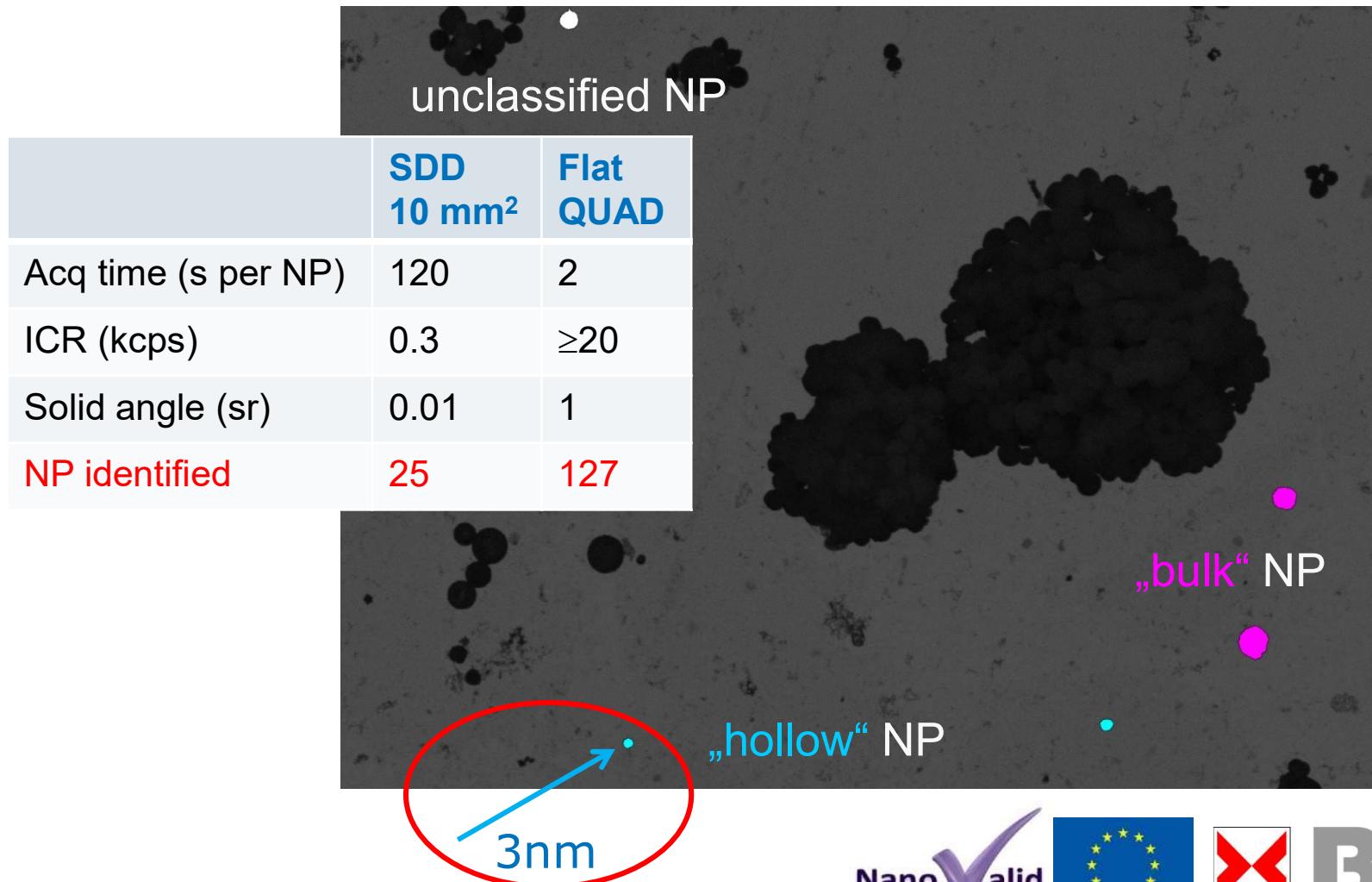


SU8230 30.0kV 11.5mm x35.0k PDBSE(CP) 1.00µm



# T-SEM-EDX of SiO<sub>2</sub> NP;

## PA: Classification, Statistics



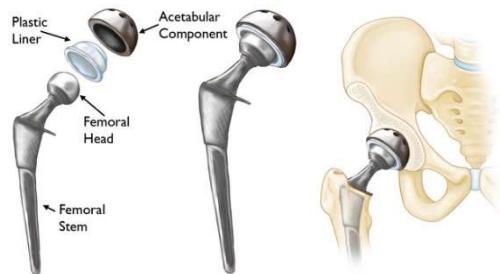
# SEM-EDX of Au-NP on TiO<sub>2</sub>

## Sponge-like coating for implants

### Overview > Particle Statistics!



