For more than 30 years, molecules have been analyzed in the solid, liquid and gaseous state on the basis of their specific vibrations. So, the (FT-IR) spectrosocopy has become an indispensable tool in the chemical and pharmaceutical industry for characterizing materials and identifying substances. In the last years, however, a completely new application field in the area of Life Science has opened up to this non-invasive analysis technique. Pharmaceutical and biotechnological companies use this technique to analyze both proteins and cellular systems in order to develop new drugs and products.

Infrared light excites molecules to vibrate. The molecules absorb only light of certains wavelengths that correspond to their specific vibration frequencies. Therefore, the position of the corresponding band in the absorption spectrum is characteristic for a certain vibration and, according to the Lambert-Beer law, the band intensity is directly proportional to the concentration of the vibrating molecule. Today’s Fourier-transform infrared (FT-IR) spectrometer cover the whole (mid) infrared region and allow the acquisition of high-quality spectra within seconds.

**Proteins**
Proteins become more and more important as active agents in medical drugs. Especially therapeutic antibodies are a promising approach for the treatment of diseases that have been incurable so far. Under what conditions remains the protein stable? How long lasts this stability? These and other questions have to be answered in the course of the formulation optimization. The classical method to detect the aggregates of the protein is the size-exclusion chromatography. The denaturation process observed with this method mostly starts mechanistically at an earlier stage, namely with conformational changes of the protein and often under the formation of a typical β-sheet structure. Using the FT-IR spectroscopy, conformational changes can be detected with a very high sensitivity. So, this technique allows to recognize instable formulations already at an early stage. Furthermore, the aggregation process can be monitored directly using the FT-IR technique. To ensure a stable formulation, the proteins are often kept in complex buffers with a large number of additives like sugars, polyalcohols or amino acids and they are stored either in dissolved state (liquid formulation) or in powder-form (solid formulation) after the lyophilization. The FT-IR spectroscopy now gives the opportunity to measure the structural changes of proteins both in liquid and solid formulations easily and quickly regardless of the buffers or additives.
Figure 1: Tissue section (1.6 x 0.8mm) of a mouse kidney (thickness: 10µm). In the visible video image, the active agent is discernible by the yellowish staining. The whole sample area has been measured using the FT-IR imaging technique. For each image point (4 x 4µm) a full IR spectrum was received. The intensity of the individual absorption bands is proportional to the concentration of corresponding absorbing molecule class. The distribution of the individual molecule classes has been visualized by a false color image reflecting the signal intensities of certain absorption bands. The figures show the distribution of the active agent, the proteins and the lipids. It is clearly visible that the active agent is mainly located in the protein-rich region, whereas, the lipids are concentrated in distinctly separated areas. The measurement has been performed using a Bruker Hyperion 3000 microscope.

Analysis of cells and search for natural substances

Already in the 1980s, Dieter Naumann and Harald Labischinski from the Robert-Koch Institut in Berlin have developed a method for identifying microorganisms that bases on the FT-IR spectroscopy. The most amazing aspect of this method is that many microorganisms can be identified even to strain level! Today, the pharmaceutical industry uses this technique increasingly to find new natural substances in the microorganisms. To do this, either the intact cells or chemical extracts from these cells are measured. Using multivariate analysis techniques (PCA, ANN), the acquired infrared spectra are compared with each other and examined for characteristic distinctions that point to the specific effect of additional substances (natural substances). For these kinds of applications, modern FT-IR microtiter plate readers allow a sample throughput of up to 10,000 samples per day.

Bioprocesses

In the optimization phase of bioprocesses, several fermentation parameters (e.g. sugar, temperature, pH value, nutrient substances) are often changed systematically in a parallel approach. For the detection of the effects of the individual parameters, it is important to measure not only the product concentration but also these parameters regularly during the fermentation process. To do this, the FT-IR spectroscopy allows a simultaneous measurement of the recombinant
product and many other parameters directly in the fermentation broth without requiring a sample clean-up. The measurement takes only a few seconds and requires no consumables. In this way, several chromatographic methods can often be substituted for the fast infrared measurement method.

**Imaging of tissues**

Only for a few years, modern FT-IR microscopes have been equipped with multielement detectors. This technique enables the analysis of tissue sections with a spatial resolution of up to 0.5 µm within a few minutes. During the measurement, a complete infrared spectrum is acquired at each image point. The result is an image showing the distribution of the main biochemical components (especially lipids, proteins and polysaccharides) in the analyzed tissue area (see figure 1). This measurement technique requires neither chemical labeling of the components nor staining of the sample. To visualize also the distribution of the individual proteins or active agents in the same sample, today’s imaging infrared microscopes are equipped with fluorescence channels. Typically, unstained cryomicrotome sections with a thickness of 6 to 10µm are analyzed. The analytical questions that can be answered with this technique range from characterizing cancer tissues and confirming the presence of implant material to detecting amyloid structures.