# Intact Protein MALDI Mass Spectrometry Imaging (MSI) to characterize MP target engagement and blood-brain barrier penetration for amyotrophic 041 lateral scierosis (ALS)

### **OVERVIEW**

- Target engagement and exposure at the site of action are critical to link protein modification to pharmacodynamics.
- Covalent drugs bind irreversibly to target proteins, which gives an opportunity to use intact protein mass spectrometry methods to detect the covalent drug-protein complex. • Fluoroscein-5-maleimide (F5M) was used as a chemical probe for proof-of-concept in ALS
- mouse brain homogenate using intact protein MALDI mass spectrometry. • Our in vivo investigation involved direct detection of drug-protein complex in situ. The drug
- candidate was S-XL6. This prototype cyclic thiosulfinate was dosed into ALS mice and intact protein MALDI MSI was used to visualize the interaction of S-XL6 with its target, copper zinc superoxide dismutase 1 (SOD1). This workflow informs target engagement and bloodbrain barrier (BBB) penetration and unlike conventional bioanalysis, also preserves the spatial component.
- Direct detection of drug-protein complexes can enable on and off-target identification in drug development and coupling to MSI enables visualization at the site of action.

### INTRODUCTION

- ALS is a fatal, neurodegenerative disease characterized by death of upper and lower motor neurons in brain and spinal cord. Drugs must therefore cross the BBB.
- Mutations in SOD1 protein are linked to familial ALS (fALS), associated with loss of quaternary structure, protein destabilization and aggregation.
- Dimer stabilization can provide as a potential therapeutic strategy for ALS.



Figure 1. Cyclic thiosulfinate cross-link SOD1 at Cys111 to form a dimer in vitro.<sup>1</sup>

### METHODS



Figure 2. Schematic diagram of MALDI workflow for detection of intact proteins. (A) MALDI MSI workflow for detection of intact proteins from ALS mouse brain sections. (B) Workflow for detection of intact proteins from mouse brain homogenate.

Rutali R. Brahme<sup>1\*</sup>, Md Amin Hossain<sup>1</sup>, Catherine M. Rawlins<sup>1</sup>, Madison H. McMinn<sup>1</sup>, Novera Alam<sup>1</sup>, Nathalie Y. R. Agar<sup>2</sup>, Jeffrey N. Agar<sup>1</sup> <sup>1</sup>Northeastern University, Boston, MA; <sup>2</sup>Brigham and Women's Hospital, Boston, MA.

## RESULTS



Figure 3. MALDI FTICR MS of intact proteins. (a) Purified fALS variant SOD1<sup>G93A</sup> modified by chemical probe F5M. (b) Control vs. sample ALS mouse brain homogenate spiked with F5M, showing target (SOD1<sup>G93A</sup>) engagement in vitro. (c) Off-target analysis. To develop methods three different sample preparations were tested and probe-modified proteins were detected in ALS mouse brain homogenate. An example of off-target was 9772 m/z, putatively identified as High affinity cationic amino acid transporter 1.



**Figure 4.** MALDI TOF MS of intact proteins *in vitro* (spiked mouse brain homogenates). The average spectrum of the entire TOF MS data set is shown in (a). Expanding the intensity scale (b) highlights the target engagement of SOD1<sup>G93A</sup> using chemical probe F5M *in vitro*.



Figure 5. MALDI TOF MS of intact proteins in vivo (mouse brain homogenates from S-XL6 dosed animals). The average spectrum of the entire TOF MS data set is shown in (a). Expanding the intensity scale (b) highlights the detection over high m/z range. Arrows indicate (1) Ubiquitin signal (m/z 8540), (2) SOD1<sup>G93A</sup> monomer (m/z 15858), (3) eYFP (m/z 26774), (4) Cross-linked SOD1<sup>G93A</sup> dimer (m/z 31836). Note that we are using transgenic G93A mice bred with mice expressing eYFP in specific cells, so that motor neurons can be identified in MALDI MSI.<sup>2</sup>



- support.
- This work was supported by the ALSA grant 18-IIA-420.





• We would like to thank Bruker Scientific, Billerica, MA for letting us use their instruments and for