#### Nanotoxicity

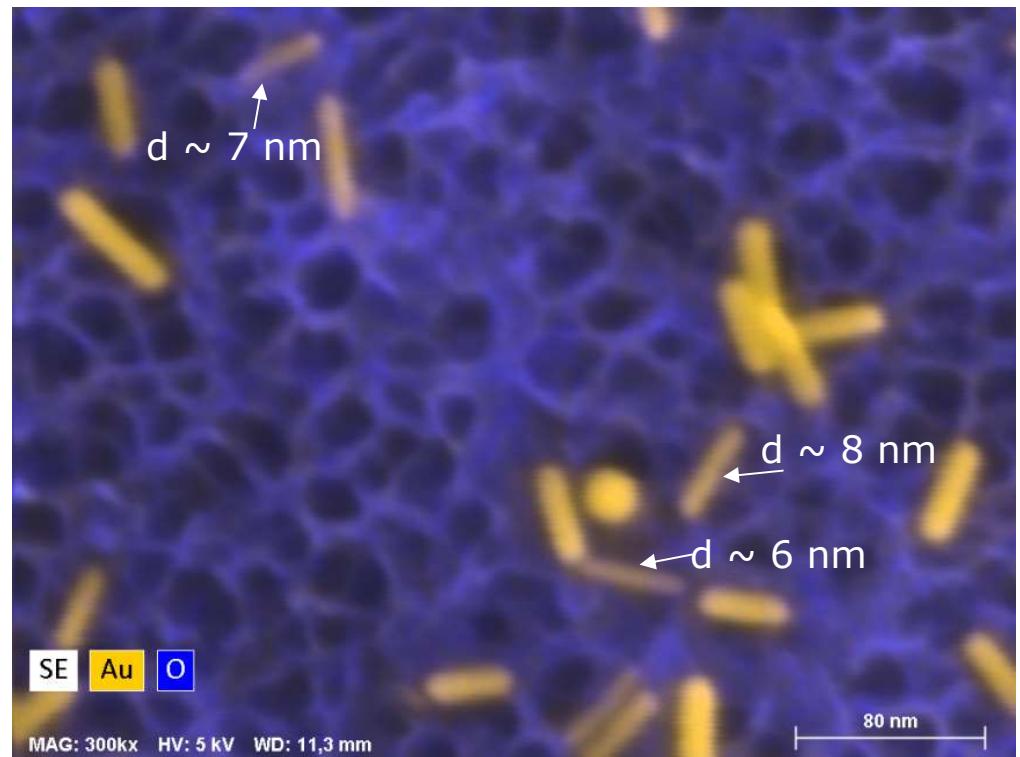


Au-NP to avoid inflammation around implant:  
NP change surface potential,  
Settling bacteria get „electrocuted“.

For successful tissue growth it is very important to judge the distribution of the NP and compare it to fluorescence light microscopy.

