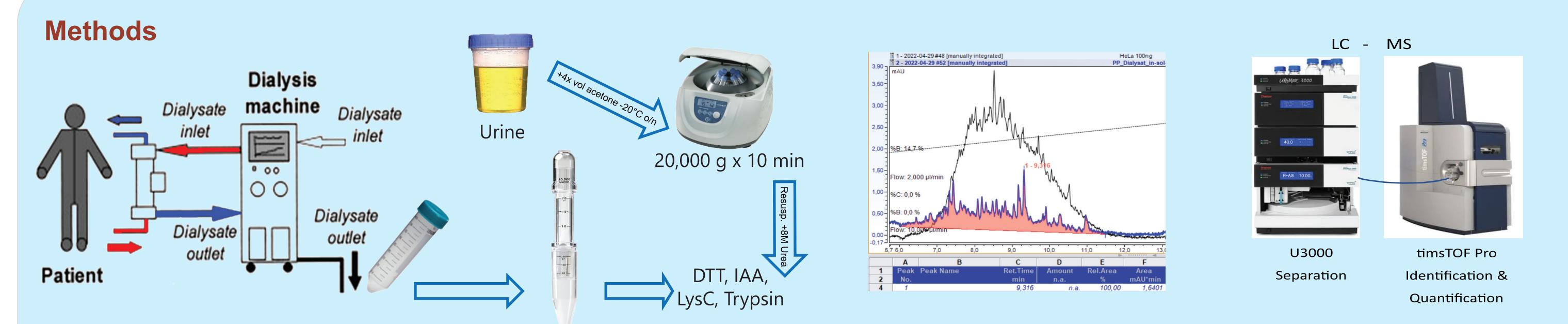
A quantitative proteomics workflow for urine and spent dialysate using datadependent and data-independent analyses on a timsTOF Pro instrument

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Introduction

Despite hemodialysis being immediately life saving for patients with kidney failure, the prognosis of patients on hemodialysis is generally poor with median survival rates of approximately five years. Interestingly, mortality of hemodialysis patients who retain RKF (residual kidney function) is lower.^{1,2)} While hemodialysis effectively removes small molecules, the clearance of medium-size molecules appears less efficient. It was hypothesized that toxic medium-size proteins, which can still be excreted into urine by hemodialysis patients with RKF, might accumulate in patients without RKF.³⁾ Alternatively, proteins that are normally reabsorbed in the kidney tubulus and that help maintain stable homeostasis, could be lost in hemodialysis. To evaluate these hypotheses, we established a high-throughput quantitative proteomics workflow for the



Dialysate flow was 500-800 mL/min i.e. 100-200 L during 3-4 hr dialysis session. Spent dialysate was collected by slow aspiration with a syringe from an adapter mounted on the dialysate outlet hose.

400 µL residual urine +4x vol acetone was precipitated o/n and centrifuged @20.000g for 10 min, followed by resuspension in 8M Urea. 10 mL of spent dialysate +8M Urea was concentrated to 200µL by repeated ultrafiltration using 2mL spin columns with 10kD cut-off

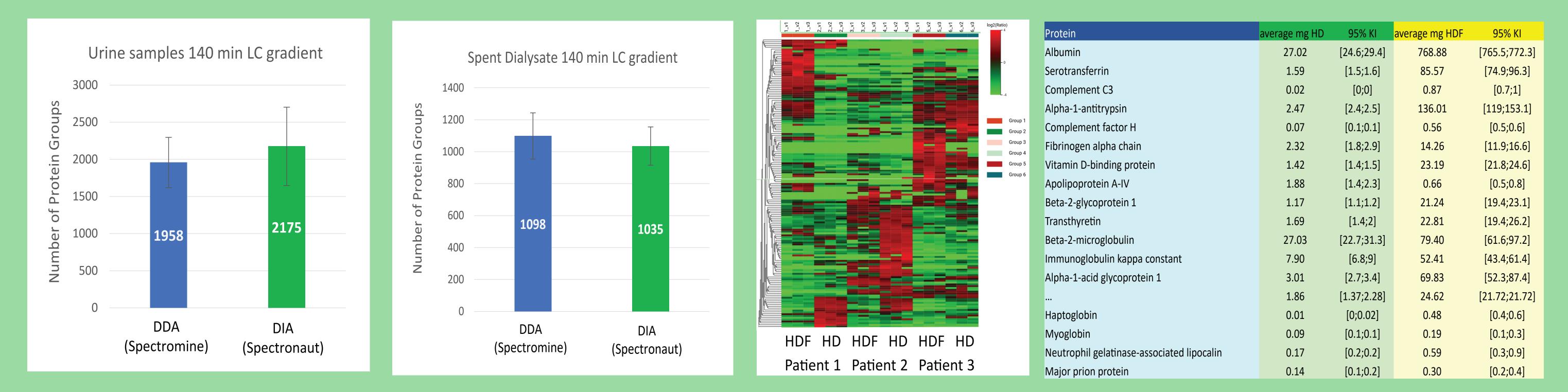
AUC @ 214 nm compared to 100ng centration of the original urine and spent dialysate samples.

Digested samples were quantified by Samples were separated on a U3000 system with PepMap 50cm or Pharmafluidics 110cm HeLa on an monolith LC. This permit- columns (140 min gradient time). Spectra were ted an estimation of the protein con- acquired on a timsTOF Pro instrument using DDA and DIA. Spectromine 3.2 and Spectronaut 16 were used for data analysis.

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Results



On average 1958 protein groups could be identified in triplicates of four urine samples using DDA (data-dependent analysis), as compared to 2175 protein groups using DIA (data-independent analysis). Data completeness was superior with DIA. This indicates that a level of sensitivity that required two-dimensional separation of samples only a few years ago, can now be achieved in 1D LC-MS runs.

With regard to spent dialysate, on average 1035 protein groups could be identified with DDA in triplicates of six dialysate samples from three patients using two dialysis modes (hemodialysis HD and hemodiafiltration HDF), as compared to 1098 groups with DIA. The protein list comprised several known uremic toxins. Results showed significant differences in the concentrations of proteins, depending on the type of dialyzer membrane and whether HD or HDF was performed. This suggests a possibility to tailor hemodialysis to individual patients' needs.

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