A combination of single-cell MALDI-MS imaging and fluorescence microscopy to explore molecular heterogeneity in cell cultures

Motivation

- MS-analysis of extracts from pooled cells provide a molecular profile but is blind to general heterogeneities within a culture
- Single-Cell heterogeneity is an inherent property of every cell population
- * Its analysis is fundamental to understanding the development, function, and role of specific cells that share the same genotype but may display different phenotypical properties

Experimental Set-up

Sample Preparation & Data Aquisition:

- Cell Culture: cells (Vero-B4 and Caki-2) were grown on 8-well chamber slides; fixed in 4% formaldehyde for 5 mins; stained with Hoechst and WGA; washed; sublimated with matrix (2,5-DHAP).
- ✤ Microscopy: digital slide scanner (SLIDEVIEW) VS200, Olympus); 20x objective; bright-field and fluorescence.
- MALDI-2-MSI: timsTOF fleX (Bruker) with 8 μm pixel size and Orbitrap Q Exactive Plus (Thermo Fisher Scientific) with $2 \mu m$ pixel size.

Data Processing:

- SCiLS Lab (Bruker): spectra reduced to signals correlating to cell regions using the co-localization function. Python: Single-Cell Segmentation (otsuthreshold and watershed) on the three microscopy channels.
- **Co-registration:** MALDI-MSI binarized and microcopy data were overlaid using 2-D- and rotationcorrelation.
- ✤ Machine learning: Single-Cell mass spectra were normalized to the TIC. A support-vector machine was trained on mono-cultures (5-fold cross-validation) and then used to classify co-cultures.

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