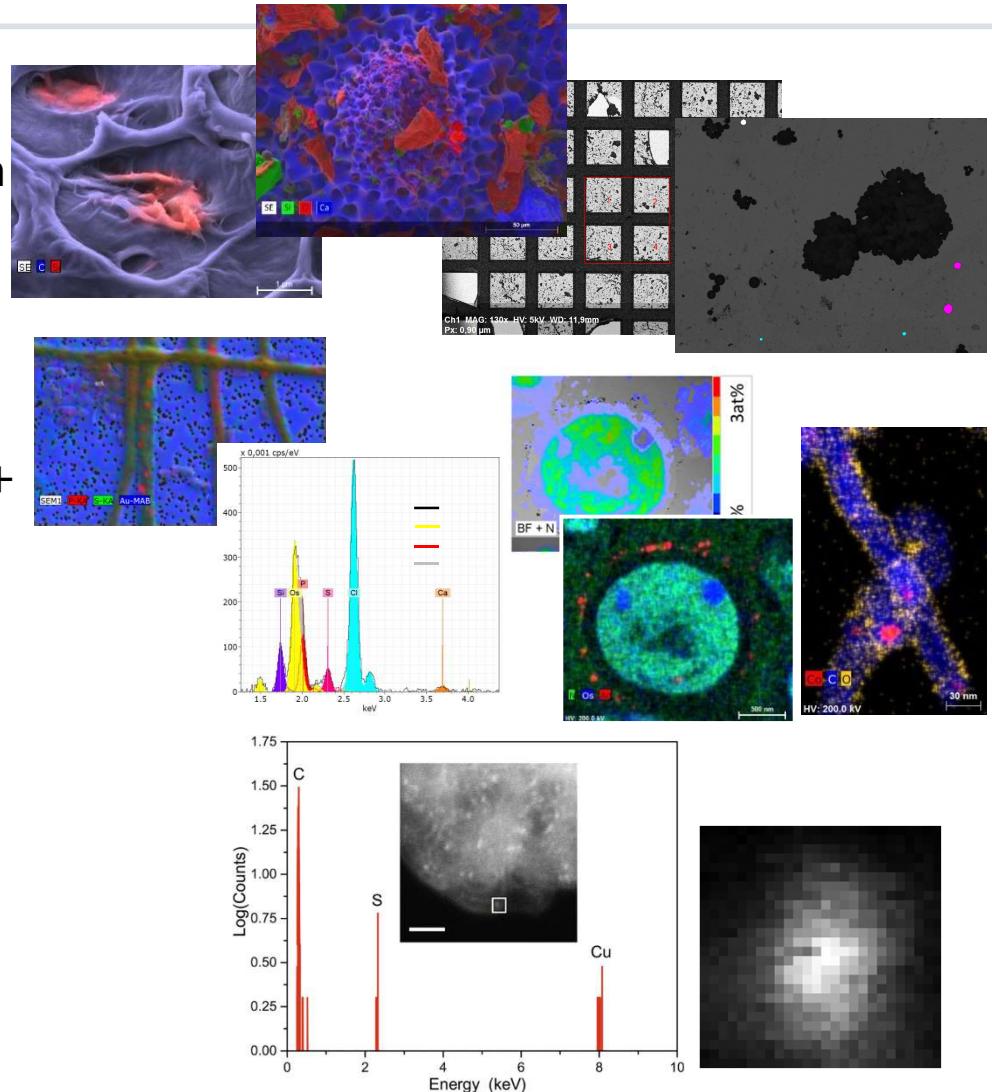
#### > Statistical analysis!

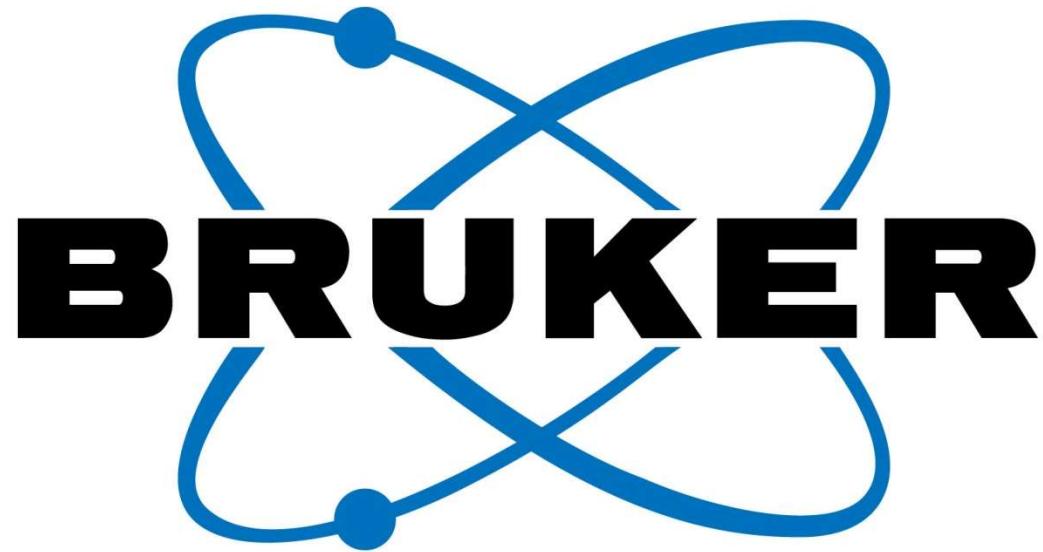
T. Yang et al., Colloids and Surfaces B: **145**, 597 (2016).



# EDS Characterization; Possible Steps: From mm via nm to single atoms

- SEM/T-SEM
  - > Overview / embedding/ statistics from mm to nm scale, Low vacuum,
  - > Use multiple/annular detectors
  - > Combine with other analysis techniques on SEM (EBSD/TKD,  $\mu$ XRF)
  
- Standard / Cs-corrected SEM/STEM + Standard EDS
  - Q-Mapping in at% at nm spatial resolution, in-situ, liquids, ice ... force
  
- Cs-corr. STEM + high brightness + high  $\Omega$  EDS
  - > Single atoms
  - > Combine with EELS, CL, diffr. ...
  - > in situ (liquids, ice, gases, temp., force)





Innovation with Integrity