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microGRID Technology For Robust High-resolution **GRID BRUKER** Imaging Down To The (Sub)cellular Level

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Introduction

MALDI imaging is a powerful technique to map biomolecules in tissue. To create a spatially resolved ion image, most MALDI instruments move the sample using an x-y stage relative to a stationary laser beam to create a mass spectrum for every pixel. However, approaching spatial resolution of only a few micrometers, over wide travel range, poses a challenge for the mechanical accuracy of most stages, affecting the quality of the MALDIimages. Here, we introduce *microGRID*, a new instrument design which combines both stage and laser beam positioning to eliminate imaging artifacts down to about 5 µm. In combination with MALDI-2, this new technique enables highly sensitive imaging at high spatial resolution without compromising on pixel fidelity.

Results

- The *microGRID* laser beam adjustment leads to the reduction of oversampling artifacts like checkerboard patterns and striping and thus increases the validity of high-resolution imaging data.
- The increase of achievable and robust lateral resolution in MALDI-MSI now enables one to clearly



(B) beam correction <u>ON</u> (A) beam correction <u>OFF</u>

visualize small structures within the testicle, which are not

distinguishable at bigger pixel sizes (Fig. 2). This emphasized the need for robust tools to perform MALDI-MSI with \leq 5 µm spatial resolution.

- The combination of laser positioning robustness and fast scanning rates of 10 pixel/sec allows for imaging of large sample areas like mouse brain or rat kidney while maintaining significant spatial information (Fig.3).
- *microGRID* imaging can resolve target structures, such as the Purkinje cell layer in the cerebellum (a) and the glomerular system of

Fig. 2 MALDI-2-MSI data of a rat testis, imaged with 5, 10 and 20 µm pitch of the MALDI sample stage movement with laser beam correction by *microGRID* technology.





Fig. 1 Combination of sample stage movement and laser beam positioning. With (A) beam correction off, limitations in stage precision lead to artifacts in pixel positioning. (B) *microGRID* exploits the laser beam steering to correct to nominal pixel position, enabling artifact-free imaging.

Methods

Optical encoders with sub-micron resolution were integrated into a regular stepper driven MALDI sample stage and monitor the actual position. Any deviation from the ideal raster is precisely detected by the encoders and sent to the adaptive smartbeam[™] 3D laser optics for automatically on-the-fly correction and irradiating precisely the targeted pixel within µm accuracy.

the kidney (b). This makes a new histological depth accessible for MALDI-MSI.

- MALDI-2boosts sensitivity to compensate for decreased ablation material at smaller pixel sizes and online calibration guarantees stable MS confidence for elongated run times.
- *m/z* 810.682 m/z 834.5996 *m/z* 331.265 *m/z* 346.057 *m/z* 792.554 m/z 465.305 PE(40:6) HexCer(d42:2) PC(40:6) Adenosine-5'-monophosphate Adrenic acid Cholesterol sulfate

Fig. 3 5 µm microGRID imaging data of (A) lipids in mouse cerebellum measured with MALDI-2 (pos), and (B) metabolites in rat kidney measured with MALDI (neg). The data demonstrates the robustness of the microGRID technology even with enlarged sample areas.

Conclusion

- microGRID allows for true high-spatial resolution MALDI-MSI down to 5 µm
- Stage movement is compensated by a highresolution positional readout and correction by the laser beam accordingly via
- The significant increase in positional accuracy leads to improved spatial resolution and thereby opens the field for the analysis of the smallest unit of life - single eucaryotic cells. With dedicated sample preparation strategies like matrix-sublimation, *microGRID* is able to push imaging resolution to its limit and allows for the visualization of cellular fine structures and organelles like the nuclei (Fig. 4).

m/z 760.584

Images were collected using smartbeamTM 3D systems with ~5 μ m laser spot size at raster spacing of 5-20 µm. We used three different kinds of samples with CHCA and DHAP matrixcoating by sublimation. MALDI-MSI data were visualized using SCiLS Lab. Tentative annotations were obtained by MetaboScape based on accurate mass.



Fig. 5 MALDI-2-MSI data imaged at 5 µm resolution using *microGRID* of (A) Caki-2 cells, overlayed with fluorescence microscopy data showing subcellular resolution (zoom-in in B). Cells were kindly prepared by Jan Schwenzfeier, University of Münster.

smartbeamTM 3D technology Sensitivity enhancement with MALDI-2 allows for visualizing of numerous molecules in various tissue types and even on the single (sub)cellular level

Technology

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